

APPENDIX

APPENDIX A:

Lists of equipments, chemicals, and reagents

Equipments	Source
SpectroMetric®3200	Shimadzu, Japan
Spectrometer 1600	Shimadzu, Japan
PTFE membrane (13 mm diameter, 0.45 µm)	LIDA, Kenosha, WI, USA
Minisart® microsyringe	MS*R250, FUJI, JAPAN
HPLC column (Waters Spherisorb® S5 ODS2) (4.6x10 mm Guard Cartridge)	Water Corporation, USA
Guard column	Water Corporation, USA
Nylon membrane (0.45 µm)	LIDA, Kenosha, WI, USA

Chemicals & Reagents	Source
Acetonitrile (HPLC grade)	Merk, Lichrosolv®, Germany
Ammonium ferrous sulphate	BDH Chemical, Sydney
2,2'-azino-bis-(3-ethylbenzothiazoline- 6-sulfonic acid (ATBS)	Fluka, Switzerland
2,2'-azobis(2-amidinopropane) dihydrochloride	Sigma, St. Louis, MO, USA
Bovine serum albumin (BSA)	Roche Diagnosis, Sydney
Buthionine sulfoximine (BSO)	Sigma, St. Louis, MO, USA
Butylated hydroxytoluene (BHT)	BDH Chemicals, Sydney
Catalase	Roche Diagnosis, Sydney
Diallyl disulfide (purity 90%)	Sigma, St. Louis, MO, USA
Dimethyl sulfoxide (DMSO)	Sigma, St. Louis, MO, USA
Di-potassium hydrogen phosphate (K ₂ HPO ₄)	Merck, Germany
3-(4,5-dimethylthiazol-2-yl)-2,5 -diphenyltetrazolium	Sigma, St. Louis, MO, USA

5,5'-dithiobis-(2-nitrobenzoic acid (DTNB)	Sigma, St. Louis, MO, USA
Folin-Ciocalteu's phenol reagent	Merck, Germany
Gallic acid	Fluka Chemika, Switzerland
Glutathione (GSH)	Sigma, St. Louis, MO, USA
Glutathione reductase (GRx)	Sigma, St. Louis, MO, USA
Guanidine hydrochloride	BDH Chemical, Sydney
Hexane (AR grade)	BDH Chemical, Sydney
Horseradish peroxidase (5,000 U)	Sigma, St. Louis, MO, USA
Hydrogen peroxide (30%, 9.79 M)	CARLO ERBA, Antibiotics SpA,
Immunitytop (aged extract power capsule)	KHAO-LA-OR Laboratory Ltd., Thailand
Kyolic (aged garlic extract power) (NZ)	Nutra-Life Health & Fitness, NZ, Ltd, Auckland, New Zealand
L-glutamine (200 mM)	Trace Scientific, VIC
Liposome from soybean lipid	Provided by Dr. Gebicki's Lab
Lipopolysaccharide (LPS)	Sigma, MO, USA
Methanol (HPLC grade)	Lab-Scan, Thailand
NADPH	Sigma, MO, USA
N-acetylcysteine (NAC)	Fluka, Buchsa, Switzerland
N-(1-Naphthyl)ethylenediamine dihydrochloride	Metheson, Coleman & Bell, Norwood, Ohio
Perchloric acid (PCA) (70-72%)	Merck, Darmstadt, Germany
Phosphoric acid	Sigma, St. Louis, MO, USA
Potassium chloride (KCl)	BDH Chemicals, Sydney
Potassium iodide (KI)	Sigma, St. Louis, MO, USA
Potassium persulfate	Retert Pharmaceutical Co. Island
Quercetin	Sigma, St. Louis, MO, USA
Sodium nitrate	Sigma, St. Louis, MO, USA
Sulfanilamide	Fluka, Buchsa, Switzerland
Tetrahydrofuran (HPLC grade)	Lab-scan, Thailand
Thiazolyl blue (MTT)	Calbiochem, San Diego, CA, USA.
2-Thiobarbituric acid	Sigma, St. Louis, MO, USA

Trichloroacetic acid (TCA)

Aldrick, Milwaukee, MO, USA

Trolox

Sigma, St. Louis, MO, USA.

Xylenol orange, sodium salt

Sigma, St. Louis, MO, USA

Cell line and Media

U937 (monocyte cell line)

Donated by Dr. Gebicki

J774.2 (mouse monocytic-macrophage cell)

Donated by Dr. Usanee

RPMI 1640

Gibco, Island, USA

Fetal calf serum (FCS)

Trace Scientific, VIC

Penicillin/Streptomycin (5,000 IU/ml penicillin G)

Sigma, St. Louis, MO, USA

Culture vessels, serological pipettes and centrifuge tubes

96-well microtitration plates

Greiner, Frickenhausen, Germany

24-well cell culture plate

Costa®, USA

6-well cell culture plate

Costa® USA

60-mm diameter petri dishes

Falcon, NJ, USA

25-cm² flasks with vented cap

Iwaki, Chiba, Japan

75-cm² flasks with vented cap

Iwaki, Chiba, Japan

10 ml sterile plastic pipettes

Sterilin, Staffordshire, UK

50 ml sterile plastic pipettes

Sterilin, Staffordshire, UK

10 ml sterile centrifuge tubes

Iwaki, Chiba, Japan

50 ml sterile centrifuge tubes

Iwaki, Chiba, Japan

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APPENDIX B:

Reagent preparation

1. Reagents for antioxidant activity assays

1.1. ABTS decolorization assay solution

14 mM ABTS solution: 1 g of ABTS dissolved in 100 ml of deionized water (14 mM)

5 mM Potassium persulfate solution; 0.007g of potassium persulfate dissolved in 2 ml of deionized water; mixed ABTS and persulfate solution, kept in dark for 12-16 h at room temperature. The stock should be used within 3-5 days.

4.7 mM Standard gallic acid solution: 2 mg of gallic acid was dissolved in 100 ml of 40% ethanol.

5 mM Standard Trolox: 0.125 g dissolved in 100 ml of ethanol

1.5 mM Standard BHT: 0.033 g dissolved in 100 ml of deionized water.

1.2. H₂O₂ scavenging assay solution

0.1 % ABTS: 0.1 g of ABTS was added in 100 ml of distilled water and keep in cool temperature at 4°C.

Peroxidase (10 U/ml): Diluted Horseradish peroxides (HRP) (5,000 U) with Phosphate buffer (pH 6.0)

H₂O₂ (0.003%): diluted H₂O₂ from stock H₂O₂ (30%) with deionized water

2. Reagents for protein and lipid hydroperoxide assays

2.1. Tri-iodide assay solution

10% Potassium iodide (PI) solution: 20 mg of PI dissolved in 200 ml of acetic acid solution (50%) in anaerobic system.

2.2. Ferrous oxidation-xylenol orange (FOX) assay solution

5 mM XO: 5 mM xylenol orange (XO) was made up in 110 mM HClO₄ and stored at room temperature.

5 mM Ferrous solution (Fe^{2+}): 5 mM ammonium ferrous sulphate was made up in 110 mM HClO_4 and stored at 4°C .

8 M guanidine HCl: this was made up in Milli Q water.

Methanol (20% BHT): 20 mg BHT was added in 100 ml of methanol (AR grade)

Chloroform (20% BHT): 20 mg BHT was added in 100 ml of Chloroform AR grade)

3. Reagents for glutathione and malondialdehyde test in blood

3.1. AAPH solution

10 mM AAPH solution: working AAPH solution at 10 mM was prepared by dissolving 0.271 g of AAPH in 100 ml of phosphate buffer (pH 7.4). Fresh preparation of AAPH was used for experiments and was kept in -80°C until assay.

3.2. Erythrocyte glutathione assay solution

DTNB reagent: 40 mg of DTNB was dissolved in 100 ml of distilled water

Precipitating agent: 0.2 g of EDTA: 1.67 g of meta-phosphoric acid and 30 g of sodium chloride in 100 ml of distilled water

Phosphate buffer: 0.3 M Na_2HPO_4 was prepared in distilled water and adjusted pH to 7.4

Glutathione standard: dissolved stock GSH at 10 mg/100 ml in distilled water.

3.3. Plasma malondialdehyde assay

TBA reagent: 4% TBA was prepared by dissolving 0.4 gram of TBA in distilled water 100 ml and kept at 4°C .

TCA reagent: 100 % of TCA was prepared by 250 g of TCA dissolved in 250 ml of 6 M HCl.

Malondialdehyde standard: The standard malondialdehyde at 100 mM was prepared by taking 20 μl from stock MDA and neutralized with 5-8 drop of concentrated HCl before being dissolved in distilled water to 100 μM .

4. Reagents for identification of bioactive compounds in HPLC

4.1. Phenolic analysis

Folin-Ciocalteu's phenol reagent

20% Na_2CO_3 was prepared by weighting Na_2CO_3 20 g and dissolving in distilled water 100 ml

Standard gallic acid; 20 mg of gallic acid was dissolved in 25 ml 40% ethanol

4.2. Diallyl disulfide analysis

Mobile phase solution: mixed acetonitrile, water and tetrahydrofuran in 70/27/3 ratio (v:v:v) and degassed with vacuum equipment.

4.3. Allyl-2 propenethiosulfinate (allicin) analysis

Mobile phase solution was 50% methanol in distilled water and degassed with vacuum equipment.

5. Reagents for oxidative stress in U937 cells line

5.1. Cell culture media

RPMI 1640 (without L-glutamate) was supplemented with 2 mM L-glutamate, 10% (v:v) heat-inactivated FCS and 100 units/ml penicillin, 100 $\mu\text{g}/\text{ml}$ streptomycin. The pH value of the medium was adjusted to 7.4 with 7.5% NaHCO_3 . All solutions were handled under sterile conditions in a sterile laminar flow hood and stored at 4 °C.

Phosphate-buffer saline (PBS): 137 mM NaCl, 2.7mM KCl, 1.5 mM KH_2PO_4 , 10 mM Na_2HPO_4 , pH 7.4 was sterilized by autoclaving and stored at room temperature.

5.2. MTT assay for cell survival assay

MTT stock solutions: MTT was dissolved in PBS at 5 mg/ml, filter sterilized and stored in the dark at 4 °C. MTT stock solutions were discarded after one month.

5.3. Intracellular glutathione assay

Phosphate-EDTA buffer: 100 mM sodium phosphate, 5 mM EDTA, pH 7.5. It was stored at room temperature

DTNB stock solution: 4 mg/ml in phosphate-EDTA buffer. It was stored at 4 °C and discarded after two weeks

NADPH stock solution: 4 mg/ml in phosphate-EDTA buffer. It was stored at 4 °C and was discarded after two weeks.

3% PCA (v:v) solution: diluted from stock 70% PCA with distilled water.

GSH solution was prepared immediately before use in cold 3% PCA solution

5.4. Griess reagent

1% sulfanilamide solution: 1 g of sulfanilamide in 100 ml of 5% phosphoric acid

0.1% NED solution: 0.1 g N-1-naphthylethylenediamine dihydrochloride in 100 ml of distilled water

Standard nitrate (0.1 M): 0.069 g of NaNO₂ in 10 ml of distilled water.

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APPENDIX C:

**The initial rates of reduction of standard antioxidant gallic acid
and BHT**

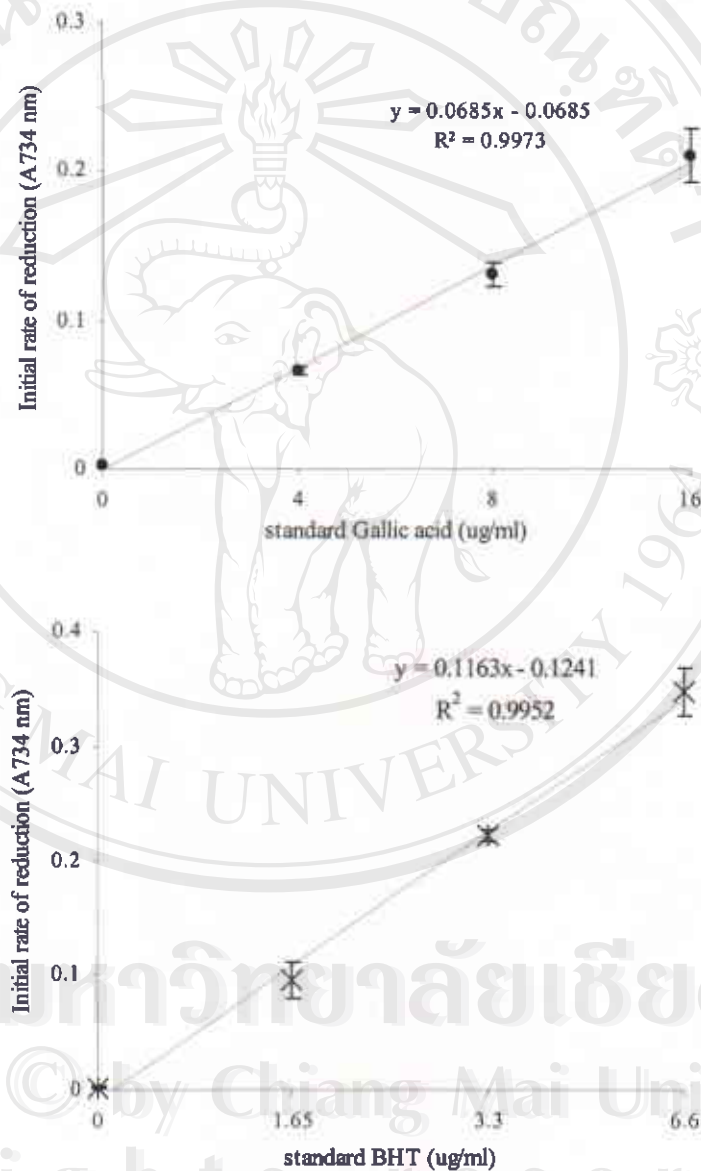


Figure 72. Antioxidant activity from initial rates of reduction in ABTS cation radicals of standard gallic acid and BHT. Initial rate of reduction (A734 nm) of standard antioxidants was determined with ABTS radical reduction within one minute. Each point represents the mean and standard deviation from triplicate trials.

APPENDIX D:

The percentage of cell survival from standard diallyl disulfide (DADS) in U937 cell line

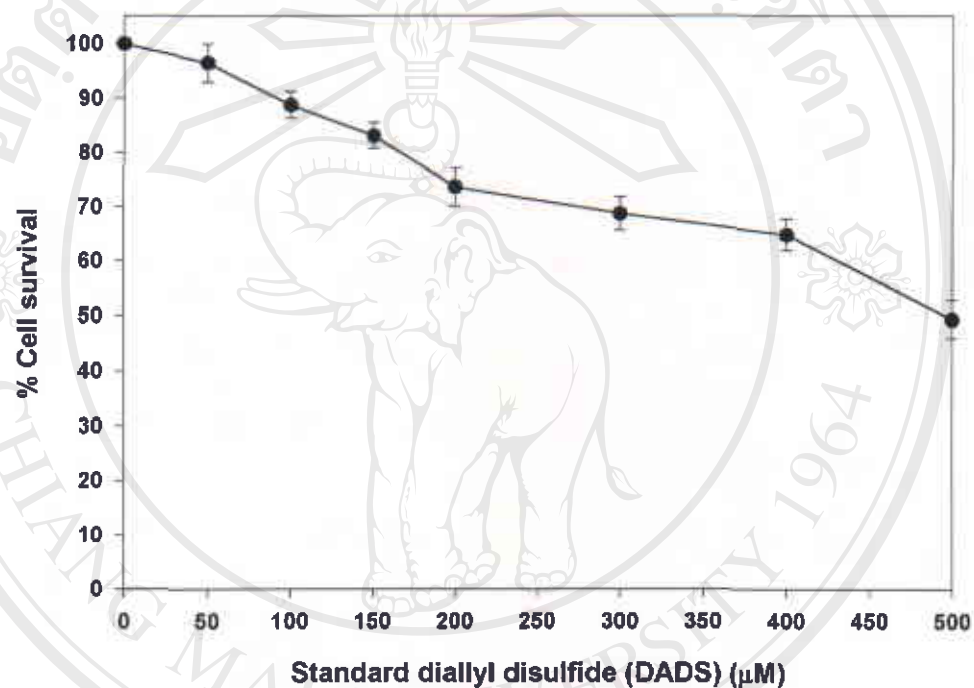


Figure 73. Effect of diallyl disulfide (DADS) on U933 cell survival. Percentage of cell survival was evaluated with MTT dye in U937 (5×10^3 cells/well) treated with standard DADS at 0-500 $\mu\text{g/ml}$ and incubation for 24 hr at 37°C , 5% CO_2 . Each point represents the mean and standard deviation from triplicate trials.

APPENDIX E:

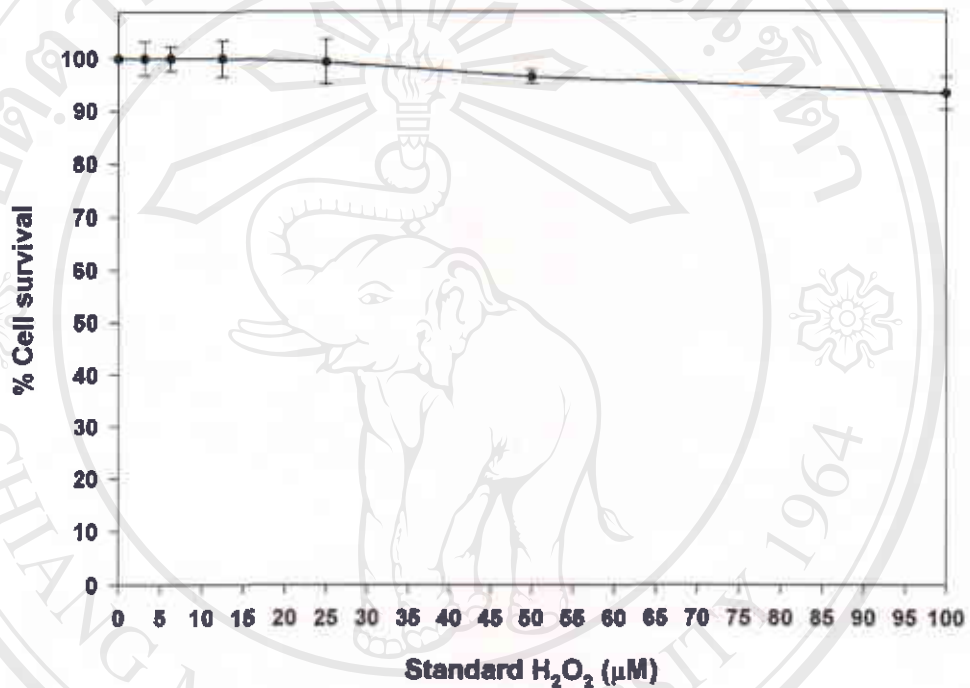
The cell survival of H₂O₂-activated U937 cell line

Figure 74. Effect of H₂O₂ on U937 cell survival. Percentage of cell survival was evaluated with MTT dye in U937 (5x10³ cells/well). The cells were treated with H₂O₂ at 0-100 µM and incubation for 24 hr at 37°C, 5% CO₂. Each point represents the mean and standard deviation from triplicate trials.

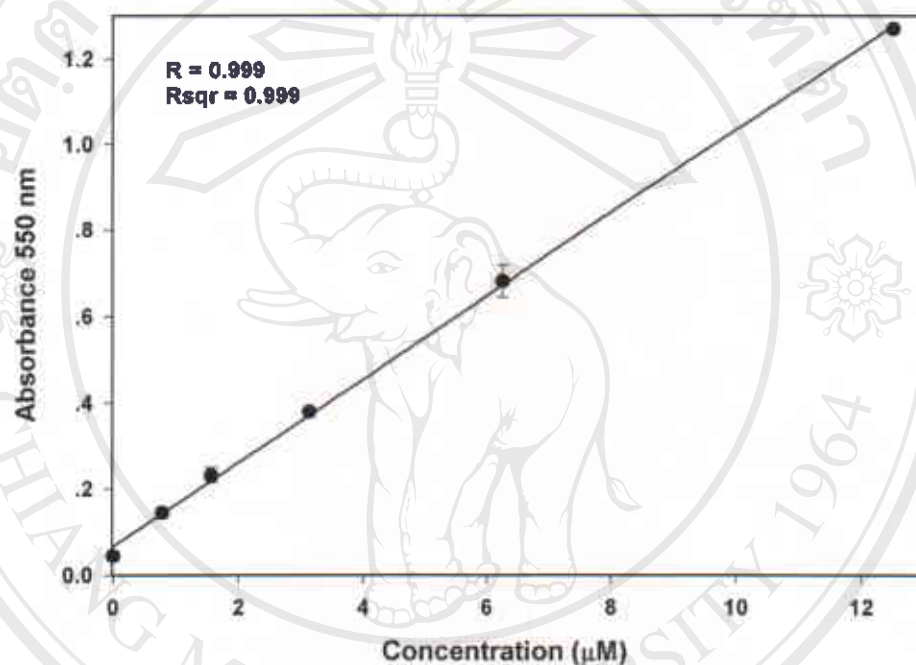
APPENDIX F:**The standard curve of sodium nitrite.**

Figure 75. Standard curve of sodium nitrite. Nitrite levels was evaluated with Griess reagent in 96-well plate at 0-12 µM. The stock working sodium nitrate (100 µl) was mixed with 100 µl of NED and Sulfanilamide, the pink color was generated and read at 550 nm. Each point represents the mean and standard deviation from triplicate samples.

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APPENDIX G:

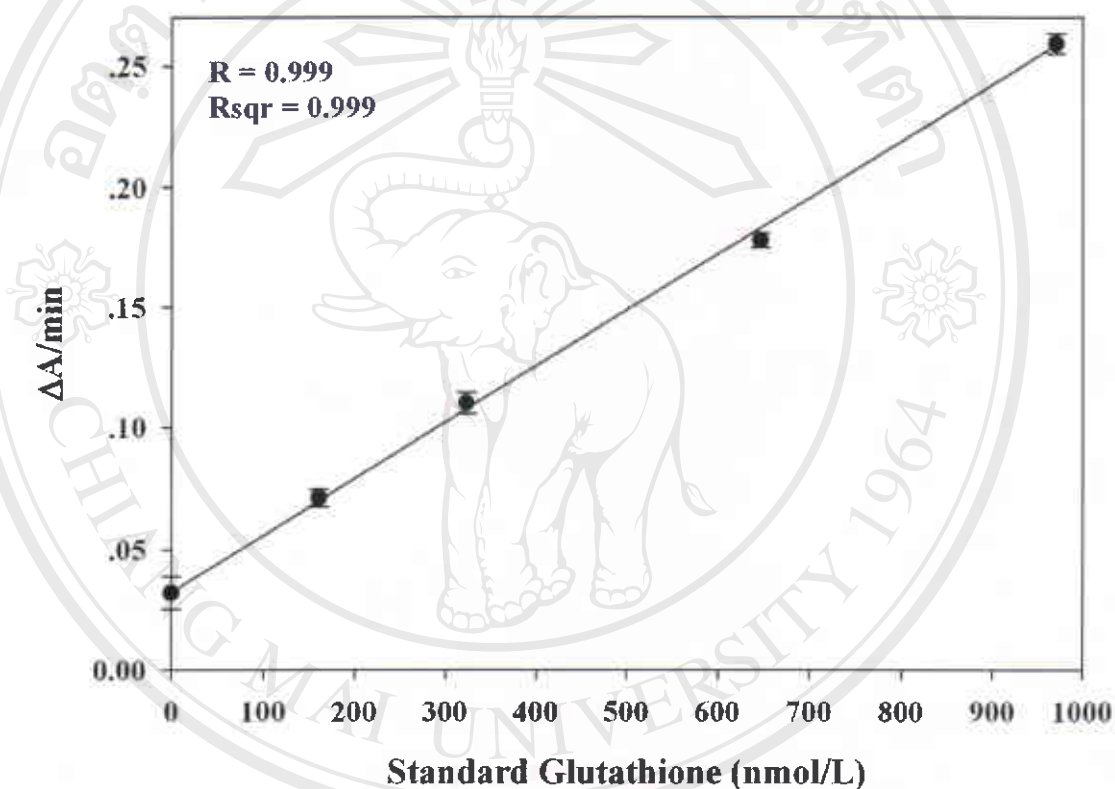
The initial rate of reduction ($\Delta A/\text{min}$) of standard glutathione

Figure 76. The initial rate of reduction of standard glutathione. The initial rate of reduction on ABTS cation radicals (734 nm) was evaluated with kinetic program. The glutathione at 0-1,000 nmol/L was prepared in 3% PCA and mixed with NADPH (4 mg/ml) and glutathione reductase. The yellow color production was recorded within 3 minutes and the initial rate of reduction ($\Delta A/\text{min}$) was used. Each point represents the mean and standard deviation from triplicate samples.

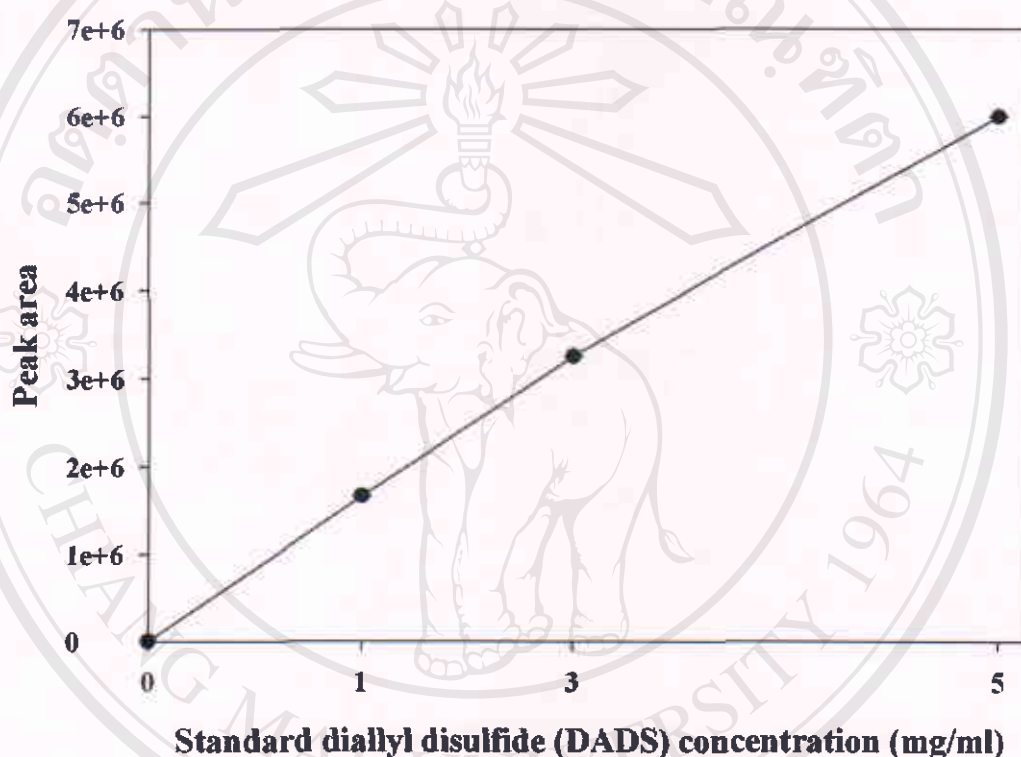
APPENDIX H:**The standard curve of diallyl disulfide (DADS)**

Figure 77. Standard curve of diallyl disulfide (DADS). The peak area of standard of DADS at 0-5.0 mg/ml are shown from HPLC chromatogram. Results from a UV detector at 240 nm at flow rate 1.0 ml/min (0.10 AUFS) eluted by acetonitrile:water:tetrahydrofuran mixture(70:23:3, v:v:v) as a mobile phase. Each point represents the mean and standard deviation from triplicate samples.

APPENDIX I:

The peaks of organosulfur compounds of standard diallyl disulfide (DADS) from gas chromatography-mass spectrum (GC-MS) analysis.

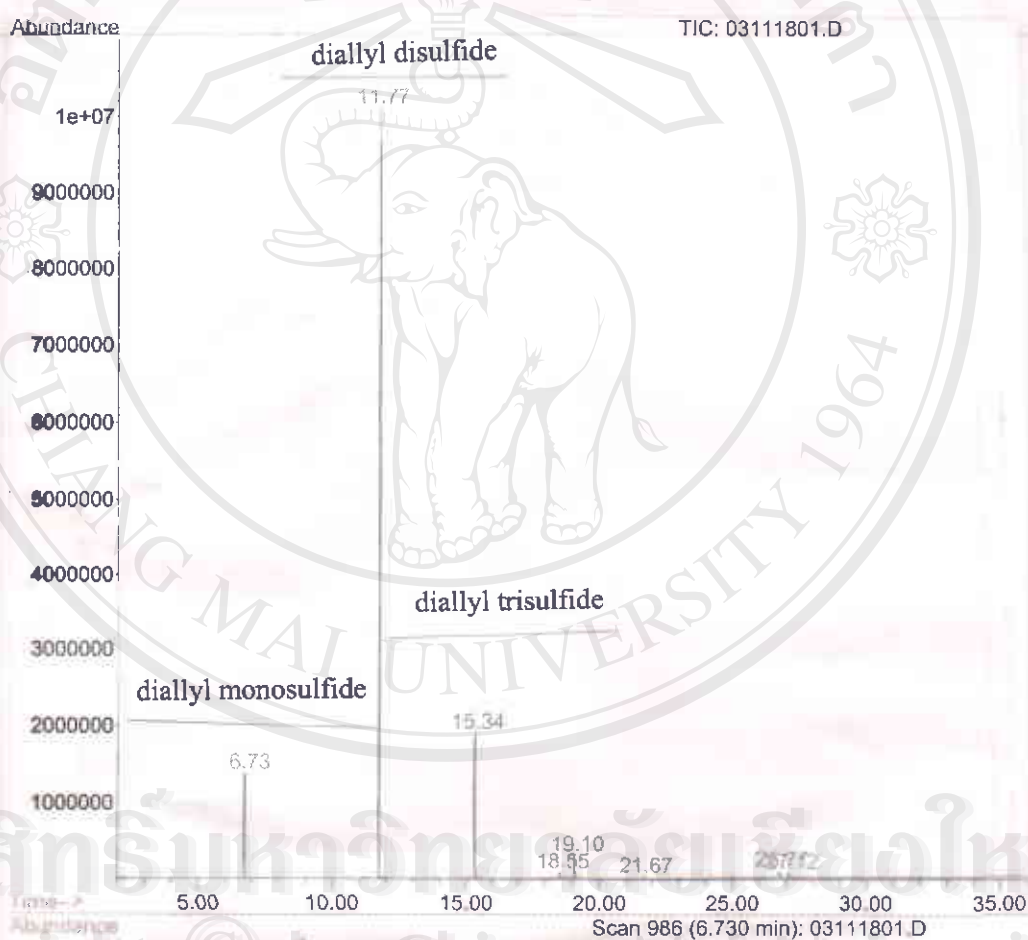


Figure 78. GC-MS chromatogram of standard diallyl disulfide (DADS). Three peaks of allyl sulfides; mono- (6.73 min), di- (11.77 min), and tri-sulfide (15.34 min) were identified with the Gases chromatography- mass spectrum (GC-MS). The 20 μ l of standard DADS was taken in vial for GC-MS analysis.

Abundance

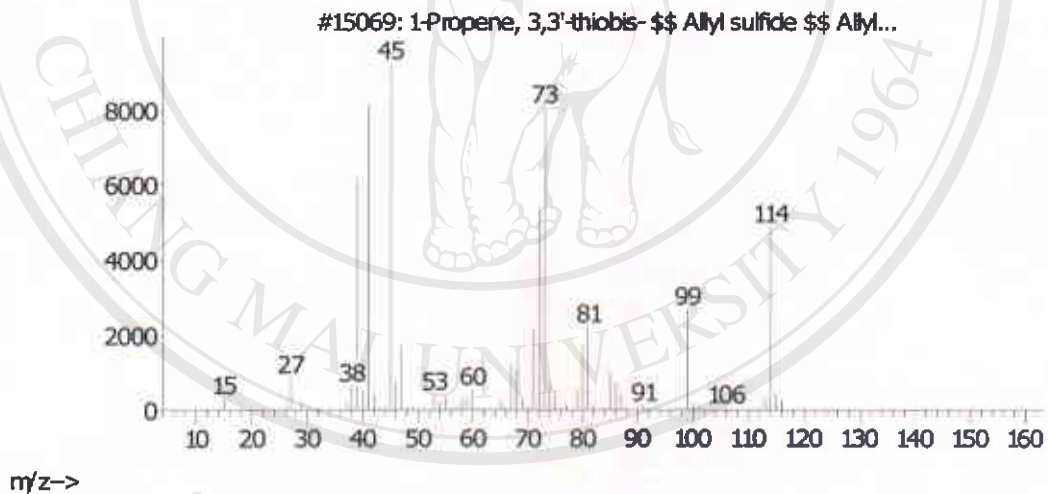
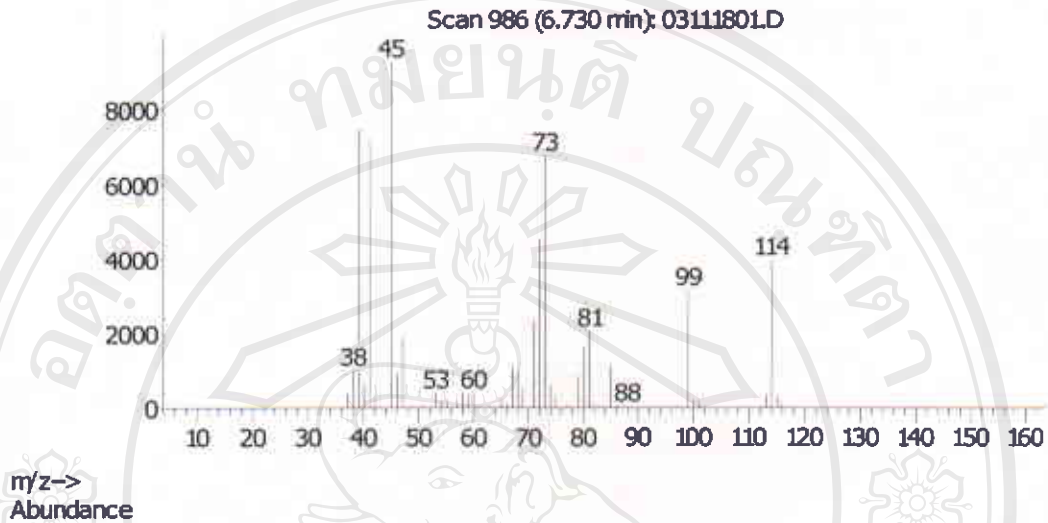
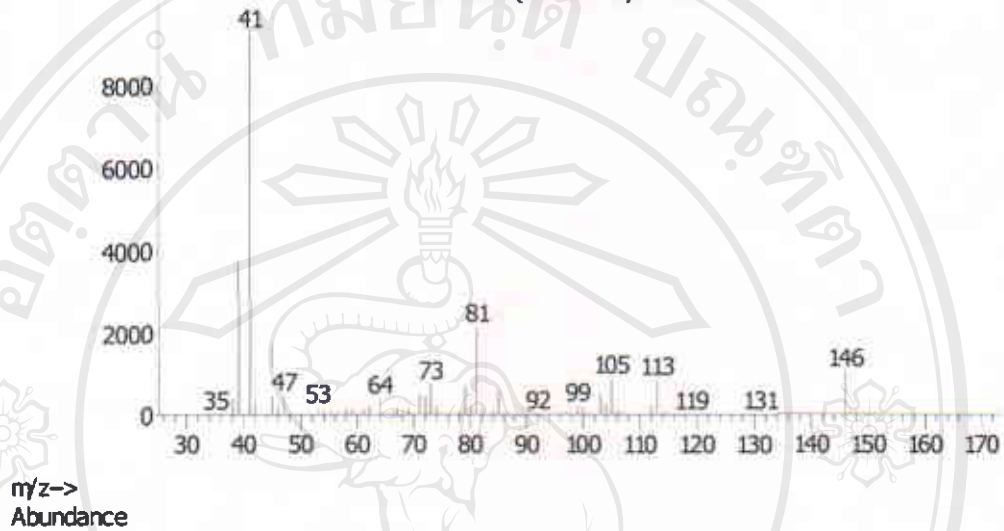


Figure 79. GC-MS chromatogram and mass spectrum of diallyl monosulfide. Mass spectra of diallyl monosulfide at 6.73 min from standard diallyl disulfide (DADS) (Sigma) that identified with the Gases chromatography- mass spectrum (GC-MS). The 20 μ l of standard DADS was taken in vial for GC-MS analysis.

Abundance

Scan 2034 (11.769 min): 03111801.D



#41859: Diallyl disulphide \$\$ Allyl disulfide \$\$ Diallyl d...

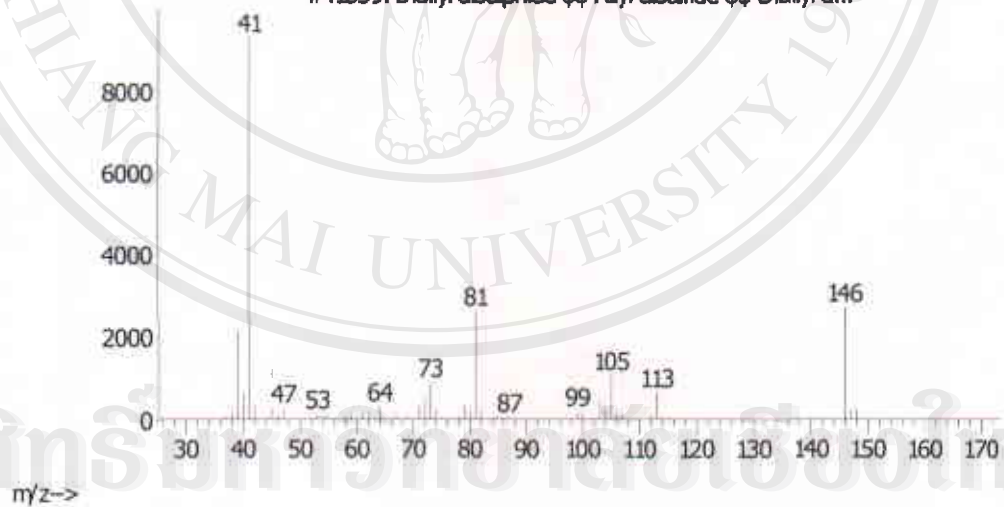
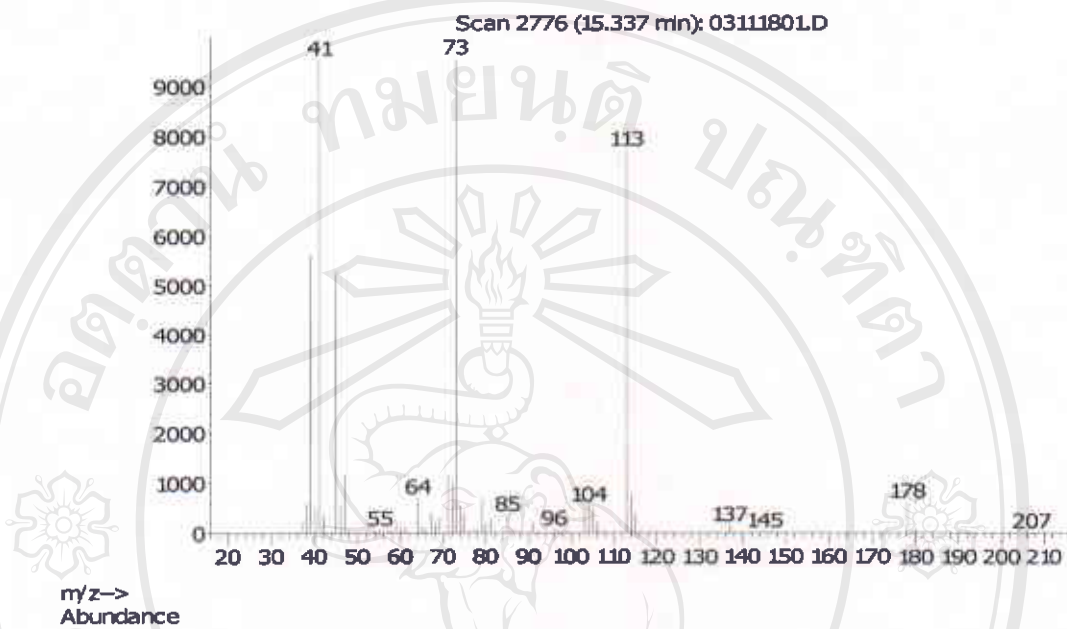


Figure 80. GC-MS chromatogram and mass spectra of diallyl disulfide (DADS). Mass spectra of diallyl disulfide at 11.76 min from standard diallyl disulfide (DADS) (Sigma) that identified with the Gases chromatography- mass spectrum (GC-MS). The 20 μ l of standard DADS was taken in vial for GC-MS analysis.

Abundance



#82586: Trisulfide, di-2-propenyl \$\$ Allyl trisulfide \$\$ D...

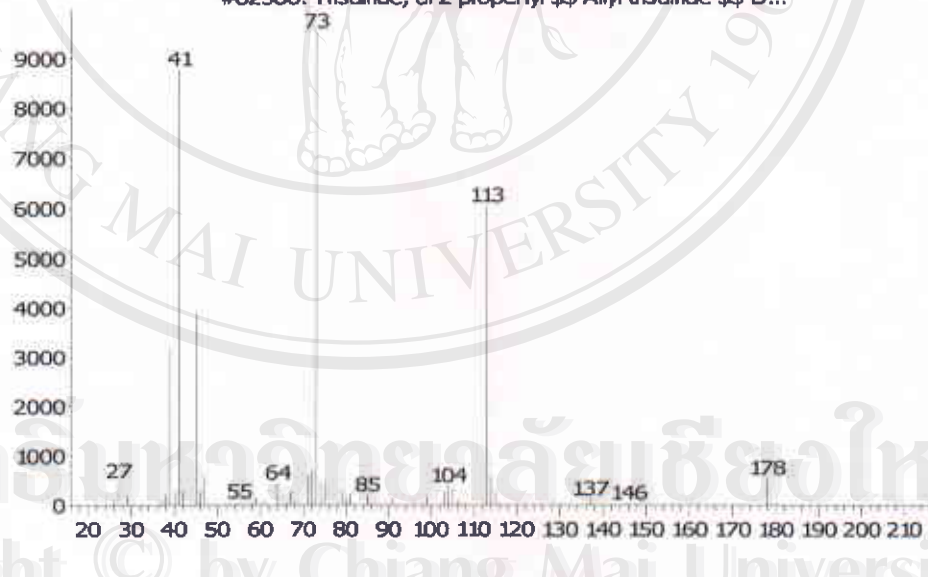


Figure 81. GC-MS chromatogram and mass spectra of diallyl trisulfide (DATS). Mass spectra of diallyl trisulfide at 15.33 min from standard diallyl disulfide (DADS) (Sigma) that identified with the Gases chromatography- mass spectrum (GC-MS). The 20 μ l of standard DADS was taken in vial for GC-MS analysis.

Pk#	RT	Area%	Library/ID	Ref#	CAS#	Qual
1	6.73	11.03	C:\DATABASE\WILEY275.L 1-Propene, 3,3'-thiobis- (CAS) \$\$	11963	000592-88-1	98
			1-Propene, 3,3'-thiobis- (CAS) \$\$	11964	000592-88-1	98
			1-Propene, 3,3'-thiobis- (CAS) \$\$	11965	000592-88-1	97
2	11.77	73.61	C:\DATABASE\WILEY275.L Disulfide, di-2-propenyl (CAS) \$\$	32189	002179-57-9	78
			Disulfide, di-2-propenyl (CAS) \$\$	32191	002179-57-9	30
			Disulfide, di-2-propenyl (CAS) \$\$	32193	002179-57-9	10
3	15.34	10.86	C:\DATABASE\WILEY275.L Trisulfide, di-2-propenyl (CAS) \$\$	61720	002050-87-5	96
			Trisulfide, di-2-propenyl (CAS) \$\$	61719	002050-87-5	64
			1-Propene, 3,3'-thiobis- (CAS) \$\$	11965	000592-88-1	43
4	18.55	0.45	C:\DATABASE\WILEY275.L Disulfide, di-2-propenyl (CAS) \$\$	32191	002179-57-9	38
			Disulfide, di-2-propenyl (CAS) \$\$	32188	002179-57-9	22
			Zinc, di-2-propenyl- (CAS) \$\$ ZINC	32206	001802-55-7	14
5	19.10	2.35	C:\DATABASE\WILEY275.L 1,2-Benzenedicarboxylic acid, diet	107342	000084-66-2	98
			1,2-Benzenedicarboxylic acid, diet	107338	000084-66-2	98
			1,2-Benzenedicarboxylic acid, diet	107340	000084-66-2	97
6	21.68	0.22	C:\DATABASE\WILEY275.L bis-TMS-C(13)-3-hydroxybutanoate \$	132496	000000-00-0	42
			Ethanedioic acid, bis(trimethylsil	119112	018294-04-7	38
			1,3-Dioxolane (CAS) \$\$ 1,3-Dioxola	1725	000646-06-0	27
7	26.77	0.93	C:\DATABASE\WILEY275.L 9-Octadecenamide, (Z)- (CAS) \$\$ OL	162712	000301-02-0	86
			9-Octadecenamide, (Z)- (CAS) \$\$ OL	162711	000301-02-0	62
			9-Octadecenamide, (Z)- (CAS) \$\$ OL	162710	000301-02-0	60
8	27.12	0.55	C:\DATABASE\WILEY275.L Hexanedioic acid, dioctyl ester (C	222005	000123-79-5	87
			Hexanedioic acid, dioctyl ester (C	222002	000123-79-5	55
			Di(2-ethylhexyl)adipate	2220A1	000103-23-1	49

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Figure 82. Comparative peaks and retention time of each diallyl sulfides. The peaks and retention time of each diallyl sulfide were used to analysis the chemical structure and scientific name by using specific of mass spectrum in library database (Wiley275.L).

CURRICULUM VITAE

Name Mr. Nuttkaan Leelarungrayub

Official address

Department of Physical Therapy, Faculty of Associated Medical Sciences,
Chiang Mai University, Chiang Mai, Thailand.

Tel; +66-53-94-9245. Fax; +66-53-94-6042

E-mail; nuttkaan@chiangmai.ac.th

Education 2544: MSc (Biochemistry) Chiang Mai University, Thailand

2536: BSc (Physical Therapy) Khonkaen University, Thailand

Research projects

Study of phenolic compound and total antioxidant capacity in fermented juices from Tomato, Carrot and Gum. Granted by Faculty of AMS (2006)

Study of response of hydroperoxide formation in blood from high intensity exercise between athletics and sedentary controls. Granted by Faculty of AMS (2005)

Study the effects of Thai shallot on total intracellular glutathione synthesis in oxidative stress and inflammation in human monocytic cell line (U937). Granted by SM.Pharmaceutical.Co (Thailand) (2002-2004)

Tendency of native shallot preparation for protective and inhibitory effects on oxidative stress in erythrocyte as commerce products. Granted by NSTDA (Northern Network), Thailand (2000-2001)

Comparative study of derivated N-acetylcysteine drugs on oxidative stress in protein and lipid hydroperoxide formation in vitro. Granted by SM.Pharmaceutical.Co. (Thailand) (1999-2002)

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Publications

A quantitative evaluation of the antioxidant properties of garlic and shallot preparation. *Nutrition*. 2006; 22: 266-274.

Exhaustive Exercise Test and Oxidative Stress Response in Athletic and Sedentary subjects. *Chiang Mai University Journal* . 2005; 4: 183-190.

Potential activity of Thai shallot (*Allium ascallonicum* L.) extract on the prevention of hemolysis and glutathione depletion in human erythrocyte from oxidative stress. . *Chiang Mai University Journal*. 2004; 3: 225-234.

Aerosol Therapy and Chest Physical Therapy on Oxidative Stress in blood and tracheal aspirate fluid (TAF) of pediatric patients with pneumonia. *Bull Chiang Mai Assoc Med Sci*. 2005; 38: 160-172.

Anti-oxidative stress and anti-inflammatory effects of Thai shallot (*Allium ascanolicum* L.) extracts in human monocytic (U937) cells. . *The Niigata Journal of Health and Welfare*. 2004; 4: 11-19.

Free radical and Exercise Tolerance in Chronic Obstructive Pulmonary Disease: COPD . *Bull Chiang Mai Assoc Med Sci* 2000;33: 212-220.

Antioxidant capacity of Thai shallot (*Allium ascalonicum* L.) on amino acid and glutathione from oxidation in vitro . *Bull Chiang Mai Assoc Med Sci*. 2004; 37: 27-36.

Chest Physical Therapy and ventilation-gas exchanges impairment. . *Bull Chiang Mai Assoc Med Sci*. 2004; 38: 40-50.

Correlation between Malondialdehyde (MDA), Hyaluronan (HA), and Alpha-tocopherol (Vit E) in Tracheal Aspirate Fluid (TAF) and Oxygen Index (PaO₂/FiO₂) in Pediatric Patients with Chronic Lung Disease . *Bull Chiang Mai Assoc Med Sci*. 2003; 36: 24-34.

Determination of GSH levels in Red Blood Cell with Dithiobis-nitrobenzoic acid method in normal and cerebrovascular disease patients. *Bull Chiang Mai Assoc Med Sci* . 2001; 34: 12-21.

Chest Physical Therapy Techniques. *Bull Chiang Mai Assoc Med Sci* 2000; 33: 29-40.

Free radical and Exercise Tolerance in Chronic Obstructive Pulmonary Disease: COPD. *Bull Chiang Mai Assoc Med Sci*. 2000; 33: 212-220.

A case study; Therapeutic effects of physical therapy in a COPD patient on weaning from ventilator . Bull Chiang Mai Assoc Med Sci. 1999; 32: 31-42.

Abnormal pulmonary assessment in pediatric patients. Bull Chiang Mai Assoc Med Sci . 1999; 32: 25-30.

A new approach of respiratory physical therapy in a patient with empyema thoracis after decortication . Bull Chiang Mai Assoc Med Sci 1998; 31: 199-206.

Biochemistry and respiratory disorder in chest physical therapy. Bull Chiang Mai Assoc Med Sci . 1998; 31: 42-56.

Exercise and Glutathione under Oxidative Stress. Bull Chiang Mai Assoc Med Sci. 1998; 31: 225-230.

Respiratory Muscles Exercise in Chest Physical Therapy. . Bull Chiang Mai Assoc Med Sci. 1997; 30: 44-56.

Chest Physical Therapy for Abdominal Surgical Patients. . Bull Chiang Mai Assoc Med Sci 1996. 1996; 29: 19-29.

Exercise and Lung Disease. . Nursing Newsletter. 1996; 23: 26-33.

Lung atelectasis and Chest Physical Therapy. Bull Chiang Mai Assoc Med Sci. 1996; 30: 77-90.

Books

Chest radiography for Chest Physical Therapy (1997)

Chest Physical Therapy; Part II (2002)

Academic Conference

The effectiveness of Thai shallot (*Allium ascanolicum* L.) and phenolics compounds against protein and lipid peroxidation. International Colloquium 2004 Health Benefit and Application of Polyphenoids. Faculty of Associated Medical Sciences, Chiang Mai University, Chiang Mai, Thailand, 25 - 26 November 2004.

Effects of Thai shallot (*Allium ascanolicum* L.) extracts on oxidative stress under intracellular glutathione and nitric oxide. International Colloquium 2004 Health Benefit and Application of Polyphenoids. Faculty of Associated Medical Sciences, Chiang Mai University, Chiang Mai, Thailand, 24 - 26 November 2004.

Anti-oxidative stress and anti-inflammatory effects of Thai shallot (*Allium ascalonicum* L.) extracts in human monocytic, . The 28th Annual Scientific Meeting on Mahidol' s Day, Faculty of Medicine, Chiang Mai University, Thailand, 24 September 2004.

A comparative study N-acetylcysteine-derived drug on scavenging free radical and hydroperoxide formation in vitro. The 4th International World Asthma Meeting, Bangkok, Thailand. Granted by S.M Pharmaceutical Co. th., 16 - 19 February 2004.

Anti-oxidative stress and Anti-inflammatory effects of Thai shallot (*Allium ascalonicum* L.) extracts in human monocytic . The 4th National symposium on graduate research. August 10-11, 2004. Lotus Hotel Pang Suan Kaew Chiang Mai, Chiang Mai, Thailand, 10 - 11 August 2004.

The relationship between gross motor function measurement score and glutathione and malondialdehyde level in children with CP. The 14th International World Physical Therapy, 7-12 June, 2003; Barcelona, Spain. 2003 Granted by Faculty of Associated Medical Sciences, Chiang Mai, Thailand. (RR-PO- 0377), 7 - 12 June 2003.

Antioxidative effect of Thai shallot (*Allium ascalonicum* L.) extracts on protein and lipid hydroperoxides. The 27th Annual Scientific Meeting on Mahidol's Day, Faculty of Medicine, Chiang Mai University, Thailand, 25 September 2003.

Effects of chest physical on biochemical changes in pediatric patient with pneumonia. The 8th General Assembly of Asian Confederation for Physical therapy, Central Grand Plaza Hotel, Bangkok, 17 - 20 November 2002.

Exhaustive exercise test and oxidative stress response to athletic and sedentary subjects. 29th Annual Scientific Meeting on Mahidol's Day, Faculty of Medicine. Chiang Mai University, Chiang Mai, Thailand, 23 September, 2548, pp 28-28.

Protein hydroperoxide relating to glutathione and lung function: A preliminary study. 29th Annual Scientific Meeting on Mahidol's Day. Faculty of Medical, Chiang Mai University, Thailand, 23 September, 2548, pp 28-28.

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Training course

25th November, 2004, Microarray Solutions for Genomics and the New Technology of Nucleic Acid Extraction with Higher Yield and Purity, Chiangmai, Thailand

28th Febuary, 2004. Tea Research for Health and Prevention of Disease, Faculty of Medicine, Chiang Mai University, Chiang Mai, Thailand.

25th-26th, November, 2003. International colloquium 2004; On Health Benefits and Applications of Polyphenols , Faculty of Associated Medical Sciences, Chiang Mai University, Chiang Mai , Thailand.

November 2003 - April 2004. Protein and Lipid hydroperoxide Assay Techniques with FOX and Tri-iodide Assays , Free radical and Biochemistry Laboratory, Department of Biological Sciences, Macquarie University, NSW , Australia



ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่

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