

## **APPENDICES**

### **APPENDIX A**

#### **EQUIPMENT, MATERIALS, MEDIA AND REAGENTS**

##### **1. Lab Equipments and Materials**

- Sterile 500, 1000 and 2000 ml Erlenmeyer flasks, sterile 250 and 500 ml beakers, and containers of capacity to accommodate samples
- Balance with a 2000 g-weights capacity and a sensitivity of 0.1 g
- Incubator, 37 and 42 C
- Laboratory refrigerator, - 20 C and -1 to 4+ C
- Water bath
- Sterile spoons for transferring faecal samples and media
- Sterile culture dishes, 15\*100 mm, glass or plastic
- Sterile pipettes
- Inoculating needle and inoculating loop (10 micrometer)
- Culture tubes, 16\*150 and 20\*150 m
- Test or culture tube racks
- Vortex mixer
- Stomacher machine.
- Sterile scissors, scalpel, and forceps
- Bunsen burner
- Stomacher bags and plastic bags
- Appendop
- Autoclave

## 2. Equipment and Material for Sample Collection

- Sterile cotton sock swabs
- Disposable hand gloves
- Stomacher bags and plastic bags
- Buffered peptone water (BPW)
- Sterile 1000 ml. Duran bottle
- Marker pens
- Alcohol, cotton, lighter
- Normal saline
- Disposal gloves, boots and lab coat
- Ice box with ice
- Snare

## 3. Media, Reagent and Chemicals

- Buffered Peptone Water (BPW)
- Nutrient agar (NA)
- Brilliant-green Phenol-red Lactose Sucrose Agar (BPLS)
- Xylose Lysine Tergitol 4 agar (XLT4)
- Muller Kaufmann Tetrathionate broth (MKTT)
- Rppaport-Vassiliadis broth (RV)
- Triple Sugar Iron Agar (TSI)
- Urea Agar
- Motility Indole Lysine Decarboxylation (MIL)
- Voges-roskauer Reaction (VPR)
- *Salmonella* polyvalent somatic (O) antiserum A- E
- *Salmonella* polyvalent somatic (O) antiserum F- 67
- *Salmonella* somatic (O) antiserum- *Salmonella* group B (O4, O5 , O27)
- *Salmonella* somatic (O) antiserum- *Salmonella* group C (O7, O8)
- *Salmonella* somatic (O) antiserum- *Salmonella* group D (O9, Vi)
- *Salmonella* somatic (O) antiserum - *Salmonella* group E (O3, O19)

- Anti- *Salmonella* flagella (H) e.g. e, f, g, h, i, k, l, m, p, q, r, s, t, u, v, w, x,  
z4, z23, z6, z29, z32, 1, 2, 5, 6, 7



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## APPENDIX B

### MEDIA PREPARATION

#### **Buffered Peptone Water (BPW; Merck KGaA, Germany)**

Preparation: Suspend 25.5 g in 1 liter of demineralized water; if desired dispense into smaller vessels; autoclave (15 min. at 121 °C). pH:  $7.0 \pm 0.2$  at 25 °C

#### **Modified Semi Solid Rappaport Visiliadis broth (MSRV; Merck KGaA, Germany)**

Preparation: Suspend 42.5 g in 1 liter of autoclave water; **Do not** autoclave this medium, boiling, prior to use added MSRV supplement after that distributed into sterile Petri dishes 10 ml.

#### **Tetrathionate broth (TT; Merck KGaA, Germany)**

Preparation: Suspend 82 g in 1 liter of demineralized water, heat briefly to boiling. Do not autoclave! After cooling, add 20 ml/l iodine potassium iodine solution and 10- ml/l 0.1% brilliant green solution (brilliant green, Cat. No.1.01310.). Dispense any eventual precipitate.

#### **Xylose-lysine-tergitol 4 (XLT4; Merck KGaA, Germany)**

Preparation: Suspend 59 g in 1 liter of demin. water, add 4.6 ml XLT4 Agar Supplement solution and heat the medium in a boiling water-batch (*not* on a heatingplate!). Cool to approx. 50 °C and pour plates. Do not overheat, do not autoclave! pH:  $7.4 \pm 0.2$  at 25 °.

**BPLS (BPLS; Merck KGaA, Germany)**

Preparation: Suspend 51 g in 1 liter of demineralized water; boiling, autoclave (15 min. at 121 °C) pH:  $7.0 \pm 0.2$  at 25 °C, plating.

**Triple Sugar Iron Agar Slants (TSI; Merck KGaA, Germany)**

Preparation: Suspend 65 g in 1 liter of demin. water by heating in a boiling water bath or in a current of stream; dispense into tubes; autoclave (15 min. at 121°C). Allow solidifying to give agar slants. pH:  $7.4 \pm 0.2$  at 25 °C

**Urea Agar (Urea; Merck KGaA, Germany)**

1. Suspend 29 g of medium in 100 ml of demineralized water.
2. Mix thoroughly and sterilize by filtration.
3. Dissolve 15 g of agar in 900 ml of demineralized water.
4. Sterilize by autoclaving at 121°C for 15 minutes.
5. Cool to 45-50°C and aseptically add the sterile Urea Agar Base.
6. Mix thoroughly and dispense into sterile tubes.
7. Cool in a slanted position so that deep butts are formed.

**MIL (MIL; Merck KGaA, Germany)**

Preparation: Suspend **20 g in 1 liter** of demineralized water by heating in a boiling water bath or in a current of stream; autoclave (15 min. at 121 °C). Pour to plates. pH:  $7.0 \pm 0.2$  at 25 °C

**Nutrient Agar (NA; Merck KGaA, Germany)**

Preparation: Suspend **20 g in 1 liter** of demineralized water by heating in a boiling water bath or in a current of stream; autoclave (15 min. at 121 °C). Pour to plates. pH:  $7.0 \pm 0.2$  at 25 °C

**APPENDIX C**  
**COMPARISON OF *SALMONELLA* SEROTYPES IN THIS STUDY**  
**WITH THE OTHER STUDIES**

Serogroup	Serotypes	Percentage		
		This study	Dorn-in (2005)	Sangwatanakul (2007)
B	<i>S. Stanley</i>	14.28	11.5	0
	<i>S. Typhimurium</i>	-	18.3	6.06
C	<i>S. Rissen</i>	57.14	45.4	18.18
E	-	-	14.6	-
D	-	-	2.0	-
II	F-67	25 (2/8)	3.7	75.76

## DECLARATION

I, the under signed, declare that the thesis is my original work and has not been presented for a degree in any University.

Name

Ruttayaporn Ngasaman

Signature.....

Date of submission.....

25 Sep 07

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## CURRICULUM VITAE

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### Educational background

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- 1991-1994 High School from Pimanpityasan School, Satun, Thailand
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### Awards and scholarships

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- CMU Scholarship student supported by the Richard Hau Global Foundation, 1999-2004
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