

## V. RESULTS

### 5.1. Isolation and characterization of the *hsp70* gene from *P. marneffei*

In a previous study by Pongpom (2004), a cDNA library was constructed in ZipLox vector from the yeast phase of *P. marneffei*. Three positive clones encoding Hsp70 (H1, H2 and H3) were obtained by immunoscreening of an expression library (20,000 pfu), using anti-*Histoplasma capsulatum* Hsp70 monoclonal antibody. An approximately 1.9 kb cDNA (H3) clone was sequenced and compared with the other *hsp70* gene. A 323 nucleotide sequence from the 5' region showed high similarity (>90%) to fungal *hsp70* gene including *Aspergillus nidulans*, *Trichophyton rubrum*, *Paracoccidioides brasiliensis* and *Histoplasma capsulatum*. Comparison with other *hsp70* genes revealed that the cDNA clone had an incomplete 5' end.

To isolate the full length *hsp70* cDNA clone, a 1.9 kb insert (clone H3), released by *NotI-SalI* digestion, was used as a probe to screen the other clone containing *hsp70* gene from the cDNA library (10,000 pfu) by using ECL Direct Nucleic Acid Labeling and Detection Kit (Amersham Pharmacia Biotech). Twenty-one positive clones from primary screening were screened for the longest insert size by PCR using T7 and H70-2 primers (Fig. 6). Secondary screening was performed on a putative full length *hsp70* cDNA, ph20, to isolate a pure phage containing *hsp70* cDNA, which was then sequenced and characterized. The cloned ph20 was 2,204 nucleotides in length and contained 96 bases at the 5' untranslated region (UTR). The cDNA had a single open reading frame (ORF). An open reading frame of *hsp70* composed of 1,911 basepairs was delimited by the well-known start (ATG) and stop (TAA) codons. The ATG codon at base 93 encoded the presumed initiating methionine. This amino acid was in the appropriate position of a consensus translation start (Kozak, 1986). The polypeptide of 636 amino acids had a calculated molecular mass of 69.5 kDa and a pI of 5.03. The cDNA included 201 bp in the 3'UTR, exclusive of the poly-A tail. The stop codon TAA was located at position 2256.

In order to find out the existence of any other intron in the *P. marneffei* *hsp70* gene, a genomic fragment corresponding to the entire region of ORF presented in the cDNA sequence was isolated by polymerase chain reaction (PCR) using PM-F and PM-R primers and sequenced (Fig. 7 and Table 3). Comparison of the nucleotide sequences from cDNA library and genomic DNA showed compatibility of both nucleotide sequences, except for two non-homologous regions from genomic DNA between nucleotides 838-894 and 898-1,095 (Fig. 7). Analysis of these regions strongly suggested that they are introns of 57 bp and 198 bp, respectively. Their 5' and 3' ends, shown in Figure 7, conformed to the basic consensus GT/AG for eukaryotic splice donor and acceptor sites (Mount, 1982; Breathnach and Chambon, 1981). Both regions contain an internal putative splice box which matches the filamentous fungus consensus sequence (NNCTPuPy) upstream from the 3'end of introns (Gurr and Unkles, 1987). Surprisingly, a microexon between two introns contains only 3 bases.

From Table 4, it can be seen that in fungal *hsp70* genes encoding Hsp70s from different cellular compartments, different numbers of introns are present, ranging from 1 in *Blastocladiella emersonii* (L22497) and *Rhizopus nigricans* (AY147870) *hsp70* genes to up to 8 in *Emericella nidulans* (X98931) *hsp70* gene. Characterization of the intron size range for intron-containing fungal *hsp70* gene is shown in Table 5. The frequency of the size distribution of intron-containing fungal *hsp70* gene was high; between 40-79, with a dominant frequency between 50-59 nucleotides. Moreover, almost all intron-containing fungal *hsp70* genes show at least 1 intron present in the amino acid terminal region in Hsp70 conserved-GIDLGTTYSCV (residues 6-16)-sequence, adenosine triphosphate-binding domain, such as V7 (*R. nigricans*, AY147871 and AY147869), S14 (*Puccinia graminis*, U26597), D13 (*B. emersonii*, L22497), G6 (*H. capsulatum*, U46464; *P. brasiliensis*, U91560), I7 (*Pneumocystis carinii* f.sp *carinii*, U80967), G15 (*E. nidulans*, X98931) and L9 (*C. neoformans* var.*neoformans*, AB126638) amino acid. It should be noted that this is a moderate conserved-characteristic of intron-containing fungal *hsp70* gene.

## 5.2 Characteristics of the deduced amino acid sequence

Deduced amino acid sequence of a clone containing the full-length *hsp70* transcript displayed strong homology to Hsp70s of both prokaryotic and eukaryotic origins. Alignment of the predicted *P. marneffei* protein sequence (*PmHsp70*) with reported sequences of Hsp70 by ClustalW analysis revealed high level of homology. Figure 8 showed the comparison of the *PmHsp70* with known Hsp70 sequences. Positions in which a gap was introduced to optimize homology were showed as dashes. High identity was found when comparing *PmHsp70* with other fungal Hsp70s. The sequence showed homology to Hsp70 sequences from other pathogenic fungi (87% to *P. brasiliensis* and 86% to *H. capsulatum* and *T. rubrum*), with the highest score identical to Hsp70 from *A. nidulans* (88 % identity). The presence of conserved sequence motifs of the Hsp70 family was observed in the amino acid sequence (Fig. 8): (1) GIDLGTTYS~~C~~V (residues 6-16); (2) NEPTAA (residues 172-177); and (3) DLGGGT~~F~~D (residues 197-204) (Rensing and Maier, 1994); (4) a KRKYKKDLTTNARALRR motif (residues 244-260), described as a putative nuclear localization signal (NLS) of eukaryotic Hsp70 homologues (Dingwall and Laskey, 1991); (5) a RARFEE motif (residues 297-302), which serves as a signature to eukaryotic non-organellar stress-70 proteins. The EELD amino acid present at the C-terminal of the protein was the same as those seen in *A. nidulans* (AY960135) and *Ustilago maydis* (EAK84826), whereas the other organism present the EEVD motif (highly conserved among members of the Hsp70 and Hsp90 protein families). Moreover, we also found the presence of five conserved sequence motifs in amino acid sequence of all 32 fungal cytoplasmic Hsp70s as followed; (I) TVPAYFNDSQRQ (residues 143-154); (II) GGSTRIP (residues 336-442); (III) VAYGAAVQA (residues 366-374); (IV) GIETAGG (residues 400-406); (V) DNQPGVLIQV (residues 431-440). The motifs I, II and III could be found in other family members of fungal Hsp70s, except that the motifs IV and V are found only fungal cytosolic Hsp70s.

### 5.3. Phylogenetic analysis

Since Hsp70 homologues are among the most highly conserved proteins, they are useful tools for phylogenetic analysis. The complete deduced protein sequences of PmHsp70 and 48 other fungal Hsp70 sequences obtained from GenBank were used for the phylogenetic tree construction. Figure 9 shows the deduced phylogeny using neighbour-joining analysis of amino acid sequences in MEGA3 program (Kumar *et al.*, 2004). The resulting tree shows 4 distinct branches. Three branches include the Hsp70 family members of different intracellular compartments (cytoplasmic, endoplasmic reticulum and mitochondria) and fourth branch represents the ribosome-associated Hsp70s. The *P. marneffei* Hsp70 was clustered with the fungal cytosolic Hsp70s and it was closely related with *A. nidulans* Hsp70.

### 5.4. Southern blot analysis of *P. marneffei* genomic DNA

To obtain the information about the genomic organization of the gene corresponding to *hsp70*, Southern blot analysis was carried out using *P. marneffei* genomic DNA (F4 strain). Total DNA was digested with the restriction endonucleases including *Bam*HI, *Sall*, *Hind*III, *Bgl*II, *Eco*RI, *Nde*I, *Xho*I, *Bss*SI and *Kpn*I . The digested DNA was fractionated on a 1% agarose gel and hybridized to the probes encoding PmHsp70, as shown in Figure 10B. High-stringency washes were used in the hybridization procedure. The uncut genomic DNA of *P. marneffei* (F4 strain) gave a strong-sharp band that indicated a complete genomic DNA. The enzymes that have no restriction site in the *Pm hsp70* gene (*Sall*, *Bgl*II and *Hind*III) resulted more than one band, whereas only *Bam*HI displayed one band matched with the expected band. The other enzymes that have one, two and three restriction site(s) (*Eco*RI, *Nde*I, *Xho*I, *Bss*SI and *Kpn*I) resulted the number of positive band more than the expected for single copy from the restriction maps of *Pmhsp70* (Fig. 10A). The results indicated that *P. marneffei* genome contained multiple copies of the *hsp70* gene.

### **5.5. Morphological changes during phase transition *in vitro***

To investigate saprobic and parasitic phase transition *in vitro*, conidial suspension of *P. marneffei* was cultured in liquid medium (BHI) at different temperatures. At 25°C (Fig. 11), spores of *P. marneffei* rapidly grew by swelling within the first 6 h. Most germ tubes were produced between 6-12 h and then they grew apically to hyphae. Hyphae growth was fully established in 3-4-day culture. The growth of *P. marneffei* in these conditions indicated saprobic phase transition. At 37°C (Fig. 11), the cellular differentiation stage was slower than at 25°C. Spore growth occurred within the first 12 h. Germ tubes were formed by polarization between 12 to 24 h. Then, some of germ tubes rapidly produced highly branched hyphae cells. Cellular differentiation to yeast-like cells began between 24 to 48 h. At 48 h, most germ tubes and branched hyphae cells went through arthroconidiation and segmentation. Most completed yeast cells were observed at 96 h after germination. This condition revealed the parasitic phase transition of *P. marneffei*. In addition, cellular differentiation from mycelial cells to yeast-like cells was observed. The dynamics of morphological changes paralleled with parasitic phase transition. However, the occurrence of yeast-like cell differentiation was earlier than those seen in parasitic phase transition. Moreover, complete yeast cells that habored with heat shock (39°C) and severe heat shock (42°C) within 3 h did not affect the morphology of this fungus and it could grow in SDA plate at 25°C.

### **5.6. Northern blot analysis of *hsp70* mRNA level during saprobic and parasitic phase transition *in vitro* and under heat shock conditions**

To investigate the differential expression of the *P. marneffei* *hsp70* gene at various stages and under heat shock condition, a Northern blot analysis was performed in which total RNA was probed with a cDNA fragment encoding the whole ORF of the Hsp70. The results showed a strong hybridization band at conidia sample, indicating an accumulation of *hsp70* transcripts in this stage (the dormant form of the organism) (Fig. 12). The faint bands were observed during the development to saprobic mycelial phase at 25°C (Fig. 12). It revealed that the *hsp70* transcript was downregulated during growth in natural conditions, with the exception of an upregulation at 48 h. A shift in the incubation temperature for the mycelium or spore from 25°C to 37°C, which required the fungus to

undergo the transition to yeast form, led to an immediate increase in the *hsp70* mRNA level (Fig. 12 and Fig. 13). However, the *hsp70* transcripts rapidly decreased paralleling with the differentiation of hyphae to yeast cells at 48 h, and until most cells were yeast cells at 72 hours.

The expression pattern of the *hsp70* gene was observed at normal homeostatic conditions (37°C) and during simulated human fever (39°C). When the incubation temperature of completed yeast cells (37°C, 96h) upshifted from 37°C to 39°C (Fig. 14), the *hsp70* mRNA rapidly increased within 30 min to the maximum level, and then moderately decreased at 3 h. However, a severe heat shock condition of 42°C resulted in lowering the *hsp70* transcript. It revealed the over-limitation condition to produce *hsp70* RNA of *P. marneffei*. In contrast to mycelial to yeast phase transition, decreasing the incubation temperature for the yeast cells from 37°C to 25°C did not affect the *hsp70* mRNA level.

### **5.7. Analysis of *hsp70* mRNA processing during saprobic and parasitic phase transition *in vitro***

The presence of putative introns in the *hsp70* gene led to investigate their occurrence in *hsp70* mRNA and splicing during saprobic and parasitic phase transition. Reverse transcription-PCR with primers flanking two introns revealed the presence of expected bands of spliced *hsp70* mRNA and intron(s) contained *hsp70* mRNA. A strong 617 bp band indicated the mature *hsp70* mRNA. The other weak higher band was likely corresponding to unspliced mRNAs. In order to confirm both intron-free and intron-containing fragments, size identity in a PCR using *hsp70*-containing plasmid (ph20) and genomic DNA as templates were performed (Fig. 15). The intron-free and intron-containing fragments were also confirmed by the DNA sequencing. Surprisingly, the DNA sequence of intron-containing fragment showed that it contained only intron II of *P. marneffei* *hsp70* gene (Fig. 16). A small population of intronII-unspliced *hsp70* mRNA was found during mycelial phase transition and during yeast phase transition. However, the unspliced *hsp70* mRNA level was low at yeast phase.

1 cctgtacactatcttctatcctcatttcctactttccgtatacccttcccatac  
 61 acacccacatacacacaacatacacattcaccATGGCCCCGCTATCGGTATCGATTGG  
   M A P A I G I D L G 10  
 121 GAACCACCTACTCCTCGGTGGGTGTCCTCCGTGATGACCGTATCGAAATCATTGCCAACG  
       T T Y S C V G V F R D D R I E I I A N D 30  
 181 ATCAGGGTAACCGAACCCACCCCTCGTCGTTGCCTTCACCGACTCCGAGCGTCTCATGG  
       Q G N R T T P S F V A F T D S E R L I G 50  
 241 GTGATGCTGCCAAGAACATCAGGTGCCATGAACCCCTACAACACAGTCATGCTAAGC  
       D A A K N Q V A M N P H N T V F D A K R 70  
 301 GTTTGATCGGCCGCAAATTCTCCGATCCTGAGGTCCAGGCTGATGCCAACGACTTCCCTT  
       L I G R K F S D P E V Q A D A K H F P F 90  
 361 TCAAGATCATCGAGAACGCCACCAAGCCGTTATCGAGGTCGAGGTTCAAGGGTGAGGTCA  
       K I I E K A T K P V I E V E F K G E V K 110  
 421 AGCAGTTCACACCTGAGGAAATCTCTTCATGGTCTGATCAAGATGCGTGAGACTGCTG  
       Q F T P E E I S S M V L I K M R E T A E 130  
 481 AGGCCTACCTCGGTGGTACCGTTAACACGCTGTACACTGTCCCCGCCTACTTCAACG  
       A Y L G G T V N N A V I T V P A Y F N D 150  
 541 ACTCCCAGCGTCAGGCCACCAAGGATGCTGGTCTCATGGCTTGAACGTCCTCCGTA  
       S Q R Q A T K D A G L I A G L N V L R I 170  
 601 TCATCAACGAACCTACTGCTGCCGCCATTGCCCTACGGTCTCGACAAGAAGGTTGAGGTG  
       I N E P T A A A I A Y G L D K K V E G E 190  
 661 AGCGCAACGTTCTCATCTCGATCTTGGTGGTACCTTCGATGTCTCTCCTCACCA  
       R N V L I F D L G G G T F D V S L L T I 210  
 721 TCGAGGACGGTATCTCGAGGTCAAGGCCACCGCCGGTACACTCACTTGGTGGTGGAGG  
       E D G I F E V K A T A G D T H L G G E D 230  
 781 ACTTCGACTCTCGCCTGTCAACCACCTTGCCTCCGAGGTTCAAGAGGAAATAAGAgt  
       F D S R L V N H F A S E F K R K Y K K 249  
 841 tgtatataccccatctgtttacaagaattgataagctaadaacttcacacagAGGgt  
   D 250  
 901 cgtcaaaaatccgaaatgaattctcattccctccgtctcttgatttttatggatatt  
 961 tggagcttgctctggagacttcctcagaaccatctgaagaatgcataatgaagactccca  
 1021 atatgaatccatctccctcccttcatccgcacttgaacttgcattttatggatattEq  
 1081 taadatgtgaaatagATTTGACCACCAATGCTCGTCTCGCCGTCTCCGCACTGCCT  
       L T T N A R A L R R L R T A C 265  
 1141 GTGAGCGTCTGCTAACCGTACCCCTCTCTTCTCCGCCAGACCTCATTGAGATCGACTCTC  
       E R A K R T L S S S A Q T S I E I D S L 285  
 1201 TCTTCGAGGGTATTGACTTCTACACCTCCATACCCGTGCTCGTTGAGGAGCTCTGCC  
       F E G I D F Y T S I T R A R F E E L C Q 305  
 1261 AGGATCTCTCCGTTCCACCATGGAGCCCGTCGAGCGTGTCCCGTGTGATGCCAACCG  
       D L F R S T M E P V E R V L R D A K T D 325  
 1321 ACAAGTCTCTGTCCACGAAATCGTCTTGGTCGGTGGTCCACCCGTATCCCCAAGATCC  
       K S S V H E I V L V G G S T R I P K I Q 345  
 1381 AGAAAGCTCGTCACCGACTTCTCAACAAGGAGCCAAACAAGTCCATCAACCCGATGAGG  
       K L V T D F F N K E P N K S I N P D E A 365  
 1441 CTGTTGCCTACGGTGCTGCCGTCCAGGCTGCTATCCTTCTGGTGACACTTCTCCAAGT  
       V A Y G A A V Q A A I L S G D T S S K S 385  
 1501 CCACCAACGAAATCTTGCCTCTCGACGTTGCTCCCTCTCCGTGGTATTGAGACTGCTG  
       T N E I L L D V A P L S V G I E T A G 405  
 1561 GAGGTGTCATGACTCCTCTCATCAAGCGAACACCACCATCCCCACCAAGAAGTCCGAGA  
       G V M T P L I K R N T T I P T K K S E T 425  
 1621 CCTTCTCCACCTACTCCGACAACCAGCCGGTGTGTTGATTCAAGGTCTACGAGGGTGAGC  
       F S T Y S D N Q P G V L I Q V Y E G E R 445  
 1681 AGTGCTCGTACCAAGGCAACAACTTGCTCGGCAAGTTCGAGCTCACTGGCATCCCCCTG  
       A R T K D N N L L G K F E L T G I P P A 465  
 1741 CTCCCTCGTGGTGTCTCAGATCGAGGTACCTCGACATGGACGCCAACGGTATCATGA  
       P R G V P Q I E V T F D M D A N G I M N 485  
 1801 AACGTCTCTGCCGTGGAAGGGTACCGGTAAGAGCAACAAGATTGTCATACCAACGACA  
       V S A V E K G T G K S N K I V I T N D K 505

**Figure 7.** The nucleotide sequence and deduced amino acid sequence of the *hsp70* gene from *P. marneffei*. The exon is indicated by upper case letters, while the intron, 5' and 3' non-translated nucleotides are indicated by lower case letters. Base numbers are on the left, and amino acid numbers are on the right. The putative splice boxes in the intron sequences are marked by rectangles. Nucleotides in bold italics represent the conserved 5' and 3' consensus of the introns.

**Table 4.** GenBank accession numbers of fungal Hsp70 sequences used in this study and numbers of introns in fungal *hsp70* genes

Organism	<i>hsp70</i> gene sequence accession number	No. of introns	Reference
<b>Ascomycota</b>			
<i>Penicillium marneffei</i>	AY960136	2	Kummasook <i>et al.</i> , 2005* this study
<i>Paracoccidioides brasiliensis</i>	U91560	2	da Silva <i>et al.</i> , 1999
<i>Ajellomyces capsulatus</i> ( <i>Histoplasma capsulatum</i> )	U46464	3	Allendoerfer <i>et al.</i> , 1996
<i>Trichophyton rubrum</i>	AF952391@	2	Rezaie <i>et al.</i> , 2000
<i>Aspergillus awamori</i>	Y12504	3	Hijarrubia <i>et al.</i> , 1997
<i>Aspergillus oryzae</i>	AB030231	3	Kasuya <i>et al.</i> , 1999
<i>Aspergillus niger</i>	Y08868	3	van Gemeren <i>et al.</i> , 1997
<i>Emericella nidulans</i> ( <i>Aspergillus nidulans</i> )	X98931	8	Kramer, 1996*
<i>Neurospora crassa</i>	U10443 Y09011	4 5	Kapoor <i>et al.</i> , 1995 Techel <i>et al.</i> , 1998
<i>Pneumocystis carinii f.sp carinii</i>	U80967 L46790	2 4	Stedman <i>et al.</i> , 1998 Stedman and Buck, 1996
<i>Pneumocystis carinii f.sp ratti</i>	U40994	3	Stedman and Buck, 1996
<b>Zygomycota</b>			
<i>Rhizopus nigricans</i> ( <i>Rhizopus stolonifer</i> )	AY147871 AY147870 AY147869	≥2 1 3	Cernila <i>et al.</i> , 1999 Cernila <i>et al.</i> , 1999 Cernila <i>et al.</i> , 2003
<b>Basidiomycota</b>			
<i>Puccinia graminis</i>	U26597	4	Staples, 1995*
<i>Cryptococcus neoformans</i> var. <i>neoformans</i>	AB126638	5	Kakeya <i>et al.</i> , 2003*
<b>Chytridiomycota</b>			
<i>Blastocladiella emersonii</i>	L22497	1	Stefani and Gomes, 1995

@ Acession number of cDNA

\* Indicates direct submission to GenBank. These entries are not included in "References"

**Table 5.** Frequency of intron lengths from fungal intron-containing *hsp70* genes

<b>Length (base) of introns</b>	<b>Frequency of <i>hsp70</i> introns</b>
40 - 49	5
50 - 59	29
60 - 69	10
70 - 79	5
80 - 89	1
90 - 99	-
100 - 109	3
110 - 119	-
120 - 129	-
130 - 139	-
140 - 149	-
150 - 159	-
160 - 169	3
170 - 179	1
180 - 189	1
190 - 199	1

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่  
 Copyright © by Chiang Mai University  
 All rights reserved





Pb-AAB53051	DNRLVNHFVNEFKRKHK	-KDLSSN-ARALRRLRTACERA	267
Pb-AAK66771	DNRLVNHFVNEFKRKHK	-KDLSSN-ARALRRLRTACERA	268
Ac-AAC05418	DNRLVNHFVSEFKRKFK	-KISPAERARALRSPTACERA	269
Tr-AAD08909	DNRLVNHFVNEFKRKKNK	-KDLSTN-ARALRRLRTACERA	268
Tv-AAQ83701	DNRLVNHFVNEFKRKKNK	-KDLSTN-ARALRRLRTACERA	268
Dt-CAA57452	DNRLVNHFSNEFKRKHK	-KDLSDN-ARALRRLRTACERA	268
Gz-EAA70431	DNRLVNHFVNEFKRKHK	-KDLSTN-VRALRRLRTACERA	268
Hj-AAP40020	DNRLVNHFVNEFKRKHKKVSSSTHVAPHSLDARHMLTPDRDLSTN	-ARALRRLRTACERA	290
Mg-EAA55301	DNRLVTHFANEFKRKHK	-KDLTTN-ARALRRLRTACERA	268
Nc-AAA82183	DNRLVNHFVQEFKRKDK	-KDLSTN-ARALRRLRTACERA	268
Pm-AY960135	DSRLVNHFASEFKRKYK	-KDLTTN-ARALRRLRTACERA	268
An-EAA62310	DNRLVNHFVTEFKRKHK	-KDLSTN-ARALRRLRTACERA	268
Pc-AAD00455	DNRLVQHFVQEFKRKHK	-KDISGN-PRALRRLRTACERA	270
Cn-AAW42238	DNRLVNHFVQEFKRKKNK	-KDLSSN-ARALRRLRTACERA	268
Cn-EAL21768	DNRLVNHFVQEFKRKKNK	-KDLSSN-ARALRRLRTACERA	268
Cn-AAW42202	DNRLVNHFVQEFKRKKNK	-KDLSSN-ARALRRLRTACERA	268
Cc-CAA72797	DNRLVNHFVQEFKRKKNK	-KDITSN-ARALRRLRTACERA	266
Um-EAK84826	DNRLVNHFVQEFKRKKNK	-KDLTTN-ARALRRLRTACERA	268
Rn-AAF13877	DNRLVAHFMQEFKRKF	-KDITGN-ARAIRRLRTACERA	270
Rn-AAN52148	DNRLVSHFMQEFKRKFK	-KDITGN-ARAIRRLRTACERA	270
Rn-AAN52149	DNRLVDHF1QEFKRKFK	-KDITGN-ARAVRRLRTACERA	268
Pg-AAB93665	DNRLVNHFVQEFKRKHK	-KDLSSN-PRALRRLRTACERA	268
Sp-CAA20787	DSRLVNHFIQEFKRKNK	-KDITGN-ARAVRRLRTACERA	268
Sp-CAA93590	DSRLVNHFAQEFKRKNK	-KDITGN-ARAVRRLRTACERA	268
Ca-CAA82929	DNRLVNFF1QEFKRKNK	-KDITGN-QRALRRLRTACERA	269
Ca-EAK94611	DNRLVNFF1QEFKRKNK	-KDITGN-QRALRRLRTACERA	268
Sc-AAA63574	DSRLVNFLAEEFKRKNK	-KDLTTN-QSRLRRLRTAAERA	268
Sc-AAC37398	DNRLVNHLATEFKRKTK	-KDISNN-QSRLRRLRTAAERA	268
Pa-AAB63968	DNRLVNHFINEFKRKNK	-KDICGN-QRALRRLRTACERA	268
Pa-CAA82570	DNRLVNHFANEFKRKYK	-KDLTTN-QRALRRLRTACERA	270
Sc-CAA31393	DNRLVNHFIQEFKRKNK	-KDLSTN-QRALRRLRTACERA	267
Sc-CAA31394	DNRLVNHFIQEFKRKNK	-KDLSTN-QRALRRLRTACERA	267
*.*** . : * * * * *		:	: * : * * * ** . ***
Pb-AAB53051	KRTLSSAAQTSIEIDSLYEGIDFYTSITRARFEELCQDLFRSTMDPVERVLRAKIDKSS	327	
Pb-AAK66771	KRTLSSAAQTSIEIDSLYEGIDFYTSITRARFEELCQDLFRSTMDPVERVLRAKIDKSS	328	
Ac-AAC05418	KRTLSSAAQTSIEIDSLYEGIDFYTSITRARFEELCQDLFRSTMEPVERVLRAKIDKSS	329	
Tr-AAD08909	KRTLSSAAQTSIEIDSLYEGVDFYTSITRARFEELCQDLFRSTMEPVERVLRAKIDKSS	328	
Tv-AAQ83701	KRTLSSAAQTSIEIDSLYEGVDFYTSITRARFEELCQDLFRSTMEPVERVLRAKIDKSS	328	
Dt-CAA57452	KRTLSSAAQTSIEIDSLFEGIDFTSNTRARFEELVGQDLFRGNMEPGERTLRRDKIDKSS	328	
Gz-EAA70431	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCQDLFRSTIOPVDRVLTDAKIDKSL	328	
Hj-AAP40020	KRTLSSAAQTSIEIDSLYEGIDYYTSITRARFEELCQDLFRSTIOPVDRVLADAKIDKSQ	350	
Mg-EAA55301	KRTLSSAAQTSIEIDSLYEGIDFTSITRARFEELCQDLFRSTLQPVDRVLTDAKIDKAQ	328	
Nc-AAA82183	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCQDLFRSTLQPVDRVLTDAKIDKSQ	328	
Pm-AY960135	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCQDLFRSTMEPVERVLRAKTDKSS	328	
An-EAA62310	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCQDLFRGTMEPVERVLRAKIDKSS	328	
Pc-AAD00455	KRSLSSSTQTSIEIDSLYEGIDLYTSITRARFEELCQDLFRGTMEPVEKVLRAKIDKSS	330	
Cn-AAW42238	KRTLSSAAQTSIEIDSLFDGIDFTSITRARFEELCQDLFRSTMDPVEKVLRDSKIDKSS	328	
Cn-EAL21768	KRTLSSAAQTTIEIDSLFDGIDFTSITRARFEELCQDLFRSTMDPVEKVLRDSKIDKSS	328	
Cn-AAW42202	KRTLSSAAQTSIEIDSLFDGIDFTSITRARFEELCQDLFRSTMDPVEKVLRDSKIDKSS	328	
Cc-CAA72797	KRTLSSAAQTSIEIDSLYDGIDFTSITRARFEELCQDLFRSTMDPVEKVLRDSKIDKSS	326	
Um-EAK84826	KRTLSSAAQTTIEIDSLFEGIDFTSITRARFEELCQDLFSHTIEPVEKVLRDSKIDKGS	328	
Rn-AAF13877	KRTLSSAAQTTIEIDSLFEGIDFTSITRARFEELCQDLFSHTIEPVEKVLRDSKIDKGS	330	
Rn-AAN52148	KRTLSSAAQTTIEIDSLFEGVDFYTSITRARFEELNQDLFRNTMEMPVEKVLRDSKIDKSQ	330	
Rn-AAN52149	KRTLSSAAQTSIEIDSLFEGVDFYTSITRARFEELNQDLFRNTMEMPVEKVLRDSKIDKSQ	328	
Pg-AAB93665	KRTLSSAAQTTIEIDSLFEGVDFYTSITRARFEELNQDLFRNTMEMPVEKVLRDSKLKG	328	
Sp-CAA20787	KRTLSSAAQASIEIDSLFEGIDFTSITRARFEELCADLFRSTLEPVEKVLRAKIDKAA	328	
Sp-CAA93590	KRTLSSAAQASIEIDSLYEGIDFTSITRARFEELCADLFRNTMEMPVEKVLRDSKVDKAS	328	
Ca-CAA82929	KRTLSSAAQTSIEIDSLYEGIDFTSITRARFEELCADLFRNTMEMPVEKVLRAKIDKSS	328	
Ca-EAK94611	KRTLSSAAQTSIEIDSLYEGIDFTSITRARFEELCADLFRNTMEMPVEKVLRAKIDKSQ	329	
Sc-AAA63574	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCADLFRSTLEPVEKVLRAKIDKSK	328	
Sc-AAC37398	KRTLSSAAQTSIEIDSLYEGIDFTSITRARFEELCADLFRSTLEPVEKVLRAKIDKSK	328	
Pa-AAB63968	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCADLFRSTLEPVEKVLRAKIDKSK	328	
Pa-CAA82570	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCADLFRSTLEPVEKVLRAKIDKSK	328	
Sc-CAA31393	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCADLFRSTLEPVEKVLRAKIDKSK	327	
Sc-CAA31394	KRTLSSAAQTSIEIDSLFEGIDFTSITRARFEELCADLFRSTLEPVEKVLRAKIDKSK	327	
*.*** . : * * * * *		:	: * * * . : * * * *



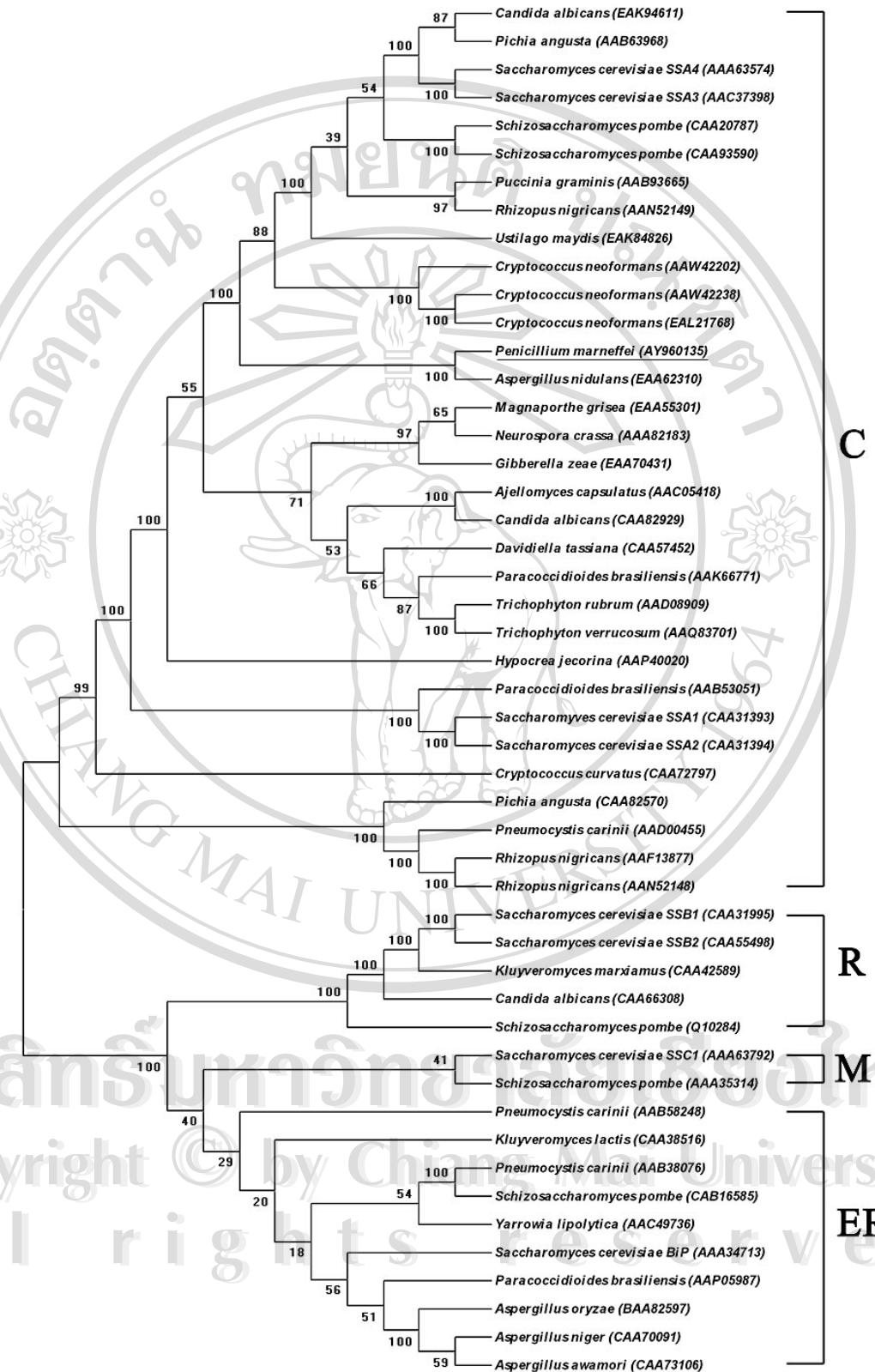


Pb-AAB53051	LKTEIDKTVSWLDENQTATKEEFEAQQKELESVANPIMMKFYGAGG--APGAGFPAG-	622
Pb-AAK66771	LKTEIDKTVSWLDENQTATKEEFEAQQKELESVANPIMMKFYGAGGEGGAPGAGFPAG-	627
Ac-AAC05418	LKSEIDKTVQWLDENQTATKEEYESQQKELEAVANPIMMKFYAGGE--GAPGG-FPGAG-	624
Tr-AAD08909	LKSEIDKVVAWLDDNQTATKEEYESQQKELEGVANPIMMKFYGAGGEGGAPGG-FPGAGA	627
Tv-AAQ83701	LKSEIDKVVAWLDDNQTATKEEYESQQKELEGVANPIMMKFYGAGGEGGAPGG-FPGAGA	627
Dt-CAA57452	LTGAIDKTVAWIDENQTATKEEYEAEQKQLESVANPVMMKIYGAEG--GAPGG-MPGQG-	624
Gz-EAA70431	LTAELDKVVQWLDDNNQQATREEEYEHQKELEGKANPIMMKFYGAGG--EGAPGGMPG-MP	625
Hj-AAP40020	LKSEIDKIVQWLDDNNQQASTEEYESHQKELEGVANPIMMKFYGAGG--E---GGMPGGMP	645
Mg-EAA55301	LNAEINKIVSWLDDESQQATKEEYEHHQKELEAVANPIMMKFYGAGG--AP--GGMPGAPG	624
Nc-AAA82183	LKSEIDKIVAWLDENQQATREEEYERQKELEAIANPIMMKFYGAGG--AP--GGMP---G	621
Pm-AY960135	LEAEIEKTIISWLDNSNQTATKDEYEAAQKQLESVANPIISAAYGAG-----AAPG--	615
An-EAA62310	VSDKIDEVISWLDNNQTAEKDEYESQQKELEGVANPIISAAYAAG-----GAPGGAA	618
Pc-AAD00455	LEKAISDVTWSLDELTNTATKEEYTSKQKDLETIAGPIMMKLYQSGE-----GVPGMG-	621
Cn-AAW42238	LSKKVEEVINALDTMQSASKEFESLQKELEGIANPIMMKFYGAGS-----GMPGGA-	617
Cn-EAL21768	LSKKVEEVINALDTMQSASKEFESLQKELEGIANPIMMKFYGAGS-----GMPGGA-	617
Cn-AAW42202	LSKKVDEVISSLDTMQSASKEFESLQKELEAVANPIMMKFYGAQG-----GAPG--	615
Cc-CAA72797	LQKAVDDCIKFLTADSASKDEIESHQKELEALSGPIMQRFYCSTG-----GAPGGAP	616
Um-EAK84826	LEKIVKEGLEWLDSNTTASTDELKDQKIEEEQVNPIMMKIYSAAG-----GAPGGMP	620
Rn-AAF13877	LNAAVDESIKWLDSEQEASKEEYESQKQLEELIANPIMMKFYQQAG-----GAPG--	620
Rn-AAN52148	LNAAVDESIKWLDSEQEASKEEYESQKQLEELEVANPIMMKFYQQAG-----GAPG--	620
Rn-AAN52149	LESAVKEAIDWMNDNSQEASKEEYESRQKELEEVANPIMMKLYQGEG-----GMPGGG-	619
Pg-AAB93665	LEDAVNSTISWLDNSQEASKEEYESHQKELEAVANPIMQKLYYAGAG-----GAPGGAP	620
Sp-CAA20787	VDKAVKETIEWLDSNTTAAKDEFEAQKQLESVANPIMAKIYQAGG-----APGGMP	619
Sp-CAA93590	IDKAVKETIEWLDSNTTAAKDEFEAQKQLESVANPIMAKIYQAGG-----APGGAP	619
Ca-CAA82929	LTKAIDETISWLDASQAASTEEYEDKRKELESVANPIISGAYGAAGGAPGGAGFPAG-	627
Ca-EAK94611	VTKAADETIWLDNSNQTATQEEFADQQKELESKANPIMTKAYQAGATPSGAAGAAP--G	624
Sc-AAA63574	LEAAQAQDAINWLDSAQAASTEEYKERQKELEGVANPIMSKFYGAAGGA-PGAG-----	619
Sc-AAC37398	LETASQETIDWLDSAQAASTDEYKDRQKELEGIANPIMTKFYGAGAGAGPGAGESG-	623
Pa-AAB63968	FTKACDDTTIAWLDENQTATAEEYDDKRKELEQAGNEVLKDLYAEGGVPG--G-APG--	620
Pa-CAA82570	LNKAIETEISWLDNNQSATTDEYEDKRKELEGIANDALKDLYAAGGVPG--GAAPG--	623
Sc-CAA31393	VTKKAEETISWLDNSNTTASKEFFDDKLKELOQDIANPIMSKLYQAGGAPGGAAGGAPG--	621
Sc-CAA31394	VTKKAEETIAWLDSNTTATKEEFDQLKELOQEVANPIMSKLYQAGGAP--EGAAPG--	618
. . : * : * : * : . : *		
Pb-AAB53051	-----GPFGFP-----AGVGGAHSGGDDGPTVEEV-----	649
Pb-AAK66771	-----GPFGFP-----AGAGGAHSGGDDGPTVEEV-----	654
Ac-AAC05418	-----GPFGFPG-----PGAGHASGGGDDGPTVEEVDLKFPMPLPWQLSRKMHRPFF	674
Tr-AAD08909	-----GGPGFP-----AGAGGAAA-DDGPTVEEV-----	654
Tv-AAQ83701	-----GGPGFP-----AGAGGAAA-DDGPTVEEV-----	654
Dt-CAA57452	-----AGAPP-----PGAG-----DDGPTVEEV-----	643
Gz-EAA70431	-----GGPGFP-----AGGPAPGAGGDDGPTVEEV-----	653
Hj-AAP40020	-----GGAGGFPG-----AGG-----APHQGGDDGPTVEEV-----	672
Mg-EAA55301	-----GAPGGFP-----AGG-----APGAGGDDGPTVEEV-----	651
Nc-AAA82183	-----AAPGGFP-----GG-----APGSNDNEGPTVEEV-----	646
Pm-AY960135	-----ATG-----ASATREADEVEERPEELD-----	636
An-EAA62310	-----PGAGAAPG-----GGAGFRNDGVVENEELD-----	644
Pc-AAD00455	-----GACSPQG-----GPFGTDDDHGPTIEEV-----	645
Cn-AAW42238	-----GAPGGFPG-----GAGGGAAQEEGPSVVEEV-----	644
Cn-EAL21768	-----GAPGGFP-----GAGGGAAQEEGPSVVEEV-----	644
Cn-AAW42202	-----GAPGGFP-----AGGAPAQEEGPSVVEEV-----	640
Cc-CAA72797	-----GGAPGGFP-----AGGPGASHEDGPSVVEEV-----	643
Um-EAK84826	-----GGAPGAAPG-----GAAPGGDDGPTVEELD-----	645
Rn-AAF13877	-----APGAAPGGFP-----G-AAPGSTDETGPSIEEV-----	651
Rn-AAN52148	-----APGAAPGGFP-----G-AAPGSTDETGPSIEEV-----	651
Rn-AAN52149	-----GMPGGGAPGGFPG-----DTGGEPTVEEV-----	645
Pg-AAB93665	-----GGFPGGAPGGFPG-----APAGEDGPSVVEEV-----	648
Sp-CAA20787	-----G-----AAPGAAPG-----AAPGAAPGGDNGPVEEV-----	647
Sp-CAA93590	-----G-----GMPGGAPG-----GAPG-----GADNGPVEEV-----	644
Ca-CAA82929	-----G-----GFPGGAPGAGGGPGGATGGESSGPTVEEV-----	656
Ca-EAK94611	-----G-----G-----GAAPEPSNDGPTVEEV-----	645
Sc-AAA63574	-----PVPG-----AGAGPTG-----APDNNGPTVEEV-----	642
Sc-AAC37398	-----GFPGS-----MPNSGATGGGEDTGPTVEEV-----	649
Pa-AAB63968	-----GFPGA-----GGAPSTEETQGPTVEEV-----	643
Pa-CAA82570	-----GFPGA-----GGAAPGAD-----QGPSVVEEV-----	645
Sc-CAA31393	-----GFP-----GGAPPAPEAEGPTVEEV-----	642
Sc-CAA31394	-----GFP-----GGAPPAPEAEGPTVEEV-----	639

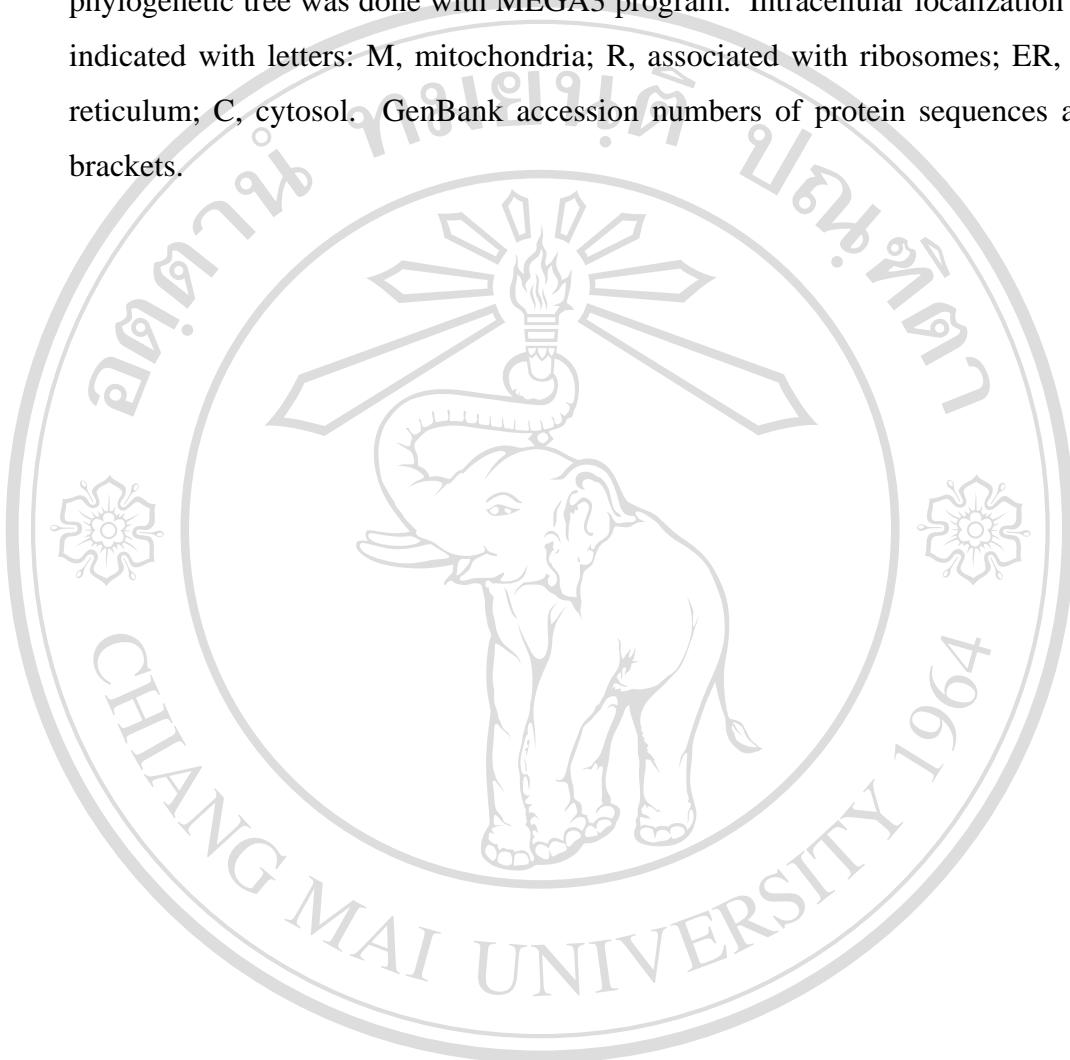
\*\* : \*

Pb-AAB53051	-----
Pb-AAK66771	-----
Ac-AAC05418	<u>L</u> FLLFLIFLIFLILFLFYFFLPVRFNESCFS 705
Tr-AAD08909	-----
Tv-AAQ83701	-----
Dt-CAA57452	-----
Gz-EAA70431	-----
Hj-AAP40020	-----
Mg-EAA55301	-----
Nc-AAA82183	-----
Pm-AY960135	-----
An-EAA62310	-----
Pc-AAD00455	-----
Cn-AAW42238	-----
Cn-EAL21768	-----
Cn-AAW42202	-----
Cc-CAA72797	-----
Um-EAK84826	-----
Rn-AAF13877	-----
Rn-AAN52148	-----
Rn-AAN52149	-----
Pg-AAB93665	-----
Sp-CAA20787	-----
Sp-CAA93590	-----
Ca-CAA82929	-----
Ca-EAK94611	-----
Sc-AAA63574	-----
Sc-AAC37398	-----
Pa-AAB63968	-----
Pa-CAA82570	-----
Sc-CAA31393	-----
Sc-CAA31394	-----

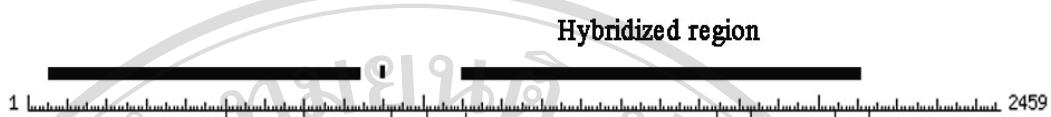
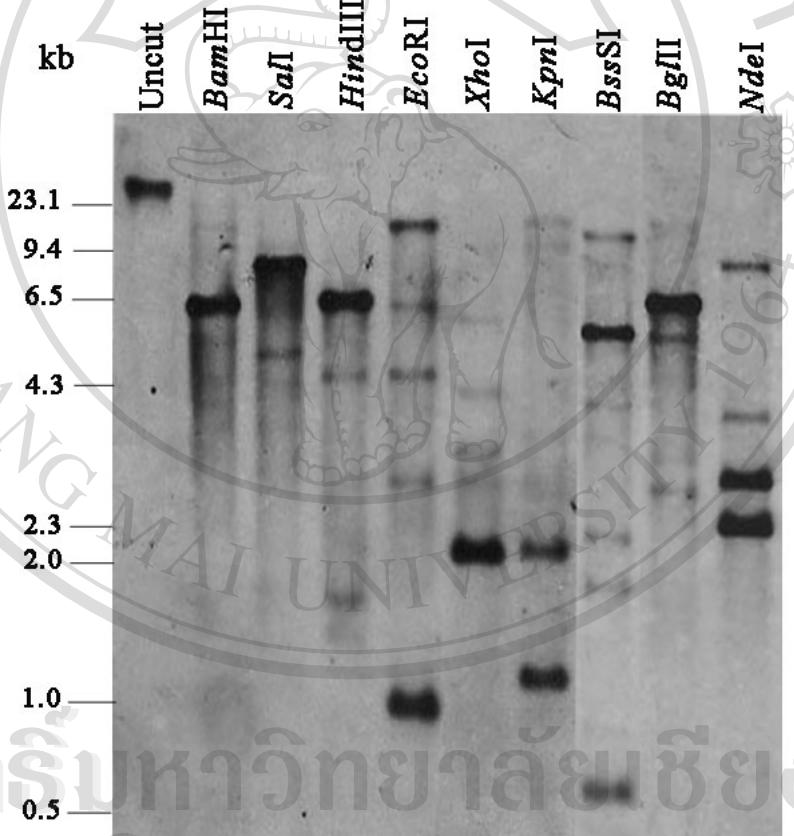
**Figure 8. Alignment of the amino acid sequences.** Amino acid sequences were obtained from *Penicillium marneffei* (Pm), *Saccharomyces cerevisiae* (Sc), *Paracoccidioides brasiliensis* (Pb), *Ajellomyces capsulatus* (Ac), *Trichophyton rubrum* (Tr), *Trichophyton verrucosum* (Tv), *Davidiella tassiana* (Dt), *Gibberella zeae* (Gz), *Hypocrea jecorina* (Hj), *Magnaporthe grisea* (Mg), *Neurospora crassa* (Nc), *Aspergillus nidulans* (An), *Pneumocystis carinii* (Pc), *Cryptococcus neoformans* (Cn), *Cryptococcus curvatus* (Cc), *Ustilago maydis* (Um), *Rhizopus nigricans* (Rn), *Puccinia graminis* (Pg), *Schizosaccharomyces pombe* (Sp), *Candida albicans* (Ca) and *Pichia angusta* (Pa). The alignment was performed using ClustalW program. The vital conserved sequence motifs are underlined. The putative conserved sequence motifs for fungal cytoplasmic Hsp70s are boxed.



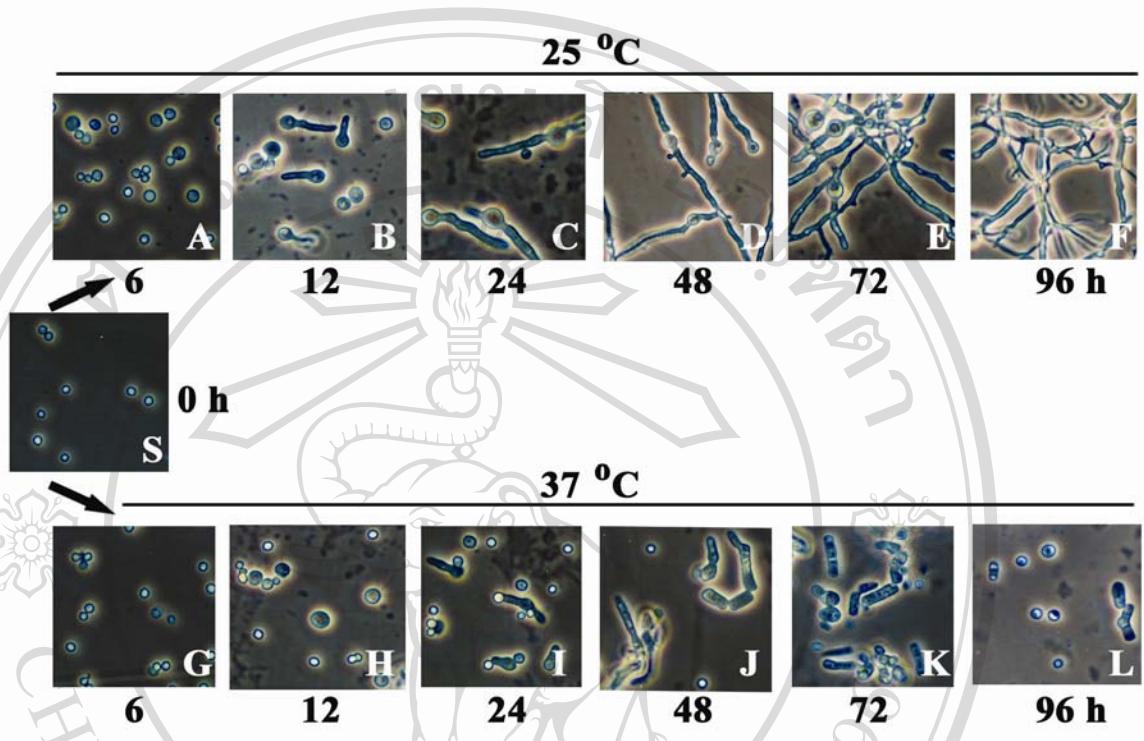
**Figure 9. Neighbor-joining phylogenetic tree of fungal heat shock protein 70 (Hsp70s).** Alignment of complete deduced amino acid sequences and construction of phylogenetic tree was done with MEGA3 program. Intracellular localization of Hsp70s is indicated with letters: M, mitochondria; R, associated with ribosomes; ER, endoplasmic reticulum; C, cytosol. GenBank accession numbers of protein sequences are shown in brackets.



ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่  
Copyright © by Chiang Mai University  
All rights reserved

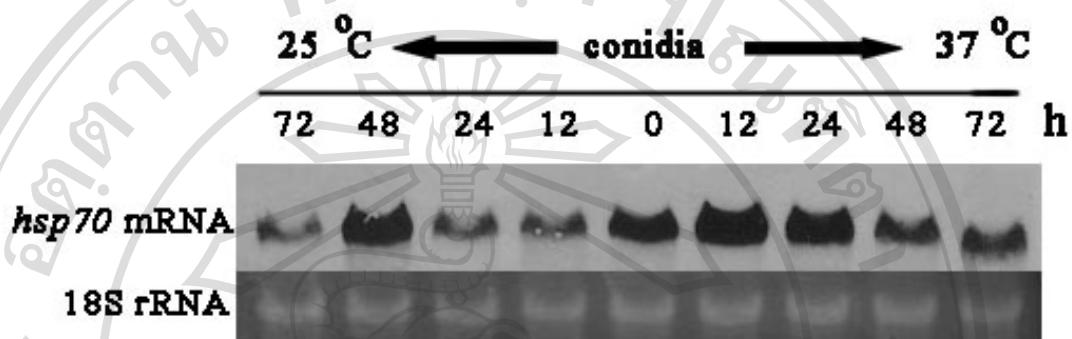
**A****B**

**Figure 10.** Southern blot analysis of *hsp70*. Restriction map (A) shows the cut sites of restriction enzymes used in Southern blot analysis (B). Immobilized digested genomic DNA of *P. marneffei* F4 strain was probed with *hsp70* gene fragment under high stringency condition. Hybridized region is shown in the black bar. Sizes of Lambda *HindIII* marker are shown in the left panel.



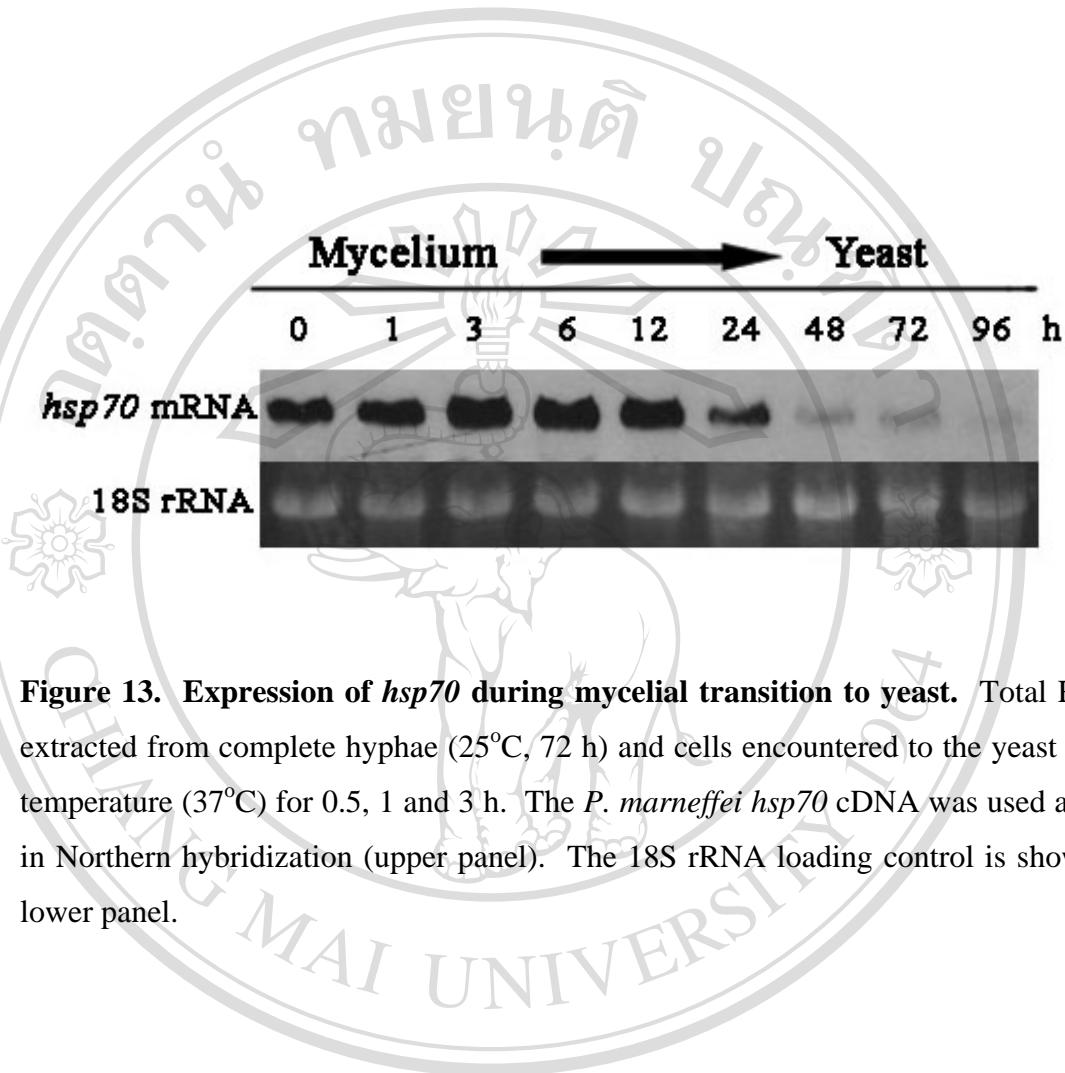
**Figure 11. Saprobic-parasitic-phase transition of *P. marneffei*.** Conidia (S) were incubated at 25°C or 37°C in brain heart infusion broth. At 25°C (A to F), conidia were swelled within 6 h. Germ tubes were produced between 6-12 h. Hyphae growth was fully established in 3-4-day culture. At 37°C (G to L), conidia growth occurred within the first 12 h. Germ tubes were formed by polarization between 12 to 24 h. Cellular differentiation to yeast-like cells began 48 h. Most completed yeast cells were observed at 96 h after germination.

Copyright © by Chiang Mai University  
All rights reserved



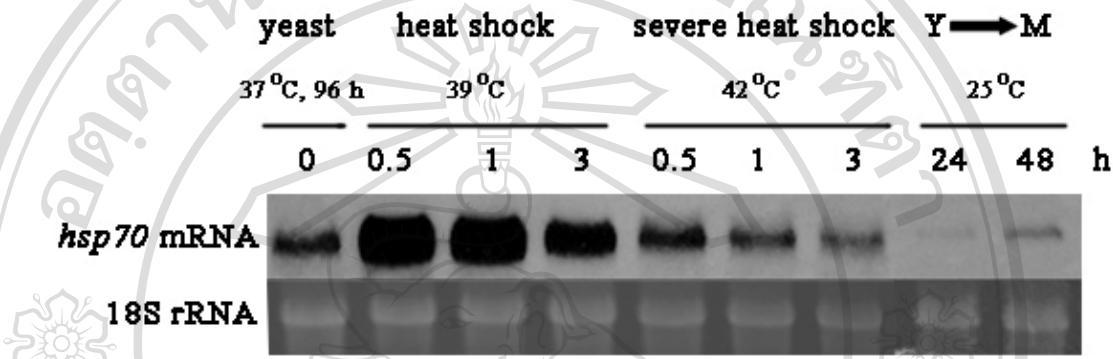
**Figure 12. Expression of *hsp70* during mold and yeast phase transition by Northern hybridization.** Total RNA was extracted from spore, mycelial and yeast cells at different time intervals during the temperature-induced morphological transition of the fungus. *P. marneffei* *hsp70* cDNA was used as a probe (upper panel) in Northern hybridization. The 18S rRNA loading control is shown in the lower panel.

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่  
Copyright © by Chiang Mai University  
All rights reserved

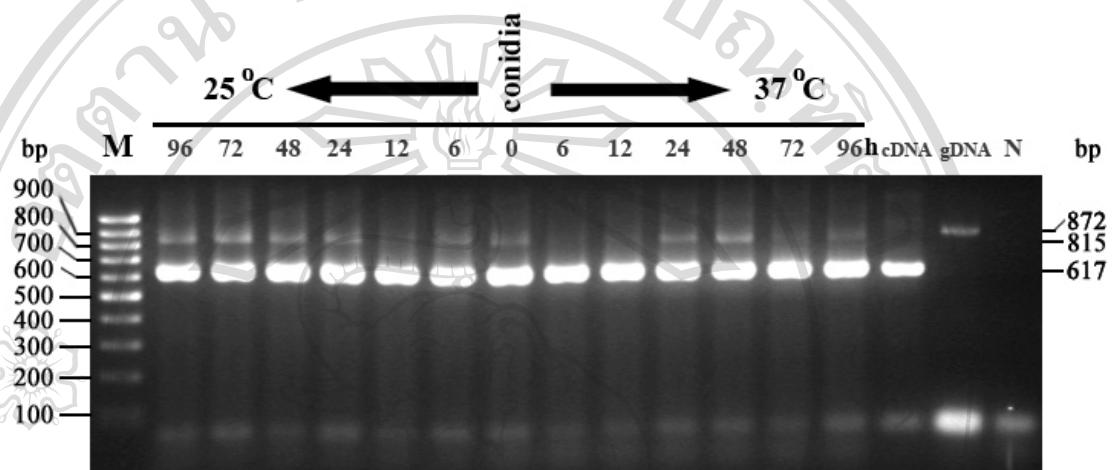


**Figure 13. Expression of *hsp70* during mycelial transition to yeast.** Total RNA was extracted from complete hyphae ( $25^{\circ}\text{C}$ , 72 h) and cells encountered to the yeast inducible temperature ( $37^{\circ}\text{C}$ ) for 0.5, 1 and 3 h. The *P. marneffei* *hsp70* cDNA was used as a probe in Northern hybridization (upper panel). The 18S rRNA loading control is shown in the lower panel.

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่  
Copyright © by Chiang Mai University  
All rights reserved



**Figure 14. Expression of *hsp70* during heat shock and the reverse to room temperature conditions.** Total RNA was extracted from complete yeast cells (37 °C, 96 h); cells encountered the heat shock (39°C) and severe heat shock (42°C) conditions for 0.5, 1 and 3 h; and after reverse to room temperature for 24 and 48 h. The *P. marneffei* *hsp70* cDNA was used as a probe in Northern hybridization (upper panel). The 18S rRNA loading control is shown in the lower panel.



**Figure 15. Analysis of *hsp70* mRNA splicing during saprobic (to 25°C) and parasitic (to 37°C) phase transition by RT-PCR.** The molecular marker indicated on the left and the expected sizes of mature *hsp70* mRNA (617 bp), intronII-unsPLICED *hsp70* mRNA (815 bp), and a fragment from PCR which contained 2 introns (872 bp) are indicated on the right. Control templates; cDNA (complementary DNA), gDNA (genomic DNA) and N (negative control).

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่  
Copyright © by Chiang Mai University  
All rights reserved

RNAM13R+F CAGTCCTCGATGCTAAGCGTTGATCGGCCGAAATTCTCGATCCTGAGGTCCAGGCTG 60  
 G20M13F+R CAGTCCTCGATGCTAAGCGTTGATCGGCCGAAATTCTCGATCCTGAGGTCCAGGCTG 60  
 17M13F+R CAGTCCTCGATGCTAAGCGTTGATCGGCCGAAATTCTCGATCCTGAGGTCCAGGCTG 60  
 \*\*\*\*\*  
**PMH70-F2**

RNAM13R+F ATGCCAAGCACTTCCCTTCAAGATCATCGAGAAGGCCACCAAGCCCCTATCGAGGTCTG 120  
 G20M13F+R ATGCCAAGCACTTCCCTTCAAGATCATCGAGAAGGCCACCAAGCCCCTATCGAGGTCTG 120  
 17M13F+R ATGCCAAGCACTTCCCTTCAAGATCATCGAGAAGGCCACCAAGCCCCTATCGAGGTCTG 120  
 \*\*\*\*\*  
 RNAM13R+F AGTCAAGGGTGAGGTCAAGCAGTCACACCTGAGGAATCTCTCCATGGTCTTGTATCA 180  
 G20M13F+R AGTCAAGGGTGAGGTCAAGCAGTCACACCTGAGGAATCTCTCCATGGTCTTGTATCA 180  
 17M13F+R AGTCAAGGGTGAGGTCAAGCAGTCACACCTGAGGAATCTCTCCATGGTCTTGTATCA 180  
 \*\*\*\*\*  
 RNAM13R+F AGATGCGTGAGACTGCTGAGGCCTACCTCGGTGGTACCGTTAACAACGCTGTCACTCG 240  
 G20M13F+R AGATGCGTGAGACTGCTGAGGCCTACCTCGGTGGTACCGTTAACAACGCTGTCACTCG 240  
 17M13F+R AGATGCGTGAGACTGCTGAGGCCTACCTCGGTGGTACCGTTAACAACGCTGTCACTCG 240  
 \*\*\*\*\*  
 RNAM13R+F TCCCCGCCTACTTCAACGACTCCCGCAGCGTCAGGCCACCAAGGATGCTGGTCTCATTGCCG 300  
 G20M13F+R TCCCCGCCTACTTCAACGACTCCCGCAGCGTCAGGCCACCAAGGATGCTGGTCTCATTGCCG 300  
 17M13F+R TCCCCGCCTACTTCAACGACTCCCGCAGCGTCAGGCCACCAAGGATGCTGGTCTCATTGCCG 300  
 \*\*\*\*\*  
 RNAM13R+F GTTGAACGTCTCCGTATCATCAACGAACCTACTGCTGCCATTGCCACGGTCTCG 360  
 G20M13F+R GTTGAACGTCTCCGTATCATCAACGAACCTACTGCTGCCATTGCCACGGTCTCG 360  
 17M13F+R GTTGAACGTCTCCGTATCATCAACGAACCTACTGCTGCCATTGCCACGGTCTCG 360  
 \*\*\*\*\*  
 RNAM13R+F ACAAGAAGGTTGAGGGTGAGCGCAACGTTCTCATCTCGATCTTGGTGGTACCTTCG 420  
 G20M13F+R ACAAGAAGGTTGAGGGTGAGCGCAACGTTCTCATCTCGATCTTGGTGGTACCTTCG 420  
 17M13F+R ACAAGAAGGTTGAGGGTGAGCGCAACGTTCTCATCTCGATCTTGGTGGTACCTTCG 420  
 \*\*\*\*\*  
 RNAM13R+F ATGTCCTCTCCCTCACCATCGAGGACGGTATCTCGAGGTCAAGGCCACCGCCGGTGACA 480  
 G20M13F+R ATGTCCTCTCCCTCACCATCGAGGACGGTATCTCGAGGTCAAGGCCACCGCCGGTGACA 480  
 17M13F+R ATGTCCTCTCCCTCACCATCGAGGACGGTATCTCGAGGTCAAGGCCACCGCCGGTGACA 480  
 \*\*\*\*\*  
 RNAM13R+F CTCACTTGGTGGTGGAGACTTCGACTCTGCCATTGTCAACCACCTTGCTCCGAGTTCA 540  
 G20M13F+R CTCACTTGGTGGTGGAGACTTCGACTCTGCCATTGTCAACCACCTTGCTCCGAGTTCA 540  
 17M13F+R CTCACTTGGTGGTGGAGACTTCGACTCTGCCATTGTCAACCACCTTGCTCCGAGTTCA 540  
 \*\*\*\*\*  
**Intron I**  
 RNAM13R+F AGAGGAAATATAAGA----- 555  
 G20M13F+R AGAGGAAATATAAGAGTATGTGATATAACCCCATCTGTTTACAAGAATTGATAAGCTAA 600  
 17M13F+R AGAGGAAATATAAGA----- 555  
 \*\*\*\*\*  
 RNAM13R+F -----AGG----- 558  
 G20M13F+R CAACATTTCACAGAGGGTACGTCAAAATCCGAAATGAATTCTCATCCCTCCGTCTCTT 660  
 17M13F+R -----AGGTACGTCAAAATCCGAAATGAATTCTCATCCCTCCGTCTCTT 603  
 \*\*\*  
**Intron II**  
 RNAM13R+F GCATTCTTATTGATATTGGAGCTGGCTCTGGAGACTTCCTCAGAACCATCTGAAGAA 720  
 G20M13F+R GCATTCTTATTGATATTGGAGCTGGCTCTGGAGACTTCCTCAGAACCATCTGAAGAA 663  
 17M13F+R  
 RNAM13R+F TGCAATATGAAGACTCCCAATATGAATCCATCTCCTCCCTTCATCCGACTTGACTCTT 780  
 G20M13F+R TGCAATATGAAGACTCCCAATATGAATCCATCTCCTCCCTTCATCCGACTTGACTCTT 723  
 17M13F+R  
 RNAM13R+F -----ATTTGACCACCAATGCTCGTGCTCTCC 585  
 G20M13F+R GAAGAACTCCCTCAGATGCTAACATGTGAAATAGATTGACCACCAATGCTCGTGCTCTCC 840  
 17M13F+R GAAGAACTCCCTCAGATGCTAACATGTGAAATAGATTGACCACCAATGCTCGTGCTCTCC 783  
 \*\*\*\*\*



**Figure 16. Comparison of the nucleotide sequences for confirming the sequences of RT-PCR products.** Partially nucleotide sequences of suspected mature *hsp70* mRNA (RNAM13R+F), intronII-unspliced *hsp70* mRNA (17M13F+R), and intronI+II containing *hsp70* gene (G20M13F+R) were used. The alignment was performed using ClustalW program. Nucleotide sequences of intron I are indicated by a line and intronII are in bold. Primer binding sites are indicated by arrows.

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่  
Copyright © by Chiang Mai University  
All rights reserved