

## VIII. APPENDIX

### A. Solutions and buffers for DNA extraction

#### 1. Red cells lysis buffer

|                         |      |    |
|-------------------------|------|----|
| 1.0 M Tris (pH 7.65)    | 2.5  | ml |
| 1.0 M MgCl <sub>2</sub> | 1.25 | ml |
| 5.0 M NaCl              | 0.5  | ml |
| distilled water to      | 250  | ml |

Sterilize by autoclaving and store at 4°C.

#### 2. Nucleic lysis buffer

|                     |      |    |
|---------------------|------|----|
| 1.0 M Tris (pH 8.0) | 12.5 | ml |
| 0.5 M EDTA (pH 8.0) | 0.5  | ml |
| 5.0 M NaCl          | 5.0  | ml |
| distilled water to  | 250  | ml |

Sterilize by autoclaving and store at 4°C.

#### 3. Pronase K solution

|                     |     |    |
|---------------------|-----|----|
| 0.5 M EDTA (pH 8.0) | 40  | μl |
| 10 % SDS            | 1.0 | ml |
| proteinase K        | 20  | mg |
| distilled water to  | 10  | ml |

Dispense into aliquots and store at -20°C.

## 4. Acid citrate dextrose (ACD)

|                    |      |    |
|--------------------|------|----|
| citric acid        | 0.48 | g  |
| sodium citrate     | 1.32 | g  |
| glucose            | 1.47 | g  |
| distilled water to | 100  | ml |

Sterilize by autoclaving and store at 4°C.

**B. Solutions and buffers for plasmid preparation**

## 1. Sucrose buffer

|                     |     |    |
|---------------------|-----|----|
| sucrose             | 30  | g  |
| 0.5 M EDTA (pH 8.0) | 20  | ml |
| 1.0 M Tris (pH 8.0) | 10  | ml |
| distilled water to  | 200 | ml |

sterilize by autoclaving and store at 4°C.

## 2. Triton X-100 buffer

|                     |     |    |
|---------------------|-----|----|
| triton X-100        | 0.2 | ml |
| 1.0 M Tris (pH 8.0) | 10  | ml |
| 0.5 M EDTA (pH 8.0) | 20  | ml |
| distilled water to  | 200 | ml |

sterilize by autoclaving and store at 4°C.

## 3. 30% PEG, 1.5 M NaCl

|                     |       |    |
|---------------------|-------|----|
| polyethylene glycol | 60    | g  |
| NaCl                | 17.54 | g  |
| distilled water to  | 200   | ml |

Sterilize by autoclaving and store at 4°C.

**C. Reaction buffers for restriction endonuclease digestion.**1. Reaction buffer supplied for use with *Pst* I and *Hin* d III

final concentration:

50 mM Tris-HCl (pH 8.0)

10 mM MgCl<sub>2</sub>

50 mM NaCl

2. Reaction buffer supplied for use with *Eco* R I and *Bgl* II

final concentration:

50 mM Tris-HCl (pH 8.0)

10 mM MgCl<sub>2</sub>

100 mM NaCl

**D. Solutions and buffers for agarose gel electrophoresis**

## 1. 10x Tris-borate (TBE) buffer

|                     |      |    |
|---------------------|------|----|
| Tris base           | 54   | g  |
| boric acid          | 27.5 | g  |
| 0.5 M EDTA (pH 8.0) | 20   | ml |

distilled water to 1000 ml

Mix thoroughly and store at room temperature.

2. Gel-loading buffer

bromophenol blue 0.25 %

sucrose in water 40 %(W/V)

Mix thoroughly and store at 4°C.

3. Stock ethidium bromide(10 mg/ml)

ethidium bromide 100 mg

distilled water 10 ml

Mix thoroughly until dissolve. Store at room temperature in a dark bottle.

**E. Solutions for Southern blotting**

1. 0.25 M HCl

Slowly add 21 ml concentrated HCl solution to 979 ml deionized water while stirring. Store at room temperature.

2. 1.5 M NaCl, 0.5 M NaOH

NaCl 87.7 g

NaOH 20 g

deionized water to 1000 ml

Mix thoroughly and store at 4°C.

## 3. 1.5 M NaCl, 1.0 M Tris (pH 7.5)

|                 |       |    |
|-----------------|-------|----|
| NaCl            | 87.7  | g  |
| Tris base       | 121.1 | g  |
| deionized water | 800   | ml |

Adjust the pH to 7.5 with concentrated HCL. Adjust the volume to 1000 ml with deionized water. Mix thoroughly and store at 4 ° C.

## 4. 20x SSC

|                           |       |    |
|---------------------------|-------|----|
| NaCl                      | 175.3 | g  |
| Sodium citrate, dihydrate | 88.2  | g  |
| deionized water           | 800   | ml |

Adjust the pH to 7.0 with 4 N NaOH. Adjust the volume to 1000 ml with deionized water. Sterilize by autoclaving and store at 4°C.

### **F. Component of BioNick<sup>TM</sup> Labelling System**

## 1. 10x dNTP Mix

0.2 mM each dCTP, dGTP, dTTP

0.1 mM dATP

0.1 mM biotin-14-dATP

500 mM Tris-HCl (pH 7.8)

50 mM MgCl<sub>2</sub>

100 mM β-mercaptoethanol

100 µg/ml nuclease-free BSA

2. 10x Enzyme Mix

0.5 units/µl DNA Polymerase I

0.0075 units/µl DNase I

50 mM Tris-HCl (pH 7.5)

5 mM magnesium acetate

1 mM β-mercaptoethanol

0.1 mM phenylmethylsulfonyl fluoride

50% (v/v) glycerol

100 µg/ml nuclease-free BSA

3. Control DNA

5 µg pBR 322 in

10 mM Tris-HCl (pH 8.0)

1 mM EDTA

4. Stop buffer

300 mM EDTA

**G. Solutions and buffers for hybridization**

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่  
Copyright© by Chiang Mai University  
All rights reserved

## 1. Prehybridization solution

| Component  | Final concentration | Amount |
|--|---------------------|--------|
| Formamide  | 50%(W/V)            | 50 ml  |
| NaCl   | 0.9 M               | 5.26 g |
| NaH <sub>2</sub> PO <sub>4</sub> .H <sub>2</sub> O | 0.06 M              | 0.83 g |
| Na <sub>2</sub> EDTA.2H <sub>2</sub> O             | 0.006 M             | 0.22 g |
| Ficoll   | 0.1%(W/V)           | 0.1 g  |
| Polyvinylpyrrolidone                               | 0.1%(W/V)           | 0.1 g  |
| Bovine serum albumin                               | 0.1%(W/V)           | 0.1 g  |
| Sodium dodecyl sulfate                             | 1.0%(W/V)           | 1.0 g  |
| Sheared,denatured salmon sperm DNA                 | 200 µg/ml           | 20 mg  |

Dissolve solid components except DNA in 40 ml deionized water. Adjust pH to 7.4 with 4 M NaOH. Add 2.0 ml of 10 mg/ml sheared, denatured salmon sperm DNA. Adjust volume to 50 ml with deionized water. Add 50 ml formamide. Store at -20°C.

## 2. 20% dextran sulfate

Dissolve 10 g dextran sulfate in formamide to give a final volume of 50 ml. High molecular weight dextran sulfate dissolves very slowly. Stir or rock the solution slowly overnight. Store at 4°C.

## 3. 2x Hybridization solution

| Component  | Final concentration | Amount |
|--|---------------------|--------|
| NaCl   | 1.8 M               | 5.26 g |
| NaH <sub>2</sub> PO <sub>4</sub> .H <sub>2</sub> O | 0.12 M              | 0.83 g |
| Na <sub>2</sub> EDTA.2H <sub>2</sub> O             | 0.012 M             | 0.22 g |
| Ficoll   | 0.2%(W/V)           | 0.1 g  |
| Polyvinylpyrrolidone                               | 0.2%(W/V)           | 0.1 g  |
| Bovine serum albumin                               | 0.2%(W/V)           | 0.1 g  |
| Sodium dodecyl sulfate                             | 2.0%(W/V)           | 1.0 g  |
| Sheared,denatured salmon sperm DNA                 | 400 µg/ml           | 20 mg  |

Dissolve solid components except DNA in 40 ml deionized water. Adjust pH to 7.4 with 4 M NaOH. Add 2.0 ml of 10 mg/ml sheared,denatured salmon sperm DNA. Adjust volume to 50 ml with deionized water. Store at -20°C.

## 4. TBS-Tween 20

| Component | Final concentration | Amount |
|-----------|---------------------|--------|
| Tris base | 100 mM              | 12.1 g |
| NaCl      | 150 mM              | 8.77 g |
| Tween 20  | 0.05%(W/V)          | 0.5 ml |



Adjust pH to 7.5 with 4 M HCl. Filter through a sterile 0.2- $\mu$  membrane. Store as a sterile solution at 4°C.

#### 5. Blocking solution

Bovine serum albumin            3    g

TBS-Tween 20                    100    ml

Adjust pH to 7.5 . Filter through a sterile 0.45- $\mu$  membrane.

Store at 4°C.

#### 6. Developer

Developer                            1    part

distilled water                    5    parts

Mix thoroughly and prepare before use.

#### 7. Fixer

distilled water                    3    parts

Fixative agent                    1    part

Mix thoroughly.

distilled water to                5    parts

Again, mix thoroughly and prepare before use.

**IX. BIOGRAPHY**

**NAME : Miss ROONGSIRI MUANGMOONCHAI**

**DATE OF BIRTH : AUGUST 3, 1968**

**PLACE OF BIRTH : RAYONG, THAILAND**

**INSTITUTION ATTENDED :**

**: PHADOONGPANYA SCHOOL, TAK...**

**MARCH 1985, CERTIFICATE OF MATHAYOM VI**

**: FACULTY OF ASSOCIATED MEDICAL SCIENCES,**

**CHIANG MAI UNIVERSITY, CHIANG MAI**

**MARCH 1990, BECHELOR OF SCIENCE IN MEDICAL**

**TECHNOLOGY, B.Sc.(MED.TECH.)**

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่  
Copyright© by Chiang Mai University  
All rights reserved