TABLE OF CONTENTS

	Page
ACKNOWLEDMENT	iii
ENGLISH ABSTRACT	iv
THAI ABSTRACT	vi
LIST OF TABLES	xi
LIST OF FIGURES	xii
ABBREVIATIONS	xiv
CHAPTER I INTRODUCTION	22.1
1.1 Statement of the problems	1
1.2 Literature reviews	4
1.2.1 Multistep chemicals carcinogenesis	4
1.2.2 Carcinogenicity test	8
1.2.3 Diethylnitrosamine and hepatocarcinogenesis	13
1.2.4 Cancer chemoprevention and carcinogenesis	14
1.2.5 Xenobiotic-metabolizing enzymes	16
1.2.6 Effect of flavanones on xenobiotic-metabolizing enzymes	24
1.2.7 Pinocembrin	27
1.3 Objectives	28
CHAPTER II MATERIALS AND METHODS	29
2.1 Chemicals and instruments	29
2.2 Animals	29
2.3 Extraction and isolation of pinocembrin from <i>Boesenbergia</i>	29
pandurata (Roxb.) Schltr. rhizome.	
2.4 Mutagenicity study of pinocembrin in rat liver	30

2.5 Partial hepatectomy	32
2.6 Isolation of hepatocytes	32
2.7 Microscopic observation and micronucleus determination	33
2.8 Inhibitory effect of pinocembrin on diethylnitrosamine-	33
induced micronucleated hepatocyte formation in rat	
2.9 Preventive effect of pinocembrin on 30 mg/kg bw of	34
diethylnitrosamine-induced micronucleated hepatocyte formation in rat	
2.10 Protective effect of pinocembrin on 20 mg/kg bw of	34
diethylnitrosamine-induced micronucleated hepatocyte formation in rat	
2.11 Effect of pinocembrin on promotion stage in diethylnitrosamine-	38
induced rat hepatocarcinogenesis	
2.12 Immunohistochemistry for GST-P	S 39
2.13 Quantitative assessment of GST-P positive foci	39
2.14 Determination of lipid peroxidation by thiobarbituric acid	42
reactive substances assay	
2.15 Determination of the expression and activities of phase I and phase II	43
xenobiotic-metabolizing enzymes	
2.16 Statistical analysis	50
CHAPTER III RESULTS	51
3.1 Mutagenicity study of pinocembrin in rat liver	51
3.2 Effect of pinocembrin on lipid peroxidation in rat liver	51
3.3 Effect of pinocembrin on xenobiotic-metabolizing enzymes	52
a d a in rat liver	
3.4 Inhibitory effect of pinocembrin on diethylnitrosamine-induced	57
micronucleated hepatocyte formation in rat	
3.5 Preventive effect of pinocembrin on diethylnitrosamine-	57
induced micronucleus formation in rat liver	

Page

3.6 Effect of pinocembrin on promotion stage in diethylnitrosamine-	62
induced rat hepatocarcinogenesis	
CHAPTER IV DISCUSSION AND CONCLUSION	69
REFERENCES	74
APPENDICES	89
APPENDIX A	90
APPENDIX B	93
APPENDIX C	95
APPENDIX D	102
CURRICULUM VITAE	106

ລິບສິກລິ້ມหາວົກຍາລັຍເชีຍວໃหม Copyright[©] by Chiang Mai University All rights reserved

Х

LIST OF TABLES

Table		Page
3-1	Mutagenicity of pinocembrin in rat liver	53
3-2	Effect of pinocembrin on the expression of cytochrome P450	55
	isoenzymes and cytochrome P450 reductase in rat liver.	
3-3	Effect of pinocembrin on the activities of some phase I and	56
	phase II enzymes in rat liver	
3-4	Inhibitory effect of pinocembrin on diethylnitrosamine-induced	59
	micronucleus formation in rat liver	
3-5	Preventive effect of pinocembrin on 30 mg/kg bw of	60
	diethylnitrosamine-induced micronucleated hepatocyte	
	formation in rat	
3-6	Protective effect of pinocembrin on 20 mg/kg bw of	61
	diethylnitrosamine-induced micronucleus formation in rat liver	
3-7	General appearance of rats in medium-term carcinogenicity	64
	experiment	
3-8	Relative organ weight of rats in medium-term carcinogenicity	65
	experiment	
3-9	Blood biochemicals analysis of rats in medium-term	66
	carcinogenicity experiment	
3-1	0 Lipid peroxidation of rats in medium-term carcinogenicity	67
	experiment	
3-1	1 Number and the distribution of size of GST-P positive foci of rats	68
	in medium-term carcinogenicity experiment	

LIST OF FIGURES

Figure	Page
1-1 Overview of genotoxic and non-genotoxic effects of carcinogens	7
1-2 Mechanisms of multistage carcinogenesis	7
1-3 A schematic diagram show the origin of micronucleus	9
1-4 Standard protocol of the medium-term liver bioassay	10
1-5 Hypothetical model for the development of GST-P positive lesions	12
1-6 Regulation of the GST-P gene expression in normal liver cells	12
and in the pre-neoplastic lesion	
1-7 Biotransformation of diethylnitrosoamine and mechanism of	13
DNA-adduct formation	
1-8 Role of dietary detoxifying enzyme inducers in chemoprevention	15
1-9 Major detoxification activities in drug metabolism	17
1-10 The microsomal NADPH-cytochrome P450 reductase system	18
1-11 The pathway of heme degradation in mammalian cells	19
1-12 NADPH: quinone oxidoreductase 1	21
1-13 Consequences of quinone metabolism	21
1-14 Role of UDP-glucuronyltransferase	22
1-15 Role of glutathione-S-transferase	23
1-16 The basic structure of flavonoids	25
1-17 Chemical structures of some representative flavanone	25
1-18 Flavanones that block or suppress multistage carcinogenesis	26
1-19 Structure of pinocembrin	27
2-1 The protocol for mutagenicity study of pinocembrin in male Wistar rat	31

xii

Figure

Page

2-2	The protocol for inhibitory effect of pinocembrin on DEN-induced	35
	micronucleus formation in rat liver	
2-3	The protocol for preventive effect of pinocembrin on 30 mg/kg bw	36
	of DEN-induced micronucleus formation in rat liver	
2-4	The protocol for protective effect of pinocembrin on 20 mg/kg bw	37
	of DEN-induced micronucleus formation in rat liver	
2-5	The protocol for the effect of pinocembrin on promotion stage in	40
	diethylnitrosamine-induced rat hepatocarcinogenesis	
2-6	The procedure of GST-P immunohistochemistry in rat liver	41
2-7	MDA-TBA adduct	42
2-8	The preparation of microsomal and cytosolic fractions obtained	44
	from rat liver	
2-9	SDS-PAGE and Western blot procedures	46
3-1	Effect of pinocembrin on lipid peroxidation	54
3-2	Western blot analysis of liver microsomes from rat treated	54
	with various doses of pinocembrin.	
3-3	Growth curve of rats in medium-term carcinogenicity experiment	63
S-1	Extraction scheme of Boesenbergia pandurata	103
S-2	Isolation scheme of pinocembrin	104

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่ Copyright[©] by Chiang Mai University All rights reserved

ABBREVIATIONS

β- NADPH β -nicotinamide adenine nucleotide phosphate (Reduced form) °C degree celcius microgram μg μl microliter μΜ micromolar micrometer μm A Ampere ALP alkaline phosphatase ALT alanine aminotransferase AST aspartate aminotransferase BSA bovine serum albumin body weight bw CDNB 1-Chloro-2, 4-dinitrobenzene CuSO₄·5H₂O copper sulphate pentahydrate CYP cytochrome P450 3, 3'-diaminobenzidine DAB 2, 6-dichlorophenolindophenol DCPIP DEN diethylnitrosamine deionized water DI dithiothreitol DTT DW distilled water ethylenediaminetetraacetic acid EDTA ethylene glycol tetraacetic acid EGTA

xiv

FAD	flavin adenine dinucleotide
g	gram
GSH	glutathione (Reduced form)
GST-P	glutathione-S-transferase placental form
H ₂ O ₂	hydrogen peroxide
нсі	hydrochloric acid
i.g.	intragastium
i.p.	intraperitoneum
IgG	immunoglobulin G
KCI	potassium chloride
KCN	potassium cyanide
kg	kilogram
KH ₂ PO ₄	potassium dihydrogen phosphate
кон	potassium hydroxide
L	liter
м	molar
mA/cm ²	milli ampere per square centimeter
MDA	malondialdehyde bis(dimethyl acetal)
mg	milligram
MgCl ₂	magnesium chloride
MgCl ₂ min	minute
ml	milliliter
mm	millimeter
mM	milli molar
MNHEPs	micronucleated hepatocytes
NaCl	sodium chloride
NaHCO ₃	sodium bicarbonate
NaH ₂ PO ₄	sodium dihydrogen phosphate
Na ₂ HPO ₄	sodium hydrogen phosphate
Na ₂ CO ₃	sodium carbonate
NaOH	sodium hydroxide

xv

NSS	normal saline solution
nm	nanometer
PBS	phosphate buffer saline
PC	pinocembrin
РН	partial hepatectomy
PMSF	phenylmethanesulphonylfluoride
SDS-PAGE	sodium dodecyl sulfate polyacrylamide
	gel electrophoresis
ТВА	thiobarbituric acid
TBARS	thiobarbituric acid reactive
	substances
TCA	trichloroacetic acid
v/v	volume by volume
w/v	weight by volume

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่ Copyright[©] by Chiang Mai University All rights reserved

xvi