



APPENDIX A
IDENTIFICATION OF FUNGI IN THIS STUDY

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gi|74135500|gb|DQ123646.1| *Penicillium citrinum* strain NRRL 35449 18S ribosomal RNA gene, partial sequence; internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence; and 28S ribosomal RNA gene, partial sequence, Length=1110

Score = 880 bits (444), Expect = 0.0
Identities = 465/465 (100%), Gaps = 0/465 (0%), Strand=Plus/Plus

```

Query 4   AGTGC GGGCCCCCTCGGGGCCAACCTCCACCCGTGTTGCCCGAACCTATGTTGCCTCGG 63
          |||
Sbjct 15   AGTGC GGGCCCCCTCGGGGCCAACCTCCACCCGTGTTGCCCGAACCTATGTTGCCTCGG 74

Query 64  CGGGCCCCGCGCCCGCCGACGGCCCCCTGAACGCTGTCTGAAGTTGCAGTCTGAGACCT 123
          |||
Sbjct 75  CGGGCCCCGCGCCCGCCGACGGCCCCCTGAACGCTGTCTGAAGTTGCAGTCTGAGACCT 134

Query 124 ATAACGAAATTAGTTAAACTTTCAACAACGGATCTCTGGTTCGGGCATCGATGAAGAA 183
          |||
Sbjct 135 ATAACGAAATTAGTTAAACTTTCAACAACGGATCTCTGGTTCGGGCATCGATGAAGAA 194

Query 184 CGCAGCGAAATGCGATAACTAATGTGAATTGCAGAATTCAGTGAATCATCGAGTCTTTGA 243
          |||
Sbjct 195 CGCAGCGAAATGCGATAACTAATGTGAATTGCAGAATTCAGTGAATCATCGAGTCTTTGA 254

Query 244 ACGCACATTGCGCCCTCTGGTATTCCGGAGGGCATGCCTGTCCGAGCGTCATTGCTGCC 303
          |||
Sbjct 255 ACGCACATTGCGCCCTCTGGTATTCCGGAGGGCATGCCTGTCCGAGCGTCATTGCTGCC 314

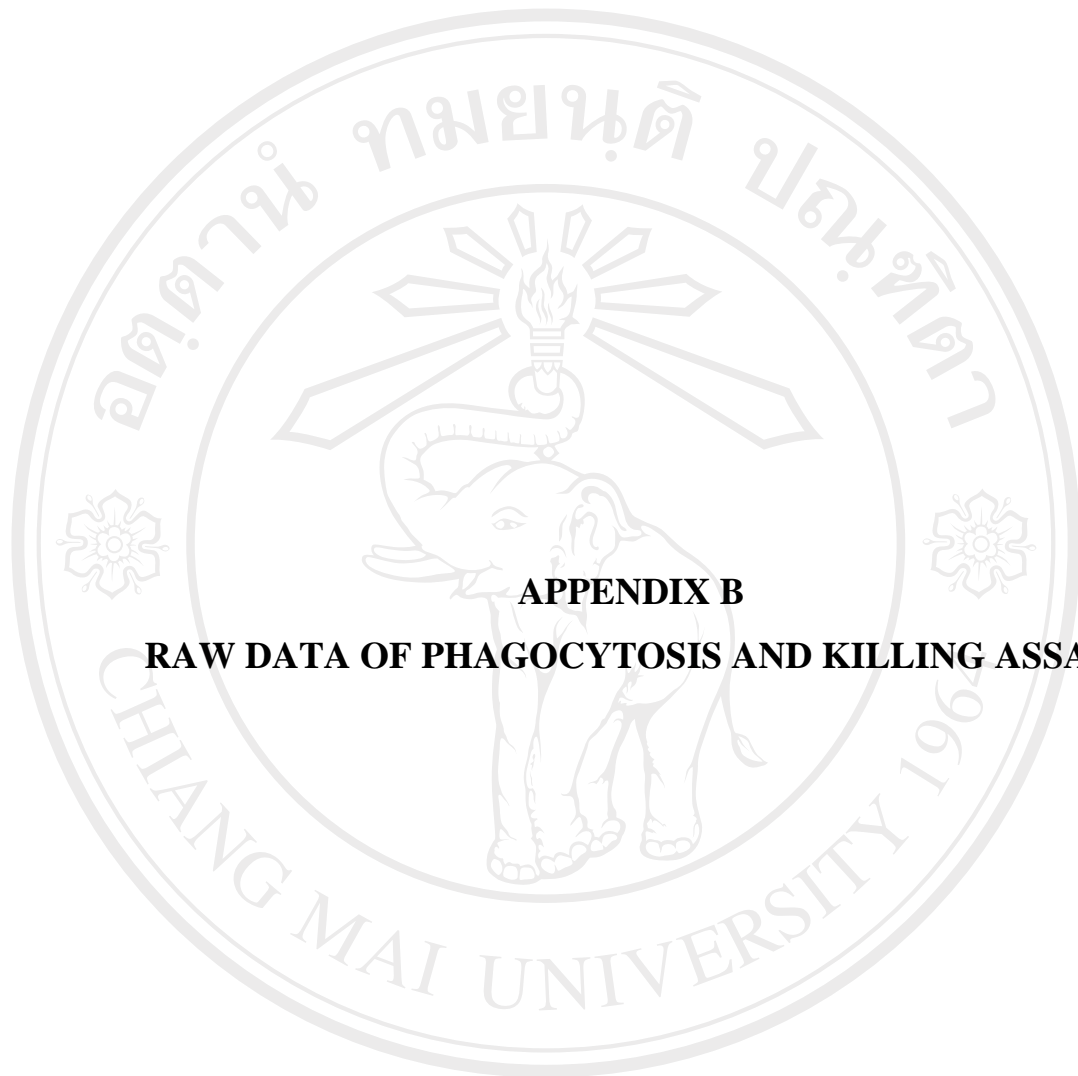
Query 304 TCAAGCCCGGCTTGTGTGTTGGGCCCGTccccccGCGGGGGGACGGGCCCGAAAGGC 363
          |||
Sbjct 315 TCAAGCCCGGCTTGTGTGTTGGGCCCGTCCCCCGCGGGGGGACGGGCCCGAAAGGC 374

Query 364 AGCGGCGCACCGCGTCCGGTCTCGAGCGTATGGGGTTCGTACCCGCTCTAGTAGGC 423
          |||
Sbjct 375 AGCGGCGCACCGCGTCCGGTCTCGAGCGTATGGGGTTCGTACCCGCTCTAGTAGGC 434

Query 424 CCGGCCGCGCCAGCCGACCCCAACCTTTAATTATCTCAGGTTG 468
          |||
Sbjct 435 CCGGCCGCGCCAGCCGACCCCAACCTTTAATTATCTCAGGTTG 479

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APPENDIX B
RAW DATA OF PHAGOCYTOSIS AND KILLING ASSAY

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Phagocytosis and killing of J774.1 macrophage on *Penicillium citrinum*

Table B-1 Phagocytosis of J774.1 on *P. citrinum* with MOI=10

Time (min)	Macrophage cell count (categorize by number of internalized conidia)											PP	PI	
	0	1	2	3	4	5	6	7	8	9	10			
0	100, 100, 100	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	-
30	54, 55, 54	27, 29, 26	14, 7, 15	6, 6, 3	2, 3, 2	1, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	50, 45, 46	1.72, 1.62, 1.59
60	34, 23, 34	30, 29, 35	18, 22, 22	10, 16, 10	3, 5, 6	4, 4, 1	1, 1, 1	0, 0, 2	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	66, 77, 77	2.03, 2.17, 2.08
120	23, 17, 20	26, 30, 40	20, 29, 21	14, 8, 7	6, 8, 8	5, 3, 2	2, 1, 1	1, 3, 3	1, 1, 0	1, 0, 0	1, 0, 0	1, 0, 0	77, 83, 80	2.64, 2.24, 2.05
240	11, 10, 15	21, 28, 36	29, 27, 18	12, 14, 14	11, 7, 8	7, 4, 4	2, 4, 2	4, 3, 3	1, 1, 0	1, 1, 0	1, 1, 0	1, 1, 0	89, 90, 85	2.93, 2.71, 2.34

Table B-2 Killing ability of J774.1 on *P. citrinum* with MOI=10

Time (min)	Control (without macrophage)		Test (co-incubate with macrophages)		% Killing
	Count	Mean CFU	Count	Mean CFU	
0	(148,144), (150,148)	(5.84 x 10 ⁵), (5.96 x 10 ⁵)	(146,144), (155,145)	(5.80 x 10 ⁵), (6.00 x 10 ⁵)	0, 0
30	(140,142), (144,151)	(5.64 x 10 ⁵), (5.90 x 10 ⁵)	(78,76), (87,92)	(3.08 x 10 ⁵), (3.58 x 10 ⁵)	45.39, 39.32
60	(139,140), (150,147)	(5.60 x 10 ⁵), (5.94 x 10 ⁵)	(49,45), (45,42)	(1.88 x 10 ⁵), (1.74 x 10 ⁵)	66.42, 70.70
120	(145,140), (146,140)	(5.72 x 10 ⁵), (5.72 x 10 ⁵)	(50,50), (41,49)	(2.00 x 10 ⁵), (1.80 x 10 ⁵)	65.03, 68.53
240	(137,146), (144,140)	(5.64 x 10 ⁵), (5.68 x 10 ⁵)	(50,58), (42,53)	(2.16 x 10 ⁵), (1.90 x 10 ⁵)	61.70, 66.54

(Test1), (Test2)

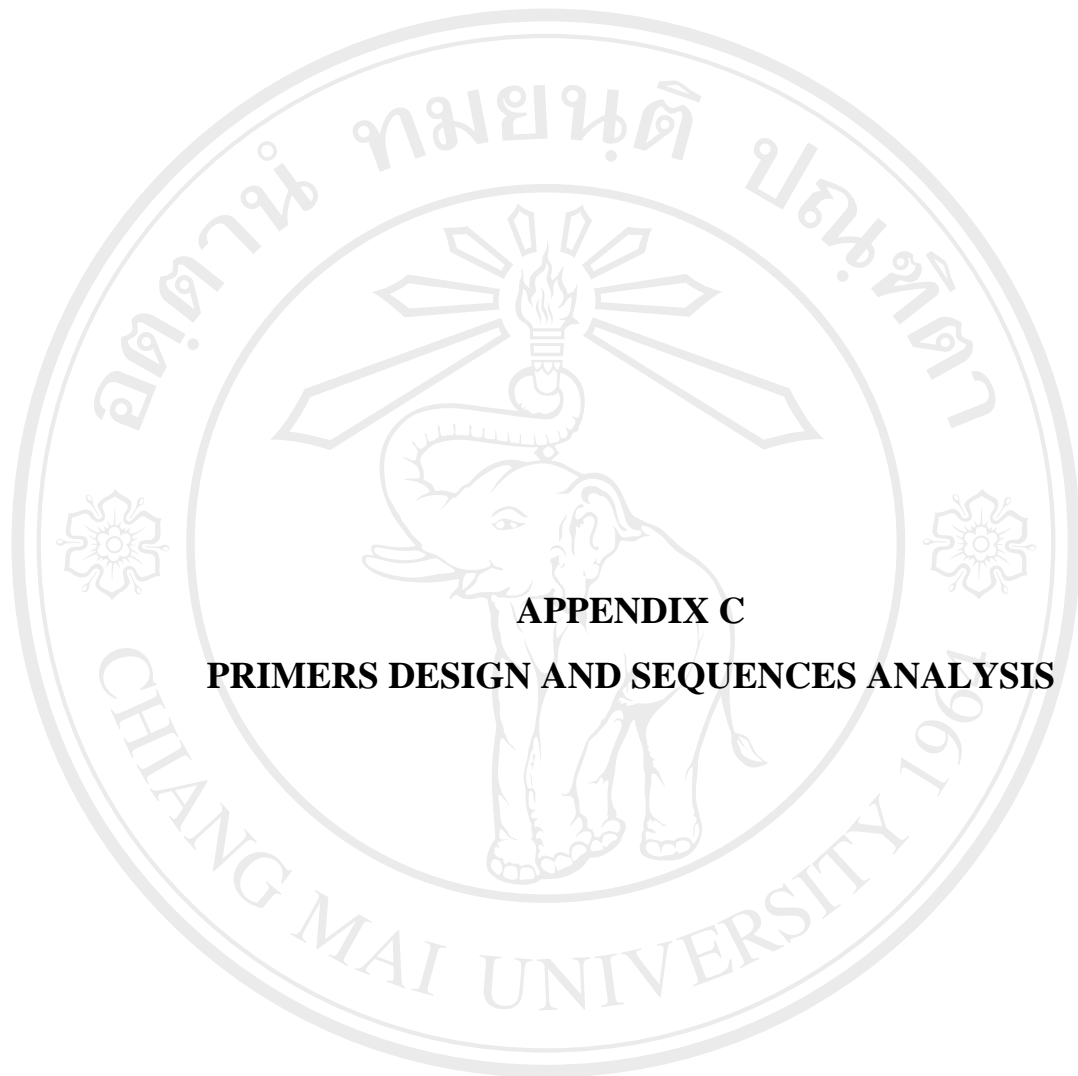
Phagocytosis and killing of J774.1 macrophage on *Penicillium marneffei*

Table B-3 Phagocytosis of J774.1 on *P. marneffei* with MOI=10

Time (min)	Macrophage cell count (categorize by number of internalized conidia)											PP	PI	
	0	1	2	3	4	5	6	7	8	9	10			
0	100, 100, 100	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	-
30	54, 55, 54	27, 29, 26	14, 7, 15	6, 6, 3	2, 3, 2	1, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	50, 45, 46	1.72, 1.62, 1.59
60	34, 23, 34	30, 29, 35	18, 22, 22	10, 16, 10	3, 5, 6	4, 4, 1	1, 1, 1	0, 0, 2	0, 0, 0	0, 0, 0	0, 0, 0	0, 0, 0	66, 77, 77	2.03, 2.17, 2.08
120	23, 17, 20	26, 30, 40	20, 29, 21	14, 8, 7	6, 8, 8	5, 3, 2	2, 1, 1	1, 3, 3	1, 1, 0	1, 0, 0	1, 0, 0	1, 0, 0	77, 83, 80	2.64, 2.24, 2.05
240	11, 10, 15	21, 28, 36	29, 27, 18	12, 14, 14	11, 7, 8	7, 4, 4	2, 4, 2	4, 3, 3	1, 1, 0	1, 1, 0	1, 1, 0	1, 1, 0	89, 90, 85	2.93, 2.71, 2.34

Table B-4 Killing ability of J774.1 on *P. marneffei* with MOI=10

Time (min)	Control (without macrophage)		Test (co-incubate with macrophages)		% Killing
	Count	Mean CFU	Count	Mean CFU	
0	(142,146), (121,128)	(5.76 x 10 ⁵), (4.98 x 10 ⁵)	(144,143), (140,110)	(5.74 x 10 ⁵), (5.00 x 10 ⁵)	0, 0
30	(139,144), (125,110)	(5.66 x 10 ⁵), (4.70 x 10 ⁵)	(125,140), (112,104)	(5.30 x 10 ⁵), (4.32 x 10 ⁵)	6.36, 8.09
60	(136,153), (120,120)	(5.38 x 10 ⁵), (4.80 x 10 ⁵)	(118,ND*), (100,90)	(4.72 x 10 ⁵), (3.80 x 10 ⁵)	12.27, 20.83
120	(136,141), (120,120)	(5.54 x 10 ⁵), (4.80 x 10 ⁵)	(103,80), (72,63)	(3.66 x 10 ⁵), (2.7 x 10 ⁵)	33.94, 43.75
240	(116,101), (122,112)	(4.52 x 10 ⁵), (4.68 x 10 ⁵)	(49,40), (48,39)	(1.78 x 10 ⁵), (1.74 x 10 ⁵)	60.62, 62.80



APPENDIX C
PRIMERS DESIGN AND SEQUENCES ANALYSIS

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C1. Cu, Zn Superoxide dismutase gene degenerate primer design

ClustalW alignment of fungal Cu, Zn SOD amino acid sequences

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Anid-Q9HEY7      MVKAVAVLRGDSKVSQVTTFEQADENSNTTVSWNITGNDPNAERGFHIHQFGDNTNGCTS 60
Afla-AAM94904    -VKAVAVLRGDSKISQVTTFEQADANAPTTVSWNITGHDANAERAFHVHQFGDNTNGCTS 59
Afum-AAL38991    MVKAVAV-RGDSKITGTVTFEQADENSPTTVSWNIKGNPNKRGFHVHQFGDNTNGCTS 59
Scer-NP_012638   MVQAVAVLKG DAGVSGVVKFEQASESEPTTVSYE IAGNSPNAERGFHIHEFGDATNGCVS 60
Psin-AAR15417    MVKAVCVLRGDSKITGIVNFEQESDSSPTTISWEISNHDADAKRGFHITPFGDNTNGCTS 60
Cneo-AAK31915    MVKAVVVLKGE SYVHGTVCFTQESENAPVCITGEIKDMDADAKRGMHVHEFGDNTNGCTS 60
                *:* * :*:: : * * * * . . . : : * . . :*:* :* : * * * * * *
                ↓
Anid-Q9HEY7      AGPHFNPFKGTGAPEDVHRVGD LGNFKTD AEGNSKGSKTDKLIK LIGAESV LGRTLVI 120
Afla-AAM94904    AGPHFNPFKGEHAPEDENRHVGD LGNFKTD AEGNAVGSKQDKLIK LIGAESV LGRTLVI 119
Afum-AAL38991    AGPHFNPHYGKTHGAPEDSERHVGD LGNFETDAEGNAVGSKQDKLIK LIGAESV LGRTLVI 119
Scer-NP_012638   AGPHFNPFKKTGAPTDEVRHVGD MGNVKTD ENGVAKGSFKDSL IKLIGPTSVVGRSVVI 120
Psin-AAR15417    AGPHFNPHGKTHGNVTDENRHVGD MGNIETDCDGN SKSISKDKLIK LIGPHSVIGRTVVI 120
Cneo-AAK31915    AGPHYNPFKKHHGAPTDSERHVGD LGNIQTNSCGAAQLDFSDKII SLYGPHSIIIGRSLV 120
                ****:* * * * * . *****:* : * : . * : * * * * * * : * : * : * : * :
                →
Anid-Q9HEY7      HAGTDDLGRGDSEESKKTGNAGARPACGVIGIAA 154
Afla-AAM94904    HAGTDDLGRSEHPESKKTGNAGARPACGVIGIAA 153
Afum-AAL38991    HAGTDDLGRGGNEESKKTGNAGARPACGVIGIAA 153
Scer-NP_012638   HAGQDDLKGKDTEESLKTGNAGPRPACGVIGLTN 154
Psin-AAR15417    HAGTDDLKGGNDESLKTGNAGPRPACGVIGVAN 154
Cneo-AAK31915    HASTDDLKGGNESLKTGNAGARLACGVIGIST 154
                ** . *****: . * * ***** * ***** :
                ←

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Primer sequences

SOD-F: SAGPHY/FNP or 5'-WSIGCIGGICCICAITWYAAAYCC-3'

SOD-R: KTGNA GA/PR or 5'-CKIGSICCIGCRTTICIGTYTT-3'

Expected PCR product: ~250 bp

Homology searching of amplified PCR product fragments

S. cerevisiae 249 bp

```

1 TGGGCGGGGC CGCAGTTTAA CCCTTTCAAG AAGACACACG GTGCTCCAAC TGACGAAGTC
61 AGACATGTCG GTGACATGGG TAACGTAAAG ACGGACGAAA ATGGTGTGGC CAAGGGCTCC
121 TTCAAGGACT CTTTGATCAA GCTTATCGGT CCTACCTCCG TTGTAGGCAG AAGCGTCGCT
181 TATCCACGCC GGCCAAGATG ACTTAGGTAA GGGTGACACT GAAGAATCTT TGAAAACCGG
241 CAATGCCCG

```

gb|AY690619.1| *Saccharomyces cerevisiae* copper-zinc superoxide
dismutase gene, complete cds, Length=496
Score = 392 bits (212), Expect = 3e-106 Identities = 223/228 (97%),
Gaps = 1/228 (0%) Strand=Plus/Plus

```

Query 22 CCTTTCAAGAAGACACACGGTGCTCCAAGTACGAAGTCAGACATGTCCGGTGACATGGGT 81
      |||
Sbjct 212 CCTTTCAAGAAGACACATGGTGCTCCAAGTACGAAGTCAGACATGTCCGGTGACATGGGT 271

Query 82 AACGTAAAGACGGACGAAAATGGTGTGGCCAAGGGCTCCTTCAAGGACTCTTTGATCAAG 141
      |||
Sbjct 272 AACGTAAAGACGGACGAAAATGGTGTGGCCAAGGGCTCCTTCAAGGACTCTTTGATCAAG 331

Query 142 CTTATCGGTCTTACCTCCGTTGTAGGCAGAAGCGTCGCTTATCCACGCCGGCCAAGATGA 201
      |||
Sbjct 332 CTTATCGGTCTTACCTCCGTTGTAGGCAGAAGCGTCG-TTATCCACGCCGGCCAAGATGA 390

Query 202 CTTAGGTAAGGGTGACACTGAAGAATCTTTGAAAACCGGCAATGCCGG 249
      |||
Sbjct 391 CTTAGGTAAGGGTGACACTGAAGAATCTTTGAAGACTGGTAATGCCGG 438

```

***P. marneffei* 251 bp**

```

1 GCCGGTCCTC ACTTCAACCC CTTCCGAAAG ACCCACGGTG CTCCCACCGA TGACGAACGC
61 CATGTCGGTG ACTTGGGTAA CTTCAAGACC GATGCTCAGG GCAATGCTGT CGGCTTCGTC
121 GAGGACAAGC TCATCAAGTT GATCGGTGCT GAGAGCGTTC TCGGACGTAC TATCGTCGTC
181 CACGCCGGTA CTGACGACCT CGGCCGGTGT GGCAACGAGG AGTCCAAGAA GACTGGCAAC
241 GCTGGTCCTC G

```

ref|XM_001215721.1| *Aspergillus terreus* NIH2624 superoxide dismutase (ATEG_06543) mRNA, complete cds, Length=492

Score = 273 bits (302, Expect = 2e-70 Identities = 208/246 (84%), Gaps = 0/246 (0%) Strand=Plus/Plus

```

Query 1      GCCGGTCCTCACTTCAACCCCTTCGGAAAGACCCACGGTGCTCCCACCGATGACGAACGC 60
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 208    GCCGGCCCTCACTTCAACCCCTTCTCCAAGACCCACGGTGCCCTGAGGATGAGGTGCCG 267

Query 61     CATGTCGGTGACTTGGGTAACCTTCAAGACCGATGCTCAGGGCAATGCTGTCGGCTTCGTC 120
          || ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 268    CACGTCGGTGACCTCGGTAACCTTCAAGACCGATGCAGAGGGCAACGCCGTCGGCTCCAAG 327

Query 121    GAGGACAAGCTCATCAAGTTGATCGGTGCTGAGAGCGTTCTCGGACGTACTATCGTCGTC 180
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 328    CAGGACAAGCTGGTCAAGCTGATCGGTGCCGAGAGCGTTCTGGGCCGTACCCTTGTCGTC 387

Query 181    CACGCCGGTACTGACGACCTCGGCCGGTGGCAACGAGGAGTCCAAGAAGACTGGCAAC 240
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 388    CACGCCGGCACTGATGATCTCGGCCGGTGGCAACGAGGAGTCCAAGAAGACTGGTAAC 447

Query 241    GCTGGT 246
          || |||
Sbjct 448    GCCGGT 453

```

gb|AF401280.1| *Aspergillus flavus* Cu,Zn-superoxide dismutase mRNA, partial cds, Length=463

Score = 268 bits (296), Expect = 8e-69 Identities = 210/251 (83%), Gaps = 0/251 (0%) Strand=Plus/Plus

```

Query 1      GCCGGTCCTCACTTCAACCCCTTCGGAAAGACCCACGGTGCTCCCACCGATGACGAACGC 60
          || || ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 179    GCTGGCCCTCACTTCAACCCCTTCCGCAAGGAGCACGGTGCTCCCGAGGATGAGAACCGC 238

Query 61     CATGTCGGTGACTTGGGTAACCTTCAAGACCGATGCTCAGGGCAATGCTGTCGGCTTCGTC 120
          || ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 239    CACGTCGGTGACTTGGGCAACCTTCAAGACCGATGCTGAGGGTAACGCCGTCGGCTCCAAG 298

Query 121    GAGGACAAGCTCATCAAGTTGATCGGTGCTGAGAGCGTTCTCGGACGTACTATCGTCGTC 180
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 299    CAGGACAAGCTTATTAAGCTGATCGGTGCCGAGAGCGTACTTGGCCGTACTCTCGTCATC 358

Query 181    CACGCCGGTACTGACGACCTCGGCCGGTGGCAACGAGGAGTCCAAGAAGACTGGCAAC 240
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 359    CACGCCGGTACTGACGACCTTGGCCGTAGTGAACACCCGAGTCCAAGAAGACTGGCAAT 418

Query 241    GCTGGTCCTCG 251
          ||||| |||||
Sbjct 419    GCTGGTGCTCG 429

```

ref|XM_748622.1| *Aspergillus fumigatus* Af293 Cu,Zn superoxide
dismutase SOD1 (AFUA_5G09240) mRNA, complete cds, Length=477
Score = 255 bits (282), Expect = 5e-65 Identities = 207/251 (82%),
Gaps = 0/251 (0%) Strand=Plus/Plus

```

Query 1      GCCGGTCCTCACTTCAACCCCTTCGGAAAGACCCACGGTGCTCCCACCGATGACGAACGC 60
             || ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 193    GCTGGTCCTCACTTCAACCCCTATGGCAAGACCCATGGAGCTCCTGAGGACTCCGAGCGC 252

Query 61     CATGTCGGTGACTTGGGTAACCTTCAAGACCGATGCTCAGGGCAATGCTGTCGGCTTCGTC 120
             ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 253    CATGTCGGTGACCTTGGTAACCTTCGAGACCGATGCTGAGGGTAACGCCGTCGGCTCCAAG 312

Query 121    GAGGACAAGCTCATCAAGTTGATCGGTGCTGAGAGCGTTCTCGGACGTACTATCGTCGTC 180
             ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 313    CAGGACAAGCTTATTAAGCTGATTGGTGCCGAGAGCGTTCTGGGCCGGACCTTGGTCGTT 372

Query 181    CACGCCGGTACTGACGACCTCGGCCGTTGGTGGAACGAGGAGTCCAAGAAGACTGGCAAC 240
             ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 373    CACGCCGGTACCGACGACCTCGGAAGGGTGGCAACGAGGAGTCCAAGAAGACTGGTAAC 432

Query 241    GCTGGTCCTCG 251
             ||||| ||||
Sbjct 433    GCTGGTGCTCG 443

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C2. Glyceraldehyde-3-phosphate dehydrogenase gene degenerate primer design

ClustalW alignment of fungal *GAPDH* nucleotide sequences

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Acap-AF273704      CGCGAGCATCCCCTGGGGCAAGCACGGCGTGAATACGTCGTTGAGTCCACCGGCGTCTT 324
Aory-AB032274     CGCTGCCATCCCCTGGGGCTCCGCTGGCGCTGCCTACATCGTCGAGTCCACTGGTGTCTT 305
Afum-XM_743052    CTCTCAGATCCCCTGGTCTGAGACCGGCGCTGCCTACATCGTCGAGTCTACTGGTGTCTT 305
Cimm-AY536450    TGCCAACATCCCATGGGCGCAAACCGGTGCCGACTACGTCATTGAATCCACTGGTGTCTT 164
Pbra-AF396657     CGCCAACATCCCCTGGGGCAAGCACGGCGTCGACTATGTCGTCGAGTCCACAGGCGTCTT 539
*      ***** ** * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Acap-AF273704      CACCACCACCGAGAAGGCCAGCGCCCACTTGAAGGGCGGTGCCAAGAAGGTCATCATCTC 384
Aory-AB032274     CACCACCACCGAGAAGGCCCTCCGCTCACTTGAAGGGTGCCGCAAGAAGGTCATCATCTC 365
Afum-XM_743052    CACCACCAAGGAGAAGGCCCTCCGCTCACTTGAAGGGTGGTGCCAAGAAGGTCATCATCTC 365
Cimm-AY536450    CACCACCACCGAGAAGGCCAGCGCTCACTTGAAGGGTGCCGCAAGAAGGTCATCATCTC 224
Pbra-AF396657     CACCACCACCTGAAAAGGCCAAGGCCATTTATCTGGTGCCGCAAAAAGGTCATCATCTC 599
*****> ** * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Acap-AF273704      CGCCCCCTCTGCAGATGCCCCCATGTTTCGTCATGGGCGTCAACGAGAAAACATACGACCC 444
Aory-AB032274     TGCTCCTTCTGCTGATGCCCCATGTTTCGTTATGGGTGTCAACAACAAGGAATACAAGAC 425
Afum-XM_743052    CGTCTCTTCTGCTGATGCCCCATGTTTCGTCATGGGTGTCAACAACAAGGTCATCATCTC 425
Cimm-AY536450    CGCCCCATCTGCCGATGCCCAATGTTTCGTAATGGGTGTCAACAACGAGACATATAAGTC 284
Pbra-AF396657     CGCCCCATCCGCTGATGCCCCCATGTTTCGTCATGGGCGTCAACGAGAAGATTACAGGCC 659
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Acap-AF273704      GAGCGTCAACGTCCTTTCCAACGCCTCCTGCACCACCAACTGCCTCGCCCCGCTTGCAAA 504
Aory-AB032274     CGACATCAACGTCCTCTCTAACGCTTCTTGCACCACCAACTGCCTTGCTCCCTTGTCTAA 485
Afum-XM_743052    CGACATCCAGGTCCTCTCCAACGCCTCTTGCACCACCAACTGCCTTGACCCCTGGCCAA 485
Cimm-AY536450    CGATATCAAGGTTCTCTCCAACGCCTCCTGCACAACCAACTGTCTTGTCTCTTGGCCAA 344
Pbra-AF396657     CGACATCTCCGTCCTCTCCAACGCCTCTTGCACCACCAACTGCCTCGCTCCACTCGCCAA 719
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Acap-AF273704      GGTTCATCAACGACAACCTTCGGCCTGACCGAGGGCCTCATGACGACCATCCACTCCTACAC 564
Aory-AB032274     GGTTCATCAACGACAACCTTCGGTCTCGTTGAGGGTCTCATGACCACTGTCCACTCCTACAC 545
Afum-XM_743052    GGTTCATCAACGACAAGTTTCGGCATTGTTGAGGGTCTCATGACCACTCCACTCCTACAC 545
Cimm-AY536450    GGTTCATCAACGACAACCTTCGGTTCGGTTGAGGGTTTGTATGACCACTCCACTCCTATAC 404
Pbra-AF396657     GGTTCATCCAGACAACCTTCGGTATTGCTGAAGACTCATGACCACTCCATTCTATAC 779
*** * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Acap-AF273704      CGCCACCCAGAAGACCGTCGACGGGCGTCTTCCAAGGACTGGCGTGGTGCCCGCACCCGC 624
Aory-AB032274     TGCTACCCAGAAGACTGTCGATGCTCCCTCCGCAAGGACTGGCGTGGTGACGTACCCGC 605
Afum-XM_743052    CGCCACCCAGAAGGTCGTCGATGCTCCCTCCAACAAGGACTGGAGAGGTGGTCTGACTGC 605
Cimm-AY536450    CGCCACTCAGAAGACTGTCGATGGCCCATCTCCAAGGACTGGCGTGGTGCCGTCGCCGC 464
Pbra-AF396657     TGCCACCCAGAAGACCGTCGATGGCCCTTCCCACAAGGACTGGCGTGGTGCCCGCACTGC 839
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

Acap-AF273704      TGCTCAGAACATCATCCCCAGCAGCACAGGCGCCGCAAGGCCGTCGGCAAGGTCATCCC 684
Aory-AB032274     CGCTCAGAACATCATCCCCAGCTCCACTGGTGTGCTGCAAGGCTGTGGCAAGGTCATTC 665
Afum-XM_743052    TGCCCAGAACATCATTTCCAGCTCGACTGGTGTGCTAAGGCCGTCGGCAAGGTCATTC 665
Cimm-AY536450    TGCTCAGAACATCATCCCTAGCAGACTGGTGTGCGCCGCAAGGCTGTGGAAAGGTCATTC 524
Pbra-AF396657     GGCTCAGAACATCATCCCCAGCAGCACTGGCGCCGCAAGGCTGTAGGAAAGGTCATCCC 899
* * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *

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Primer sequences

GAP-F: 5'-ACHGGYGYTTTACCACCA-3'

GAP-R: 5'-ATGACCTTKCCDACRGCCT-3'

Expected size ~ 400 bp

C3. Calmodulin gene

Homology searching of amplified PCR product fragments

H. capsulatum calmodulin gene fragment 570 bp

```

1 AAGGAGGCCT TCTCTCTATT TGTAAGTTGA CTTTCATTTTT GCCCCACACC TAACAGGCTT
61 ACCAACGGCG CCTCAGGACA AGAACTAACA TCGGCATTTA TTTTACAGG ATAAAGATGG
121 CGATGGTGAT TATTTCCACT TCCACTTGTT TCCAGAAGCT TAGGAAATGA GAGATGTAAA
181 AAATTTGTCG ACTGGGACTA GAGCTGACCA CTTGTTTAAA TGAAACAGGG CAAATCACCA
241 CCAAAGAGTT GGAACAGTA ATGCGGTCTC TGGGACAAA CCCTCCGAA TCAGAACTAC
301 AGGACATGAT CAACGAGGT GATGCAGATA ACAATGGAAC CATTGACTTC CCGGTATGG
361 TGCCAGTTA TTTCTCAGAT GATTTGATGC TATCTGCAAT GATGCTGTGT CTAACCGCTT
421 CTCCCTGTTT CACAGAGTTT TTGACCATGA TGGCAAGGAA AATGAAGGAT ACCGATTCTG
481 AAGAAGAAAT TCGAGAAAGC ATTCAAAGTT TTCGATCGTG ATAACAACGG TTTTATCTCC
541 TGCTGCCGAG TTACGCCCCG TGATGACTTC

```

gb|AF072882.1|AF072882 *Ajellomyces capsulatus* calmodulin (CAM1) gene, complete cds, Length=2100

Score = 941 bits (509), Expect = 0.0 Identities = 552/572 (96%), Gaps = 6/572 (1%) Strand=Plus/Plus

```

Query 1      AAGGAGGCCTTCTCTCTATTGTAAGTTGACTTCATTTTTGCCCCACACCTAACAGGCTT 60
|
Sbjct 600    AAGGAGGCCTTCTCTCTATTGTAAGTTGACTTCATTTTTGCCCCACACCTAACAGGCTT 659

Query 61     ACCAACGGCGCCTCAGGACAAGAATAACATCGGCATTATTTTACAGGATAAAGATGG 120
|
Sbjct 660    ACCAACCGCCTCAGGACAAGAATAACATCGGCATTTA-TTTTACAGGATAAAGATGG 718

Query 121    CGATGGTGATTATTTCCACTTCCACTTGTTTCCAGAAGCTTAGGAAATGAGAGATGTAAA 180
|
Sbjct 719    CGATGGTAATTATTTCCACTTCCACTTGTTTCCAGAAGCTTAGGAAATGAGAGATGTAAA 778

Query 181    AAATTTGTCGACTGGGACTAGAGCTGACCCTGTTTAAATGAAACAGGGCAAATCACCA 240
|
Sbjct 779    ACATTTGTTGACTGGGACTAGAGCTGACCCTGTTTAAATGAAACAGGGCAAATCACCA 838

Query 241    CCAAAGAGTTGGGAACAGTAATGCGGTCTCTGGGACAAAACCTTCCGAATCAGAACTAC 300
|
Sbjct 839    CCAAAGAGTTGGGAACAGTAATGCGGTCTCTGGGACAAAACCTTCCGAATCAGAACTAC 898

Query 301    AGGACATGATCAACGAGGTTGATGCAGATAACAATGGAACATTGACTTCCCGGTATGG 360
|
Sbjct 899    AGGACATGATCAATGAGGTTGATGCAGATAACAATGGAACATTGACTTCCCGGTATGG 958

Query 361    TGCCAGTTATTTCTCAGATG-ATTTGAT-GCTATCTGCAATGATGCTGTGTCTAACC GC 418
|
Sbjct 959    TGCCAGTTATTTCTCAGATTCAATTTGATCG-TATCTGCAATGATGCTGTGTCTAACC GC 1017

Query 419    TTCTCCCTGTTTCACAGAGTTTTTGACCATGATGGCAAGGAAAATGAAGGATACCGATTC 478
|
Sbjct 1018   TTCTCCCTGTTTCACAGAGTTTTTGACTATGATGGCAAGGAAAATGAAGGATACCGATTC 1077

Query 479    TGAAGAAGAAATTCGAGAAAGCATTCAAAGTTTTTCGATCGTGATAACAACGGTTTTATCT 538
|
Sbjct 1078   TGAAGAAGAAATTCGAGAA-GCATTCAAAGTTTTTCGATCGTGATAACAACGGTTTTATCT 1136

Query 539    CCTGCTGCCGAGTTACGCCCCGTGATGACTTC 570
|
Sbjct 1137   C-TGCTGCCGAGTTACGCCACGTGATGACTTC 1167

```

***P. marneffei* calmodulin gene fragment 550 bp**

```

1 CTCTCTATTT GTAAGTTTGG ATTTCTGGTT GTCGCAATGT TGTGGTGGGT GGTTCGCTGA
61 CTAGCCGTTT GGATGAATAG GACAAGGATG GTGATGGTGA GTGACGCCAC GAACACCAGA
121 CATATAGTCT TCGAACAAAA AGTTATTACT GCGAACAGAT ATTAATAACA TCAATAGGTC
181 AAATTACAAC CAAGGAACTG GGCACCGTCA TGC GTTCCCT CGGCCAGAAC CCTTCCGAAT
241 CCGAATTGCA GGACATGATC AACGAGGTCG ACGCTGACAA CAACGGCACA ATCGATTTC
301 CTGGTATGAT GCAGCCTCTA TTTATCGCAG CCGTTTCCGA TCATAAGGGC AGATACTGAC
361 TGCCTTAGAA TTCTTGACAA TGATGGCCCG CAAAATGAAG GATACCGACT CCGAGGAAGA
421 GATCCGCGAG GCTTTCAAGG TGTTTGATCG TGACAACAAT GGATTCATCT CTGCTGCTGA
481 ATTGCGCCAC GTTATGACCT CGATTGGCGA AAAGTTGACC GACGACGAAG TTGACGAGAT
541 GATTCCGCGAG

```

gb|AY678609.1| *Talaromyces flavus* strain AS3.6681 calmodulin (cmd)
 gene, partial cds, Length=540
 Score = 592 bits (320), Expect = 6e-166 Identities = 469/537 (87%),
 Gaps = 26/537 (4%) Strand=Plus/Plus

```

Query 1 CTCTCTATTTGTAAGTTTGG-ATTTCTGGTTGTCGCAATGTTGTGGTGGGTGGTTTCGCTG 59
Sbjct 25 CTCTCTTTTTGTAAGTTTAGCAATT-TGTTTTCGCAATGTTGTGGTGGGTGGTT-GCTG 82
Query 60 ACTAGCCGTTTGGATGAATAGGACAAGGATGGTGATGGTGAGTGACG-CCACGAACACCA 118
Sbjct 83 ACTAGCCGTTTGTATGAGTAGGACAAGGATGGAGATGGTGAGT--CGGCCACGAACAATA 140
Query 119 GACATATAGTC-TTCGAACAAAAAGTTATTACTGCGAACAGATATTAA-TAACATC-AAT 175
Sbjct 141 TCCAAA-AGGCGTT-GAACAAGG-GTTATCGCCGGAACAGATATTGACTAT-ATCGAAT 196
Query 176 AGGTCAAATTACAACCAAGGAACTGGGCACCGTCATGCGTTCCCTCGGCCAGAACCCCTC 235
Sbjct 197 AGGTCAAATCACAACCAAGGAACTGGGCACCTGTATGCGTTCCCTCGGCCAGAACCCCTC 256
Query 236 CGAATCCGAATTGCAGGACATGATCAACGAGGTCGACGCTGACAACAACGGCACAATCGA 295
Sbjct 257 CGAATCCGAATGACAGGACATGATCAACGAAGTCGACGCTGACAACAACGGCACAATCGA 316
Query 296 TTTCCCTGGTATGATGCAGCCTCTATTTATCGCAGCCGTTTCCGATCATAAGGGCAGATA 355
Sbjct 317 TTTCCCTGGTATGAT--A---TA-ATTGCTCACAG--GAT---ATTATGATGGCAG-TA 363
Query 356 CTGACTGCCTTAGAATTCTTGACAATGATGGCCCGCAAAAATGAAGGATACCGACTCCGAG 415
Sbjct 364 CTAAGTCCCGCAGAAATCCTGACAATGATGGCCCGCAAAAATGAAGGACACCGACTCCGAG 423
Query 416 GAAGAGATCCGCGAGGCTTTCAAGGTGTTTGATCGTGACAACAATGGATTTCATCTCTGCT 475
Sbjct 424 GAAGAGATCCGCGAGGCTTTCAAGGTGTTTGACCGTGACAACAATGGATTTCATCTCTGCT 483
Query 476 GCTGAATTGCGCCACGTTATGACCTCGATTGGCGAAAAAGTTGACCGACGACGAAGTT 532
Sbjct 484 GCCGAATTGCGCCACGTTATGACCTCGATTGGCGAGAAAGTTGACCGACGACGAGGTT 540

```

gb|AY678604.1| *Penicillium pinophilum* strain AS3.6551 calmodulin
(cmd) gene, partial cds, Length=541
Score = 592 bits (320), Expect = 6e-166 Identities = 469/537 (87%),
Gaps = 25/537 (4%) Strand=Plus/Plus

```

Query 1 CTCTCTATTTGTAAGTTTGGATTTCTGGTTGTCGCAATGTTGTGGTGGGTGGTTCGCTGA 60
      |||
Sbjct 25 CTCTCTATTTGTAAGTTTGGAAATCTGGTTGTCGCAATGTTGTGGTGGATGGTTAGCTGA 84

Query 61 CTAGCCGTTTGGATGAATAGGACAAGGATGGTGATGGTGAGTGACCCACGAACACCAGA 120
      |||
Sbjct 85 CTAGCCGTTTGGATGAGTAGGACAAGGATGGAGATGGTGAGT-CCGCCACGAACA-C-GA 141

Query 121 C-ATA-TAGTCTTCGAACAAAAAG-TTATTACTGCGAACAGATATTAATAACATC-AATA 176
      |||
Sbjct 142 CGATATTTGTC-TCGAAC-AAAGGTTTTTTTC-ACGAGCATATATTGATAAAAATCTAATA 198

Query 177 GGTCAAATTACAACCAAGGAACTGGGCACCGTCATGCGTTCCTCGGCCAGAACCCCTCC 236
      |||
Sbjct 199 GGCCAAATTACAACCAAGGAACTGGGCACCGTTATGCGTTCCTCGGCCAGAACCCCTCC 258

Query 237 GAATCCGAATTGCAGGACATGATCAACGAGGTCGACGCTGACAACAACGGCACAATCGAT 296
      |||
Sbjct 259 GAATCCGAACTGCAGGACATGATCAACGAAGTCGACGCTGACAACAACGGCACAATCGAT 318

Query 297 TTCCCTGGTATGATGCAGCCTCTATT-TATCGCAGCCGTTTCCGATCATAAGGGCAGATA 355
      |||
Sbjct 319 TTCCCTGGTATGAT--A---T-AATTGT-TCAC-G-GGTTT---AT-ACGATGGCAG-TA 364

Query 356 CTGACTGCCTTAGAATCTTGACAATGATGGCCCGCAAAATGAAGGATACCGACTCCGAG 415
      |||
Sbjct 365 CTAAGTGCCTTAGAATCTTGACAATGATGGCCCGCAAAATGAAGGATACCGACTCCGAG 424

Query 416 GAAGAGATCCCGGAGGCTTTCAAGGTGTTTGATCGTGACAACAATGGATTTCATCTCTGCT 475
      |||
Sbjct 425 GAAGAGATCCCGTGAAGCTTTCAAGGTGTTTGACCGTGACAACAATGGTTTTCATCTCTGCT 484

Query 476 GCTGAATTGCGCCACGTTATGACCTCGATTGGCGAAAAGTTGACCGACGACGAAGTT 532
      |||
Sbjct 485 GCTGAACTGCGCCACGTCATGACCTCGATTGGCGAGAAGTTGACCGACGATGAGGTT 541

```


C4. Catalase gene primer design

P. marneffei amplified PCR fragment 558 bp

```

1 GCGGTGGATT CGTTTCTTAC CCCGAGAAGA TCCAGGCAAT CAAAAAGCGT AGCCAGGGAC
61 CCAAGTTCCG CGAACATCAC AACCAGGCGC AGTTATCTA CAACTCTCTT ACTGAATACG
121 AGCAACACCA CGTCGCCAAA GCGTTTAGCT TTGAGCTTGA CCATTGCGAC GACCCTGTGG
181 TCTACCAGCG CATGGCCTTG CGTATTGACG AGATTGATCT TTCCCTCGCC CAGAAAGTCG
241 CCGTAATGGT CGGCGCCCA ACACCTGAAA CTCCCGGAAA GCAGAACCAC GGCAAAAAGA
301 CTCGCGGTCT TTCACAAATT GACTTCAAGC CTCGTGTACC AACTGTGGCC AGCAGACGTA
361 TCGCCATAAT CATTGGCGAC GGCTTCGACT CTGTCGCTTT TAACGGAGTG TATACTGCCA
421 TCAAAGCTGC TGGCGCTCTG CCATTTGTCA TCGGCACAAA GAGACAGCCC ATCTTCGCAG
481 ATGGCGTTGA CCCCAGACC GGAAATGGCG TAACACCCGA ACACCAATAC GAGGGTGTGC
541 GTTCTACAAT GTTCGACG

```

ClustalW alignment of *cat* gene fragments

```

Pmar-AL683912      ACTAAGGGAACAGTCAACTACTGGCCGAATCGGTTTGATGCGGTACCACCAATCAAGCCT 60
Pmar-PCR          -----

Pmar-AL683912      GAGGACGGTGGATTTCGTTTCTTACCCCGAGAAGATCCAGGCAATCAAAAAGCGTAGCCAG 120
Pmar-PCR          ----GCGGTGGATTTCGTTTCTTACCCCGAGAAGATCCAGGCAATCAAAAAGCGTAGCCAG 56
                    *****

Pmar-AL683912      GGACCCAAGTTCGCGAACATCACAACCAGGCGCAGTTATTCTACAACCTCTTACTGAA 180
Pmar-PCR          GGACCCAAGTTCGCGAACATCACAACCAGGCGCAGTTATTCTACAACCTCTTACTGAA 116
                    *****

Pmar-AL683912      TACGAGCAACACCACGTCGCCAAAGCGTTTAGCTTTGAGCTTGACCATTGCGACGACCCCT 240
Pmar-PCR          TACGAGCAACACCACGTCGCCAAAGCGTTTAGCTTTGAGCTTGACCATTGCGACGACCCCT 176
                    *****

Pmar-AL683912      GTGGTCTACCAGCGCATGGCCTTGCCTATTGCAGAGATTGATCTTTCCCTCGCCAGAAA 300
Pmar-PCR          GTGGTCTACCAGCGCATGGCCTTGCCTATTGCAGAGATTGATCTTTCCCTCGCCAGAAA 236
                    *****

Pmar-AL683912      GTCGCCGTAATGGTCGGCGCCCCAACACCTGAAACTCCCGGAAAGCAGAACCACGGCAAA 360
Pmar-PCR          GTCGCCGTAATGGTCGGCGCCCCAACACCTGAAACTCCCGGAAAGCAGAACCACGGCAAA 296
                    *****

Pmar-AL683912      AAGACTCGCGGTCTTTCACAAATTGACTTCAAGCCTCGTGTACCAACTGTGGCCAGCAGA 420
Pmar-PCR          AAGACTCGCGGTCTTTCACAAATTGACTTCAAGCCTCGTGTACCAACTGTGGCCAGCAGA 356
                    *****

Pmar-AL683912      CGTATCGCCATAATCATTGGCGACGGCTTCGACTCTGTGCTTTTAAACGGAGTGTATACT 480
Pmar-PCR          CGTATCGCCATAATCATTGGCGACGGCTTCGACTCTGTGCTTTTAAACGGAGTGTATACT 416
                    *****

Pmar-AL683912      GCCATCAAAGCTGCTGGCGCTCTGCCATTTGTATCGGCACAAAGAGACAACCCATCTTC 540
Pmar-PCR          GCCATCAAAGCTGCTGGCGCTCTGCCATTTGTATCGGCACAAAGAGAGACAACCCATCTTC 476
                    *****

Pmar-AL683912      GCAGATGGCGTTGACCCCCAGACCGGANATGGCGTAACACCCGAACACCAATACGAGGGT 600
Pmar-PCR          GCAGATGGCGTTGACCCCCAGACCGGAAATGGCGTAACACCCGAACACCAATACGAGGGT 536
                    *****

Pmar-AL683912      GTGCGTTTACAATGTTTCGACGCTACTTTTCATCCCCGGTGGACCACACGTCAA 653
Pmar-PCR          GTGCGTTTACAATGTTTCGACG----- 558
                    *****

```


dbj|AB079611.1| *Aspergillus oryzae* catA gene for catalase, complete cds, Length=3210

Score = 165 bits (182), Expect = 2e-37 Identities = 378/563 (67%), Gaps = 11/563 (1%) Strand=Plus/Plus

```

Query 3      GGTGGATTTCGTTTCTTACCCCGAGAAGATCCAGGCAATCAAAAAGCGTAGCCAGGGACCC 62
          ||||| ||||| || ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 1982    GGTGGCTTCGCGTCTTACCCTGCGAAACTGAACGGTATCAAGAAGCGCGGCCTGAGCCCC 2041

Query 63     AAGTTCGCGAACATCACACCAGGCGCAGTTATTCTACAACCTCTTACTGAATACGAG 122
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2042    AAGTTCGCGAGCACCACAACCAGGCTCAACTCTTCTACAACCTCTCTCCGAGCACGAG 2101

Query 123    CAACACCACGTCGCCAAGCGTTTAGCTTTGAGCTTGACCATTGCGACGACCCTGTGGTC 182
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2102    AAGTCCACGTCGAAGAAGGCCTTCGGCTTCGAACTGGACCCTGCGACGACCCCATCGTC 2161

Query 183    TACCAGCGCATGGC---CTTGCGTATTGCAGAGATTGATCTTCCCTCGCCAGAAAGTC 239
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2162    TACGAGCGCCTCGCCGCCACCGTCTCGCCGAGATCGATCTCACTCTCGCCAGGAAGTC 2221

Query 240    GCCGTAATGGTCGGCGCCCCAACACCTGAAACTCCCGAAAGCAGAACCACGGCAAAAAG 299
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2222    GCCGAGCTCGTCGGCGCCCCGATCCAGACAAGGCACTTCGCCCGAACCATGAAAGCGC 2281

Query 300    A-CTCGGGTCTTTCACAAATTGACTTCAAGCCTCGT--GTA-CCAACTGTGGCCAGCAG 355
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2282    AGCAAGC-ATCTTTCGCAGACCGAGTTC---CCGGTAAGCAGCCGACGATCGCCAGTCG 2337

Query 356    ACGTATCGCCATAATCATTGGCGACGGCTTCGACTCTGTGCTTTTAACGGAGTGTATAC 415
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2338    CCGAATCGCCATCATTATCGGCGACGGATACGACCCCGTTCGCTTTCAATGGCCTCAAGGG 2397

Query 416    TGCCATCAAAGCTGCTGGCGCTCTGCCATTTGTCATCGGCACAAAGAGACAGCCCATCTT 475
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2398    CGCCATCACGGCGGTTGGAGCCTTACCCTTCGTCATTGGCACCAAGCGGTCACCTATCTA 2457

Query 476    CGCAGATGGCGTTGACCCCCAGACCGGAAATGGCGTAACACCCGAACACCAATACGAGGG 535
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2458    CGCCGACGGTGAGGACAAATCATCTTCCAAGGGCGTGATCGCCGACCACCAGTATGACGG 2517

Query 536    TGTGCGTTCTACAATGTTTCGACG 558
          ||||| ||||| ||||| |||||
Sbjct 2518    ACAGCGTTCGACGATGTTTGACG 2540

```

gb|AY103479.1| *Ajellomyces capsulatus* catalase A (CATA) gene, complete cds, Length=4558

Score = 107 bits (118), Expect = 4e-20 Identities = 146/204 (71%), Gaps = 0/204 (0%) Strand=Plus/Plus

```

Query 62     CAAGTTCGCGAACATCACACCAGGCGCAGTTATTCTACAACCTCTTACTGAATACGA 121
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2455    CAAGTTCGCGAGCATATCAACCAAGCGCAATTATTCTATAACTCTCTCTCCGAATACGA 2514

Query 122    GCAACACCACGTCGCCAAGCGTTTAGCTTTGAGCTTGACCATTGCGACGACCCTGTGGT 181
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2515    AAAGTCCATGTCAATAATGCTTTTTCGCTTTGAGTTGGATCACTGCGATGACCCATTGT 2574

Query 182    CTACCAGCGCATGGCCTTGCCTATTGCAGAGATTGATCTTCCCTCGCCAGAAAGTCGC 241
          ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
Sbjct 2575    CTACAATAGACTTGTGTCTCGGATTTCTGAAATCGACCATGCCCTCGCCAAAGCCGTTGC 2634

Query 242    CGTAATGGTCGGCGCCCCAACACC 265
          ||||| ||||| ||||| |||||
Sbjct 2635    AGTGAAAGTCGGCGCACCGACCCC 2658

```

C5. Primer for *sodA* expression

→

```

cDNA      ATGGTCAAGGCTGTCGCTGTCCTCCGTTGGAGACTCCAACATCAAGGGTACCGTCACCTTT 60
gDNA      ---GTCAAGGCTGTCGCTGTCCTCCGTTGGAGACTCCAACATCAAGGGTACCGTCACCTTT 57
          *****

cDNA      GAACAGGCTGACGAGAACTCCCTTACCACCATCTCATGGAACATCACCGGCCACGACGCC 120
gDNA      GAACAGGCTGACGAGAACTCCCTTACCACCATCTCATGGAACATCACCGGCCACGACGCC 117
          *****

cDNA      AACGCTGAGCGTGGCATCCACGTTACCAGTTCGGTGACAACACCAACGGCTGCACATCT 180
gDNA      AACGCTGAGCGTGGCATCCACGTTACCAGTTCGGTGACAACACCAACGGCTGCACATCT 177
          *****

cDNA      GCCGGTCTCACT----- 193
gDNA      GCCGGTCTCACTGTATGCTTCCGACCTCCTCACTCCTCACTTCTCACTAAAGTGTGGC 237
          *****

cDNA      -----TCAACCCCTTCGGAAAGACCCACGGTGCCTCCAC 227
gDNA      ATCACCGTACTAATAGAAATTCATAGTCAACCCCTTCGGAAAGACCCACGGTGCCTCCAC 297
          *****

cDNA      CGATGACGAACGCCATGTCCGGTACTTGGGTAACCTCAAGACCGATGCTCAGGGCAATGC 287
gDNA      CGATGACGAACGCCATGTCCGGTACTTGGGTAACCTCAAGACCGATGCTCAGGGCAATGC 357
          *****

cDNA      TGTCCGGCTTCGTCGAGGACAAGCTCATCAAGTTGATCCGGTGCTGAGAGCGTTCTCGGA-- 345
gDNA      TGTCCGGCTTCGTCGAGGACAAGCTCATCAAGTTGATCCGGTGCTGAGAGCGTTCTCGGAGT 417
          *****

cDNA      -----CGT 348
gDNA      ATGTCCCAATTTCTATGAGCATCTAGATGTTGTGTATATGTACTAACCGTTAATAGCGT 477
          ***

cDNA      ACTATCGTCGTCCACGCCGGTACTGACGACCTCGGCCGTGGTGGCAACGAGGAGTCCAAG 408
gDNA      ACTATCGTCGTCCACGCCGGTACTGACGACCTCGGCCGTGGTGGCAACGAGGAGTCCAAG 537
          *****

cDNA      AAGACTGGCAACGCTGGTCTCGTCCCTGCTTGCG----- 444
gDNA      AAGACTGGCAACGCTGGTCTCGTCCCTGCTTGCGGTAAGCTGTCTATTCTCAAACAAAGG 597
          *****
    
```

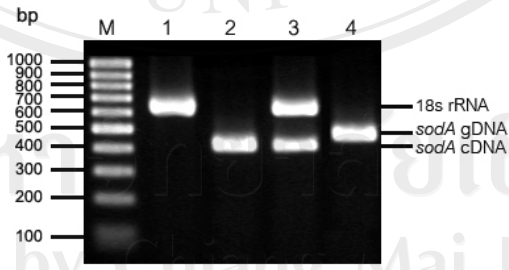


Figure C5. The amplified product of duplex RT-PCR with RRF1/RRH1 (18S rRNA) and specific primers for *sodA*. The product of 18S rRNA is 630 bp (lane 1), 448 bp of *sodA* is shown in lane 2 and mixed primers in lane 3. Lane 4 shows 521 bp amplified product form gDNA of *P. marneffei*.

C6. Primer for *gapdh* expression

```

cDNA      →
gDNA      GTCCGAGACTGGTGCCGCTTACATTGTCGAGTCCACCGGTGCTTCACCACCACCGAGAA
          GTCCGAGACTGGTGCCGCTTACATTGTCGAGTCCACCGGTGCTTCACCACCACCGAGAA
          *****

cDNA      GGCCTCTGCTCACTTGAAGGGTGGCGCCAAGAAGGTCGTCATCTCTGCTCCTTCCGCAGA
gDNA      GGCCTCTGCTCACTTGAAGGGTGGCGCCAAGAAGGTCGTCATCTCTGCTCCTTCCGCAGA
          *****

cDNA      TGCTCCT-----ATG
gDNA      TGCTCCTGTCAGTCCTCTCCCCTTCGTGGCCCTGTGGCCTGTTGCTAACCTATGTAGATG
          *****

cDNA      TTCGTCATGGGTGTCAACCACACAACCTACAAGAGCTCAGACACCATCATCTCCAACGCT
gDNA      TTCGTCATGGGTGTCAACCACACAACCTACAAGAGCTCAGACACCATCATCTCCAACGCT
          *****

cDNA      TCTTGACCAACCAACTGCTGGCTCCCTCGCCAAGATTGTCAACGACAACCTGGGGTCTT
gDNA      TCTTGACCAACCAACTGCTGGCTCCCTCGCCAAGATTGTCAACGACAACCTGGGGTCTT
          *****

cDNA      GTTGAGGGTCTCATGACCACCACTTCACTCCTACACTGCTACCCAGAAGACCGTTGATGGT
gDNA      GTTGAGGGTCTCATGACCACCACTTCACTCCTACACTGCTACCCAGAAGACCGTTGATGGT
          *****

cDNA      CCCTCCGCCAAGGACTGGCGTGGTGGTGTACCGCTGCTCAGAACATCATTTCCAGCAGC
gDNA      CCCTCCGCCAAGGACTGGCGTGGTGGTGTACCGCTGCTCAGAACATCATTTCCAGCAGC
          *****

cDNA      ACTGGTGCCGCCAAGGCTGTGCGAAAGGTCATTCTGCCCTTAACGAAAGCTCACTGGA
gDNA      ACTGGTGCCGCCAAGGCTGTGCGAAAGGTCATTCTGCCCTTAACGAAAGCTCACTGGA
          *****

cDNA      ATGTCATGCGTGTTCCTACCTCCAACGTCTCCGTTGTTGACTTGACCTGCCGCACTGAG
gDNA      ATGTCATGCGTGTTCCTACCTCCAACGTCTCCGTTGTTGACTTGACCTGCCGCACTGAG
          *****
    
```

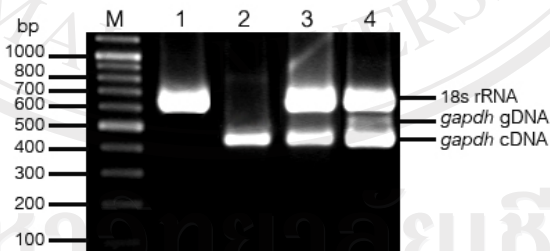


Figure C6. The amplified product of duplex RT-PCR with RRF1/RRH1 (18S rRNA) and specific primers for *gapdh*. The product of 18S rRNA is 630 bp (lane 1), 420 bp of *gapdh* is shown in lane 2 and mixed primers in lane 3. Lane 4 shows 470 bp amplified product of contaminated gDNA of *P. marneffei*.

C7. Primers for *acuD* expression

cDNA	GGCTGATATCATGGGTA-----	712
gDNA	GGCTGATATCATGGGTATGACTGTTTGCCATCTCCCAATGGAAAGAATGA	2250

cDNA	----- <u>CCGACCTCCTTGCCATTGCACGTACCG</u>	739
gDNA	ATCTAACATTGACTATATAGGTACCGACCTCCTTGCCATTGCACGTACCG	2300

cDNA	ACTCCGAAGCTGCTACTCTCATCACTTCTACCATCGACTACCGCGACCAC	789
gDNA	ACTCCGAAGCTGCTACTCTCATCACTTCTACCATCGACTACCGCGACCAC	2350

cDNA	GCCTACTTACTCGGATCTACCAACCCAGCCTCCAACCTCTTAATGACCT	839
gDNA	GCCTACTTACTCGGATCTACCAACCCAGCCTCCAACCTCTTAATGACCT	2400

cDNA	GATGGTTGCTGCCGAACAGTCCGGCAAATCCGGTGAACAACCTCCAAGCCA	889
gDNA	GATGGTTGCTGCCGAACAGTCCGGCAAATCCGGTGAACAACCTCCAAGCCA	2450

cDNA	TCGAAGACTCATGGATCGCTCAAGCCGGCATCAAGAAATTTGACGACGCC	939
gDNA	TCGAAGACTCATGGATCGCTCAAGCCGGCATCAAGAAATTTGACGACGCC	2500

cDNA	GTGATCGACACCATCAACCAAGGCAGCTTTGCCAATAAGAAAGAACTCAT	989
gDNA	GTGATCGACACCATCAACCAAGGCAGCTTTGCCAATAAGAAAGAACTCAT	2550

cDNA	CAACAGGTACCTCACAGCTGCCAAGGGCAAGAGCAACTCTGAAGCCCGTG	1039
gDNA	CAACAGGTACCTCACAGCTGCCAAGGGCAAGAGCAACTCTGAAGCCCGTG	2600

cDNA	CAATTGCCAAGGGAATCACCGGCATGGATATTTACTGGAACGGGATGCG	1089
gDNA	CAATTGCCAAGGGAATCACCGGCATGGATATTTACTGGAACGGGATGCG	2650

cDNA	CCCCGAACCCGAGGGGTTTCTATAGGTACCAGGGTGGTACGCAATGCGC	1139
gDNA	CCCCGAACCCGAGGGGTTTCTATAGGTACCAGGGTGGTACGCAATGCGC	2700

cDNA	CGTCAACCGTGCCGTC <u>GCTTATGCACCATTGCGCGATCT</u> GATTGGATGG	1189
gDNA	CGTCAACCGTGCCGTCGCTTATGCACCATTGCGCGATCTGATTGGATGG	2750

cDNA	AGAGCAAGCTTCTGATTACAAGCAAGCCAAGGAGTTTGCTGATGGTGTG	1239
gDNA	AGAGCAAGCTTCTGATTACAAGCAAGCCAAGGAGTTTGCTGATGGTGTG	2800

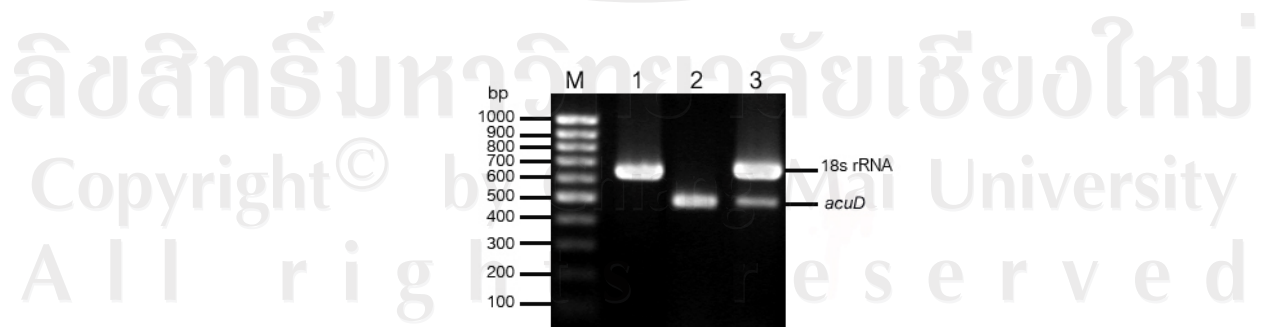


Figure C7. The amplified product of duplex RT-PCR with RRF1/RRH1 (18S rRNA) and specific primers for *acuD*. The product of 18S rRNA is 630 bp (lane 1), 450 bp of *acuD* is shown in lane 2 and mixed primers in lane 3.

C8. Primer for *hsp70* expression

gDNA	ACTTCGACTCTCGCCTTGTCACCACTTTGCCTCCGAGTTCAAGAGGAAATATAAGAGTA	840
cDNA	ACTTCGACTCTCGCCTTGTCACCACTTTGCCTCCGAGTTCAAGAGGAAATATAAGAGT---	838

gDNA	TGTGATATACCCCATCTGTTTACAAGAATTGATAAGCTAACAACCTTTCACAGAGGGTA	900
cDNA	-----AGG-----	
gDNA	CGTCAAAAATCCGAAATGAATTTCATTCCTCCCGTCTCTTGCATTCTTATTTGATATT	960
cDNA	-----	
gDNA	TGGAGCTTGGCTCTGGAGACTTCCCTCAGAACCATCTGAAGAATGCATATGAAGACTCCCA	1020
cDNA	-----	
gDNA	ATATGAATCCATCTCCTTCCCTTTCATCCGCACTTGACTCTTGAAGAACTCCTCAGATGC	1080
cDNA	-----	
gDNA	TACATGTGAAATAGATTGACCACCAATGCTCGTCTCCGCCGCTCTCCGCACTGCCT	1140
cDNA	-----ATTGACCACCAATGCTCGTCTCCGCCGCTCTCCGCACTGCCT	885

gDNA	GTGAGCGTGCTAAGCGTACCCTCTCTTCTCCGCCAGACCTCCATTGAGATCGACTCTC	1200
cDNA	GTGAGCGTGCTAAGCGTACCCTCTCTTCTCCGCCAGACCTCCATTGAGATCGACTCTC	945

gDNA	TCTTCGAGGGTATTGACTTCTACACCTCCATCACCCGTGCTCGTTTCGAGGAGCTCTGCC	1260
cDNA	TCTTCGAGGGTATTGACTTCTACACCTCCATCACCCGTGCTCGTTTCGAGGAGCTCTGCC	1005

gDNA	AGGATCTCTCCGTTCCACCATGGAGCCCGTCGAGCGTGTCTCCGTGATGCCAAGACCG	1320
cDNA	AGGATCTCTCCGTTCCACCATGGAGCCCGTCGAGCGTGTCTCCGTGATGCCAAGACCG	1065

gDNA	ACAAGTCTTCTGTCCACGAAATCGTCTTGGTCGGTGGTCCACCCGATCCCCAAGATCC	1380
cDNA	ACAAGTCTTCTGTCCACGAAATCGTCTTGGTCGGTGGTCCACCCGATCCCCAAGATCC	1125

gDNA	AGAAGCTCGTCACCGACTTCTTCAACAAGGAGCCCAACAAGTCCATCAACCCCGATGAGG	1440
cDNA	AGAAGCTCGTCACCGACTTCTTCAACAAGGAGCCCAACAAGTCCATCAACCCCGATGAGG	1185

gDNA	CTGTTGCCTACGGTGTGCGCTCCAGGCTGCTATCCTTTCTGGTGACACTTCTTCCAAGT	1500
cDNA	CTGTTGCCTACGGTGTGCGCTCCAGGCTGCTATCCTTTCTGGTGACACTTCTTCCAAGT	1245

gDNA	CCACCAACGAAATCTTGCTTCTCGACGTTGCTCCTCTCTCCGTCGGTATTGAGACTGCTG	1560
cDNA	CCACCAACGAAATCTTGCTTCTCGACGTTGCTCCTCTCTCCGTCGGTATTGAGACTGCTG	1305

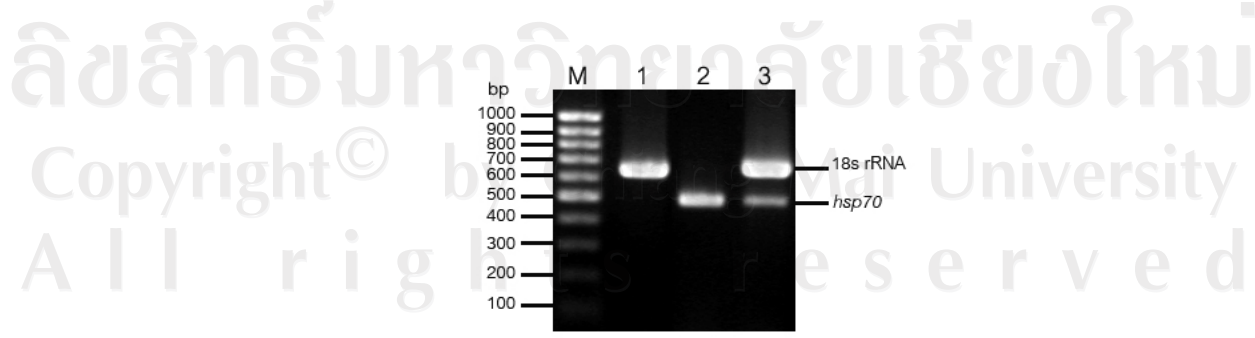


Figure C8. The amplified product of duplex RT-PCR with RRF1/RRH1 (18S rRNA) and specific primers for *hsp70*. The product of 18S rRNA is 630 bp (lane 1), 440 bp of *hsp70* is shown in lane 2 and mixed primers in lane 3.

C9. Primer for *P. marneffei cam* expression

PM-CAM	CTCTCTATTTGTAAGTTTGGATTCTGGTTGTCGCAATGTTGTGGTGGGTTGCTGA	60
PrimerF	-----CTGGTTGTCGCAATGTTGTG-----	20

PM-CAM	CTAGCCGTTTGGATGAATAGGACAAGGATGGTGATGGTGAGTGACGCCAGAACACCAGA	120
PM-CAM	CATATAGTCTTCGAACAAAAGTTATTACTGCGAACAGATATTAATAACATCAATAGGTC	180
PM-CAM	AAATTACAACCAAGGAACCTGGGCACCGTCATGCGTTCCTCGGCCAGAACCCTTCCGAAT	240
PM-CAM	CCGAATTGCAGGACATGATCAACGAGGTCGACGCTGACAACAACGGCACAATCGATTTCC	300
PM-CAM	CTGGTATGATGCAGCCTCTATTTATCGCAGCCGTTTCCGATCATAAGGGCAGATACTGAC	360
PM-CAM	TGCCTTAGAATTCTTGACAATGATGGCCCGCAAATGAAGGATACCGACTCCGAGGAAGA	420
PM-CAM	GATCCGCGAGGCTTTCAAGGTGTTTGATCGTGACAACAATGGATTTCATCTCTGCTGCTGA	480
	-----TGCTGA-----	

PM-CAM	ATTGCGCCACGTTATGACCTCGATTGGCGAAAAGTTGACCGACGACGAAGTTGACGAGAT	540
	ATTGCGCCACGTTATG-----	

C10. Primers for *P. marneffei cat* expression

PM-CAT	CAAAAAGACTCGCGGTCTTTCACAAATTGACTTCAAGCCTCGTGTACCAACTGTGGCCAG	420
PrimerF	-----TGTACCAACTGTGGCCAG-----	18

PM-CAT	CAGACGTATCGCCATAATCATTGGCGACGGCTTCGACTCTGTGCTTTTAAACGGAGTGTA	480
PrimerF	CAGA-----	22

PM-CAT	TACTGCCATCAAAGCTGCTGGCGCTCTGCCATTTGTCATCGGCACAAAGAGACAACCCAT	540
PM-CAT	CTTCGCAGATGGCGTTGACCCCCAGACCGAAATGGCGTAACACCCGAACCAATAACGA	600
PM-CAT	GGGTGTGCGTTCTACAATGTTTCGACGCTACTTTTCATCCCCGGTGGACCACGTCAGAC	660
PM-CAT	CCTTGCCAGGATCGGACGGATTGCCACTGGATAACAGAAACATTCGGTCACCTCAAAGC	720
PM-CAT	GCTCGGCGCAGAGGTGAGGCCGTCGATTTGTCAAGCAGGCGTTCTAGGTGTGGAGGC	780
PM-CAT	TGTACAATTTGCTAGTCAGAGTAGCACTGGTGTGCTTGAGTTCGTATGGCGTGGTTACTGC	840
PrimerR	-----AGCACTGGTGTGCTTGAGTCGT-----	22

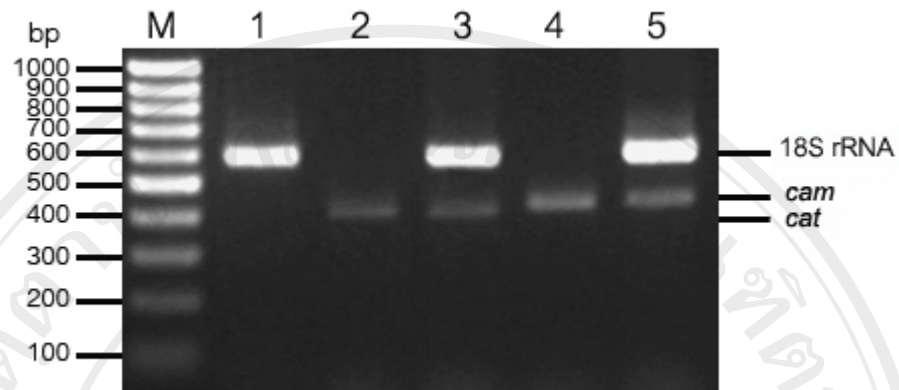


Figure C9-10. The amplified product of duplex RT-PCR with RRF1/RRH1 (18S rRNA) and specific primers for *cat* and *cam*. The product of 18S rRNA is 630 bp (lane 1), 420 bp of *cat* and 450 bp of *cam* are shown in lane 2 and 4, respectively. Duplex reactions of *cat* and *cam* are shown in lane 3 and 5, respectively.

C11. Primers for *cpeA* expression

cDNA
PrimerF GATACGCCGTCACAGTACCCTTCACTCCCGGTCGCACAGATGCTTCCCAAGAGCAGACTG 1800
-----CAAGAGCAGACTG 13

cDNA
PrimerF ACGTCCAATCCTTCAGCGACATGGAGCCATTGCTGATGGTTTCCGTAACACGGCTCAT 1860
ACGTCCA----- 20
***** →

cDNA CCACCTCTCGCGTTCGTGCTGAGGAGTGGCTCATCGATAAGGCACAGCTTTTGACCCTCA 1920

cDNA GTGCACCCGAGTTGGCCGTCTCATCGGCGGTCTCCGTGTCCTCAACACAAACTACGACG 1980

cDNA GCTCTGCTCACGGTGTCTTACCCAGCGCCAGGCAAGTTGACCAACGACTTCTTCGTCA 2040

cDNA ACCTCTTGACATGAACACCGCATGGAATCAATGGTGGTGTGACCTCTACGAGGGCA 2100

cDNA CAGATCGCAAGACTGGCGCCAAGAAGTGGACTGCTACTCGTAACGATCTCGTCTTTGGCT 2160

cDNA
PrimerR CCAACGCTGAGTTGCGTGCTATTGCTGAGGTGTACGGTAGCTCTGATGGCCAGGAGAAGT 2220
-----AGGTGTACGGTAGCTCTGATGG----- 22
←*****

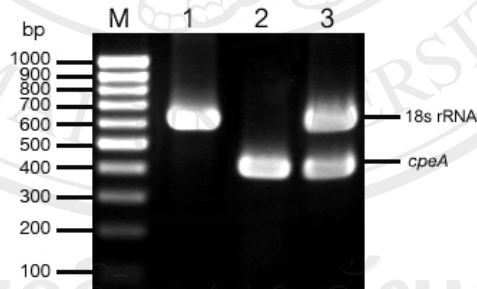


Figure C11. The amplified product of duplex RT-PCR with RRF1/RRH1 (18S rRNA) and specific primers for *cpeA*. The product of 18S rRNA is 630 bp (lane 1), 400 bp of *cpeA* is shown in lane 2 and mixed primers in lane 3.



APPENDIX D
STATISTIC ANALYSIS

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่

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Phagocytosis and killing assay

1. Percentage of phagocytosis

Table D-1 Paired Samples statistics of percentage of phagocytosis

	Mean	N	Std. Deviation	Std. Error Mean
Pair 1 PP30-PC	47.000	3	2.646	1.528
PP30-PM	49.667	3	4.726	2.728
Pair 2 PP60-PC	73.000	3	6.083	3.512
PP60-PM	72.333	3	3.512	2.028
Pair 3 PP120-PC	80.000	3	3.000	1.732
PP120-PM	90.667	3	9.292	5.364
Pair 4 PP240-PC	88.000	3	2.646	1.528
PP240-PM	93.667	3	7.506	4.333

Table D-2 Statistic significance of percentage of phagocytosis by Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)
	Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
				Lower	Upper			
Pair 1 PP30PC-PM	-2.667	6.506	3.756	-18.83	13.496	-0.710	2	0.551
Pair 2 PP60PC-PM	0.667	9.452	5.457	-22.81	24.146	0.122	2	0.914
Pair 3 PP120PC-PM	-10.667	7.095	4.096	-28.29	6.957	-2.604	2	0.121
Pair 4 PP240PC-PM	-5.667	8.737	5.044	-27.37	16.037	-1.123	2	0.378

PP = Percentage of phagocytosis, PM = *P. marneffeii*, PC = *P. citrinum*

2. Phagocytic index

Table D-3 Paired Samples statistics of phagocytic index

	Mean	N	Std. Deviation	Std. Error Mean
Pair 1 PI30 PC	1.643	3	0.068	0.039
PM	1.600	3	0.208	0.120
Pair 2 PI60 PC	2.093	3	0.071	0.041
PM	2.503	3	0.046	0.027
Pair 3 PI120 PC	2.310	3	0.301	0.174
PM	3.980	3	0.676	0.390
Pair 4 PI240 PC	2.660	3	0.298	0.172
PM	4.570	3	0.439	0.253

Table D-4 Statistic significance of phagocytic index by Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)
	Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
				Lower	Upper			
Pair 1 PI30PC-PM	0.043	0.167	0.096	-0.370	0.457	0.451	2	0.696
Pair 2 PI60PC-PM	-0.410	0.046	0.026	-0.524	-0.296	-15.497	2	0.004*
Pair 3 PI120PC-PM	-1.670	0.390	0.225	-2.638	-0.701	-7.422	2	0.018*
Pair 4 PI240PC-PM	-1.910	0.450	0.260	-3.028	-0.792	-7.352	2	0.018*

PI = Phagocytic index, PM = *P. marneffeii*, PC = *P. citrinum*, * $P < 0.05$

3. Percentage of killing

Table D-5 Paired Samples statistics of percentage of killing

	Mean	N	Std. Deviation	Std. Error Mean
Pair 1 PK30 PC	42.400	2	4.229	2.990
PM	7.225	2	1.223	0.865
Pair 2 PK60 PC	68.560	2	3.026	2.140
PM	16.550	2	6.053	4.280
Pair 3 PK120 PC	66.780	2	2.475	1.750
PM	38.845	2	6.937	4.905
Pair 4 PK240 PC	64.120	2	3.422	2.420
PM	61.710	2	1.541	1.090

Table D-6 Statistic significance of percentage of killing by Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)
	Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
				Lower	Upper			
Pair 1 PK30PC-PM	35.175	3.005	2.125	8.174	62.176	16.553	1	0.038*
Pair 2 PK60PC-PM	52.010	3.026	2.140	24.82	79.201	24.304	1	0.026*
Pair 3 PK120PC-PM	27.935	4.462	3.155	-12.153	68.023	8.854	1	0.072
Pair 4 PK240PC-PM	2.410	1.881	1.330	-14.489	19.309	1.812	1	0.321

PK = Percentage of killing, PM = *P. marneffeii*, PC = *P. citrinum*, * $P < 0.05$

Relative genes expression by RT-PCR analysis

4. Relative genes expression levels of *P. marneffei* in different phases

Table D-7 Paired Samples statistics of relative genes expression in different phases

		Mean	N	Std. Deviation	Std. Error Mean
<i>sodA</i>	Mycelium	0.190	2	0.085	0.060
	Conidia	0.420	2	0.085	0.060
	Yeast	0.635	2	0.120	0.085
<i>cpeA</i>	Mycelium	0.215	2	0.007	0.005
	Conidia	0.535	2	0.035	0.025
	Yeast	0.915	2	0.021	0.015
<i>hsp70</i>	Mycelium	0.675	2	0.007	0.005
	Conidia	0.695	2	0.021	0.015
	Yeast	0.755	2	0.007	0.005
<i>acuD</i>	Mycelium	0.435	2	0.092	0.065
	Conidia	0.450	2	0.042	0.030
	Yeast	0.490	2	0.057	0.040
<i>gapdh</i>	Mycelium	0.615	2	0.021	0.015
	Conidia	0.755	2	0.007	0.005
	Yeast	0.525	2	0.078	0.055

Table D-8 Statistic significance of relative genes expression in different phases by Paired Samples Test

		Paired Differences				t	df	Sig. (2-tailed)	
		Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
					Lower				Upper
<i>sodA</i>	Mycelium-Conidia	-0.230	0.170	0.120	-1.755	1.295	-1.917	1	0.306
	Conidia-Yeast	-0.215	0.035	0.025	-0.533	0.103	-8.600	1	0.074
	Mycelium-Yeast	-0.445	0.035	0.0250	-0.762	-0.127	-17.800	1	0.036*
<i>cpeA</i>	Mycelium-Conidia	-0.320	0.042	0.030	-0.701	0.061	-10.667	1	0.060
	Conidia-Yeast	-0.380	0.014	0.010	-0.507	-0.253	-38.000	1	0.017*
	Mycelium-Yeast	0.700	0.282	0.200	-0.954	0.446	-35.000	1	0.018*
<i>hsp70</i>	Mycelium-Conidia	-0.020	0.014	0.010	-0.147	0.107	-2.000	1	0.295
	Conidia-Yeast	-0.060	0.014	0.010	-0.187	0.067	-6.000	1	0.105
	Mycelium-Yeast	0.800	0.141	0.100	-0.047	0.207	8.000	1	0.079
<i>acuD</i>	Mycelium-Conidia	-0.015	0.050	0.035	-0.460	0.430	-0.429	1	0.742
	Conidia-Yeast	-0.040	0.014	0.010	-0.167	0.087	-4.000	1	0.156
	Mycelium-Yeast	-0.055	0.035	0.025	-0.372	0.262	-2.200	1	0.272
<i>gapdh</i>	Mycelium-Conidia	-0.140	0.028	0.020	-0.394	0.114	-7.000	1	0.090
	Conidia-Yeast	0.230	0.085	0.060	-0.532	0.992	3.833	1	0.162
	Mycelium-Yeast	0.090	0.056	0.040	-0.418	0.598	2.250	1	0.266

* $P < 0.05$

5. Relative genes expression levels of *P. marneffei* during macrophage infection

Table D-9 Paired Samples statistics of relative *sodA* expression during macrophage infection

	Mean	N	Std. Deviation	Std. Error Mean
Pair 1 Control 2 h	0.290	2	0.028	0.020
Infect 2 h	0.515	2	0.035	0.025
Pair 2 Control 4 h	0.510	2	0.014	0.010
Infect 4 h	0.675	2	0.007	0.005
Pair 3 Control 8 h	0.425	2	0.035	0.025
Infect 8 h	0.760	2	0.014	0.010

Table D-10 Statistic significance of relative *sodA* expression during macrophage infection by Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)
	Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
				Lower	Upper			
Pair 1 Ctl-Inf 2 h	-0.225	0.007	0.005	-0.289	-0.161	-45.000	1	0.014*
Pair 2 Ctl-Inf 4 h	-0.165	0.007	0.005	-0.229	-0.102	-33.000	1	0.019*
Pair 3 Ctl-Inf 8 h	-0.335	0.021	0.015	-0.526	-0.144	-22.333	1	0.028*

* $P < 0.05$

Table D-11 Paired Samples statistics of relative *cpeA* expression during macrophage infection

	Mean	N	Std. Deviation	Std. Error Mean
Pair 1 Control 2 h	0.065	2	0.007	0.005
Infect 2 h	0.630	2	0.000	0.000
Pair 2 Control 4 h	0.430	2	0.014	0.010
Infect 4 h	0.600	2	0.000	0.000
Pair 3 Control 8 h	0.445	2	0.021	0.015
Infect 8 h	0.560	2	0.000	0.000

Table D-12 Statistic significance of relative *cpeA* expression during macrophage infection by Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)
	Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
				Lower	Upper			
Pair 1 Ctl-Inf 2 h	-0.565	0.007	0.005	-0.6285	-0.5015	-113.00	1	0.006*
Pair 2 Ctl-Inf 4 h	-0.170	0.014	0.010	-0.2971	-0.0429	-17.000	1	0.037*
Pair 3 Ctl-Inf 8 h	-0.115	0.021	0.015	-0.3056	0.0756	-7.667	1	0.083

* $P < 0.05$

Table D-13 Paired Samples statistics of relative *hsp70* expression during macrophage infection

	Mean	N	Std. Deviation	Std. Error Mean
Pair 1 Control 2 h	0.740	2	0.156	0.110
Infect 2 h	0.825	2	0.219	0.155
Pair 2 Control 4 h	0.710	2	0.099	0.070
Infect 4 h	0.870	2	0.170	0.120
Pair 3 Control 8 h	0.550	2	0.042	0.030
Infect 8 h	0.705	2	0.163	0.115

Table D-14 Statistic significance of relative *hsp70* expression during macrophage infection by Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)
	Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
				Lower	Upper			
Pair 1 Ctl-Inf 2 h	-0.0850	0.064	0.045	-0.657	0.487	-1.889	1	0.310
Pair 2 Ctl-Inf 4 h	-0.1600	0.071	0.050	-0.795	0.475	-3.200	1	0.193
Pair 3 Ctl-Inf 8 h	-0.1550	0.120	0.085	-1.235	0.925	-1.824	1	0.319

Table D-15 Paired Samples statistics of relative *acuD* expression during macrophage infection

	Mean	N	Std. Deviation	Std. Error Mean
Pair 1 Control 2 h	0.270	2	0.000	0.000
Infect 2 h	0.295	2	0.064	0.045
Pair 2 Control 4 h	0.305	2	0.078	0.055
Infect 4 h	0.340	2	0.000	0.000
Pair 3 Control 8 h	0.400	2	0.071	0.050
Infect 8 h	0.395	2	0.007	0.005

Table D-16 Statistic significance of relative *acuD* expression during macrophage infection by Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)
	Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
				Lower	Upper			
Pair 1 Ctl-Inf 2 h	-0.025	0.064	0.045	-0.597	0.547	-.0556	1	0.677
Pair 2 Ctl-Inf 4 h	-0.035	0.078	0.055	-0.734	0.664	-0.636	1	0.639
Pair 3 Ctl-Inf 8 h	0.005	0.078	0.055	-0.694	0.704	0.091	1	0.942

Table D-17 Paired Samples statistics of relative *gapdh* expression during macrophage infection

	Mean	N	Std. Deviation	Std. Error Mean
Pair 1 Control 2 h	0.360	2	0.113	0.080
Infect 2 h	0.370	2	0.156	0.110
Pair 2 Control 4 h	0.395	2	0.191	0.135
Infect 4 h	0.550	2	0.156	0.110
Pair 3 Control 8 h	0.680	2	0.085	0.060
Infect 8 h	0.625	2	0.148	0.105

Table D-18 Statistic significance of relative *gapdh* expression during macrophage infection by Paired Samples Test

	Paired Differences					t	df	Sig. (2-tailed)
	Mean	Std. Deviation	Std. Error Mean	95% Confidence Interval of the Difference				
				Lower	Upper			
Pair 1 Ctl-Inf 2 h	-0.010	0.042	0.030	-0.391	0.371	-0.333	1	0.795
Pair 2 Ctl-Inf 4 h	-0.155	0.035	0.025	-0.473	0.163	-6.200	1	0.102
Pair 3 Ctl-Inf 8 h	0.055	0.064	0.045	-0.517	0.627	1.222	1	0.437



APPENDIX E
RECIPES

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่

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1. Culture Media

Brain heart infusion (BHI) agar

BHI agar (dehydrated)	52.00 g
Distilled water	1,000.00 ml

Melt, disperse in tubes and autoclave at 121 °C 15 lb/sq.in. for 15 min. Allow tubes to cool in slant position. Or, after autoclave, pour plates and allow cooling.

BHI broth

BHI	37.00 g
Distilled water	1,000.00 ml

Dissolve, disperse 30 ml in each 125 ml-Erlenmeyer flask. Sterile by autoclaving for 20 min at 15 lb/sq. in. on liquid cycle.

Luria-Bertani (LB) Medium

LB	20.00 g
Deionized H ₂ O up to	1,000.00 ml

Shake until the solutes are dissolved. Sterilize by autoclaving for 20 min at 15 lbs on liquid cycle. To make an agar medium add 20 g of agar to the 1 L solution and sterile by autoclave.

Sabouraud Dextrose Agar (SDA)

Dehydrated SDA agar (Becton Dickinson)	65.00 g
Distilled water	1,000.00 ml

Suspend 65 g of the powder in 1 liter of distilled water. Mix thoroughly, heat with frequent agitation and boil for 1 min to completely dissolve the powder. Autoclave at 121 °C 15 lbs. for 15 min.

SOC medium

Per liter:

To 950 ml of deionized water, add:

Tryptone	20	g
Yeast extract	5	g
NaCl	0.5	g

Shake until the solutes are dissolved. Adjust the pH to 7.0 with 5 N NaOH. Adjust the volume of the solution to 980 ml with the deionized water. Sterile by autoclaving, allow it to cool to 60°C or less and then add 20 ml of a sterile 1 M solution of glucose.

The glucose solution is made by dissolving 18 g of glucose in 90 ml of deionized water. After the sugar is dissolved, adjust the volume of the solution to 100 ml and sterile by filtration through a 0.2- μ m filter.

2. Solutions for working with bacteriophage λ **20 % Maltose**

Maltose	20	g
H ₂ O to	100	ml

Sterilize the solution by filtration through a 0.2- μ m filter. Store the sterile solution at room temperature.

SM

Per liter:

NaCl	5.8	g
MgSO ₄ • 7H ₂ O	2	g
1 M Tris-Cl (pH 7.5)	50	ml
2 % gelatin solution	5	ml
H ₂ O to	1	L

Sterilize the buffer by autoclaving for 20 min at 15 lb/sq.in. on liquid cycle. After the solution is cooled, dispense 50-ml aliquots into sterile containers. SM may be stored indefinitely at room temperature.

A 2 % gelatin solution is made by adding 2 g of gelatin to a total volume of 100 ml of H₂O and autoclaving the solution for 15 min at 15 lb/sq.in. on liquid cycle.

10 mM MgSO₄ (λ diluent)

Per liter:

1 M Tris-Cl (pH 7.5)	10	ml
MgSO ₄ • 7H ₂ O	2	g
H ₂ O to	1	L

Sterilize the buffer by autoclaving for 20 min at 15 lb/sq.in. on liquid cycle. After the dilution has cooled, dispense 50-ml aliquots into sterile containers. λ diluent may be stored indefinitely at room temperature.

3. Alkaline lysis buffers for minipreparation of plasmid DNA

Solution I

50 mM glucose
25 mM Tris-Cl (pH 8.0)
10 mM EDTA (pH 8.0)

Sterilize by autoclaving for 20 min at 15 lb/sq.in. on liquid cycle and store at 4°C.

Solution II

0.2 N NaOH (freshly diluted from a 10 N stock)
1 % SDS

Solution III

5 M potassium acetate	60	ml
Glacial acetic acid	11.5	ml
H ₂ O	28.5	ml

The resulting solution is 3 M with respect to potassium and 5 M with respect to acetate.

4. Reagents

50X Tris-acetate (TAE) buffer

Tris base	242	g
Glacial acetic acid	57.1	ml
0.5 M EDTA	100	ml

The working solution (1X) contains 0.04 M Tris acetate and 0.0001 M EDTA

6X agarose gel-loading buffer

Bromophenol blue	0.025	g
Xylene cyanol FF	0.025	g
Glycerol	3	ml
Distilled water	6.95	ml

5. Tissue culture medium

ATCC complete growth medium

Dulbecco's modified Eagle's medium (4 mM L-glutamine and 4.5% g/l glucose)	90%
Fetal bovine serum	10%

Freeze medium

Complete growth medium	85%
Fetal bovine serum	10%
DMSO	5%

Storage in liquid nitrogen vapor phase

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