

## APPENDIX A

### INSTRUMENTS

**Instruments used in this study were listed as following:**

- Analytical balance (Adventurer-ohaus AR 2130, USA)
- Blotting module (Amersham Biosciences, USA)
- CAMAG UV Lampe (CAMAG, German)
- Centrifuge (Kubota 5200, Japan)
- Electrophoresis module (Amersham Biosciences, USA)
- Eppendorf centrifuge 5417R (Eppendorf, Germany)
- Magnetic stirrer (Thermolyne Co., USA.)
- pH meter. Model 3560 (Beckman, USA)
- Power supply (Amersham Biosciences, USA)
- Roller mixer SRT1 (Stuart scientific, UK)
- Scan densitometer (BioRad, USA)
- Ultrasonic Processor UP50H (Hielscher, German)
- Vortex mixture, (Scientific Industries, New York, USA.)
- Water bath (Mettler)

**APPENDIX B**  
**CHEMICALS AND REAGENTS**

All chemicals (essential) used were analytical grade:

- Acrylamide (Sigma Chemical Co., USA. No. A 8887)
- Amersham Full-Range Rainbow Molecular Weight Markers (GE Healthcare, UK. RPN800E)
- Ammonium persulfate (Bio-Rad Laboratories, No. 161-0700)
- Anti-GAPDH antibody [6C5] (Abcam, Japan. No. ab8245)
- BCA protein assay kit (PIERCE, USA. No.23227).
- Boric acid (Sigma Chemical Co., USA. No. B 7901)
- Brilliant blue R-250 (Biorad Chemical Company, USA.)
- Bromophenol blue (Sigma Chemical Co., USA. No. B5225)
- ECL Plus detection kit (Amersham Biosciences, UK.)
- Ethanol (Merck Darmstadt, Germany. No. K 32820483 402)
- Glycine (Sigma Chemical Co., USA. No. G8898)
- Hydrochloric acid (Merck Darmstadt, Germany)
- Lyophilized rabbit polyclonal antibody p53 protein (CM5) (Novocastra, UK. No. NCL-p53-CM5p)
- 2-mercaptoethanol (Sigma Chemical Co., USA. No. M4125)
- Methanol (Merck Darmstadt, Germany)

- Mouse Monoclonal Anti-GRP94 (2H3) (Santacruz, inc., USA. No. sc-53929)
- N,N'-Methylene-bis-acrylamide (sigma Chemical Co., USA. No. M 7256)
- N,N,N',N'-tetramethylethylenediamine: TEMED (Sigma Chemical Co., USA. No. T8133)
- p53 protein (DO-7) liquid mouse monoclonal antibody (Novocastra, UK. No. NCL-L-p53-DO7 )
- Peroxidaase-Conjugated Rabbit Anti-Rat Immunoglobulins (DakoCytomation, Denmark. No. P0450)
- Polyclonal Goat Anti-Mouse Immunoglobulins/HRP (DakoCytomation, Denmark. No. P0447)
- Polyclonal Goat Anti-Rabbit Immunoglobulins/HRP (DakoCytomation, Denmark. No. P0448)
- Protease inhibitor cocktail tablets (Complete, Mini, EDTA-free, Roche Applied Science, Germany. Cat. No. 11836170001)
- Protein G Agarose (Pierce, USA. No. 20398)
- Rabbit polyclonal antibody Glucosidase II $\beta$  (H-195) (Santacruz, inc., USA. No. sc-10774)
- Rabbit TrueBlot: Anti-Rabbit IgG HRP (eBioscience, inc., USA. Cat. No. 18-8816)
- Rat Monoclonal Anti-GRP78 (76-E6) (Santacruz, inc., USA. No. sc-13539)
- SuperSignal west pico chemiluminescent substrate (Thermo scientific, USA. No.34080)

- Tris [hydroxymethyl]-aminomethane hydrochloride (Merck, Darmstadt, Germany. No. 648311)
- Tunicamycin from *Streptomyces* sp. (Sigma-aldrich, USA. No. T7765)
- Tween20 (Fluke, Germany. No. 93773)
- Other chemicals were analytical grade

## APPENDIX C

### REAGENT PREPARATIONS

#### I. Reagents for preparation of protein from tissues and cell lines

##### 1. SDS lysis buffer

0.5 M Tris pH 6.8	2.5	ml
SDS	0.8	g
Glycerol	2	ml

Adjust volume to 10 ml with deionized distilled water and store at -20 °C.

##### 2. Nuclear isolation buffer (NIB)

Sucrose	4.27	g
NP-40	0.125	g
EDTA	0.093	g
NaCl	0.29	g
Tris	0.061	g

Adjust to pH 7.5 with HCl.

Make up to 50 ml with deionized distilled water and stored at -20 °C.

## II. Reagents for electrophoresis on polyacrylamide gel

### 1. Separating gel buffer stock (1.5 M Tris-HCl pH 8.8)

Tris	36.3	g
ddH <sub>2</sub> O	150	ml

Adjust to pH 8.8 with HCl.

Make up to 200 ml with deionized distilled water and store at 4 °C.

### 2. Stacking gel buffer stock (0.5 M Tris-HCl pH 6.8)

Tris	6	g
ddH <sub>2</sub> O	60	ml

Adjust to pH 6.8 with HCl.

Make up to 100 ml with deionized distilled water and stored at 4 °C.

### 3. 10% APS

Ammonium persulfate	20	mg
ddH <sub>2</sub> O	200	μl

Mix together. (Prepared before using)

### 4. 2X Sample buffer

SDS	0.8	g
0.5 M tris-HCL, pH 6.8	2.5	ml
Glycerol	2	ml
0.5% bromophenol blue	2	mg

Make up to 10 ml with deionized water and stored at -20 °C

## 5. 30% Acrylamide stock solution

Acrylamide	150	g
Bis-acrylamide	4.0	g

Total volume 500 ml with deionized water mix and store at 4 °C

**III. Reagents for SDS-PAGE and western blot analysis**

## 1. 7% Separating gel solution

ddH <sub>2</sub> O	7.5	ml
30% Acrylamide stock solution	3.5	ml
Separating gel buffer stock	3.75	ml
10% SDS	150	μl
10% ammonium persulfate	100	μl
TEMED	20	μl

Swirl gently to mix and pour the solution into the gel cassette.

## 2. 4% Stacking gel solution

ddH <sub>2</sub> O	3	ml
30% Acrylamide stock solution	665	μl
Stacking gel buffer stock	1.25	ml
10% SDS	50	μl
10% ammonium persulfate	50	μl
TEMED	10	μl

Swirl gently to mix and pour the solution into the gel cassette.

## 3. 10X Running buffer

Tris-base	30.3	g
Glycien	144	g
SDS	10	g

Make up to 1L with deionized water and stored at room temperature.

## 4. 1X Running buffer

10X Running buffer	100	ml
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Make up to 1L with deionized distilled water.

## 5. 10X Transfer buffer

Tris-base	30.3	g
Glycien	144	g

Make up to 1L with deionized distilled water and stored at room temperature.

## 6. 1X Transfer buffer

10X Transfer buffer	100	ml
Methanol	200	ml

Make up to 1L with deionized distilled water.



## 7. Washing buffer (0.05% TBS-Tween)

Tris	6	g
NaCl	9	g
dH <sub>2</sub> O	600	ml
Adjust to pH 7.5 with HCl.		
Tween 20	0.5	ml

Make up to 1L with deionized distilled water and stored at room temperature.

## 8. Stripping buffer

Tris	3.78	g
SDS	10	g
dH <sub>2</sub> O	400	ml

Adjust to pH 6.7 with HCl.

Make up to 500 ml with deionized distilled water and stored at room temperature.

Add 2-mercaptoethanol (2-ME) 7  $\mu$ l/1 ml before using.

## CURRICULUM VITAE

**Name** Miss Benjamart Suradej

**Date of Birth** April 21, 1986

### **Institution attended**

- Kowittamrong Chiangmai School, March 1997  
: Elementary Certificate
- Doisaketwittayakom School, March 2000  
: Primary high school Certificate
- Yupparajwittayalai School, March 2003  
: High school Certificate
- Chiang Mai university, March 2007  
: B. Sc (Medical Technology)
- Faculty of Associated Medical Sciences, Chiang Mai university, May 2009 –  
present  
: M. Sc Student in Medical Technology

### **Poster presentation**

- Suradej B., Lertprasertsuke N., Cressey R. The Association of matrix metalloproteinase-9 (MMP-9) gene polymorphism and cigarette smoking on the risk of lung cancer in Northern Thai population. Graduate school, Chiang Mai University, 2007

**Oral presentation/Proceeding**

- Suradej B., Lertprasertsuke N., Kasinrerker W., Cressey R. Overexpression of endoplasmic reticulum glucosidase II in non-small lung cancer: a novel biomarker for diagnosis. The 1<sup>st</sup> Asean Plus Three Graduate Research Congress (AGRC 2012), the Empress Hotel, Chiang Mai, Thailand 1-2 March 2012

**Publication**

- Suradej B., Pata S., Lertprasertsuk N., Kasinrerker W and Cressey R. Similarity of structure and behavior in response to endoplasmic reticulum (ER) stress and UV irradiation of glucosidase II to p53 tumor suppressor: a potential novel cancer biomarker (2011). **Cancer Investigation**, under review

**Honors and Awards**

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|------|---|
| 2012 | Excellent Oral Presentation at The 1st ASEAN Plus Three Graduate Research Congress (AGRC 2012) at The Empress Hotel, Chiang Mai, Thailand |
| 2007 | Good Poster Presentation at Faculty of Associated Medical Sciences, Chiang Mai University   |