

CHAPTER 3

MATERIALS AND METHODS

3.1 Sampling sites

Investigation on *Pediastrum* spp. in standing freshwater was carried out during June 2011- May 2012. In total, 204 water samples from 68 sampling sites including lakes, dams, reservoirs, pond and ditch from 48 provinces were collected in this study (Figures 3 and 4). There were 8 sampling sites in the northern region, 18 in the central region, 20 in the north-eastern region, 6 in the eastern region, 6 in the western region and 10 in the southern region. The ecological data of each sampling site were recorded i.e. latitude and longitude and utilization of sampling site such as forest, agriculture and community. The list of sampling sites together with some general data are given in Table 2.

Table 2 Location and some characteristics of sampling sites

Sampling site	Code	Size	Ordinations	
1. Chiang Rai				
1.1 Canal (roadside)	CHR1	M	99°49'27"E	19°54'28"N
2. Chiang Mai				
2.1 Pond (Wat Umong)	CHM1	S	100°07'55"E	15°42'11"N
2.2 Pond (700 th Anniversary Chiang Mai Sports Complex)	CHM2	S	100°07'55"E	15°42'11"N
2.3 Chiang Mai Moat	CHM3	L	98°58' 770"E	18°46' 900"N
3. Phayao				
3.1 Ang Leung reservoir	PHY1	M	99°53'45"E	19°01'37"N
3.2 Pond (in Phayao University)	PHY2	S	99°53'31"E	19°01'58"N
4. Phrae				
4.1 Pond (roadside)	PHR1	S	100°00'57"E	17°48'03"N

Size of reservoir was characterized from its capacity

S= capacity of reservoir <1 million m³ and/or surface area of reservoir < 1km²

M= capacity of reservoir >1 million m³ and <100 million m³ and/or surface area of reservoir > 1km² and <15 km²

L= capacity of reservoir ≥100 million m³ and/or surface area of reservoir >15 km²

Table 2 (Continued)

Sampling site	Code	Size	Ordinations	
5. Uttaradit				
5.1 Pond (roadside)	UTD1	S	9°57'25"E	17°34'44"N
6. Sukhothai				
6.1 Pond (Wat Traphang Tong)	SKT1	L	99°42'32"E	17°01'06"N
7. Phitsanulok				
8.1 Pond (near Latina Hotel)	PSL1	S	100°16'33"E	16°45'23"N
8. Nakhon Sawan				
8.1 Nong Somboon	NKS1	L	100°07'55"E	15°42'11"N
8.2 Pond (in University of Central Thailand)	NKS2	S	100°06'50"E	15°41'54"N
9. Phichit				
9.1 Fish pond	PHC1	S	100°24'2.5"E	16°10'58"N
10. Phetchabun				
10.1 Huai Pa Dang Reservoir	PCB1	L	101°05'14"E	16°26'48.6"N
10.2 Pond (in Provincial Waterworks Authority)	PCB2	S	101°8'53.7"E	16°26'9.7"N
11. Saraburi				
11.1 Canal Pure (near restaurant)	SRB1	L	100°55'50.6"E	14°31'40"N
12. Pathum Thani				
12.1 Pond (Country Place Resort Club)	PTT1	S	100°45'00.4"E	14°02'43.5"N
13. Samut Prakan				
13.1 Canal (roadside)	SMP1	L	100°36'35.5"E	13°35'17.9"N
14. Phra Nakhon Si Ayutthaya				
14.1 Pond (in Red Cross Office)	PNS1	S	100°34'3.2"E	14°21'1.5"N
15. Sing Buri				
15.1 Pond (in restaurant)	SBR1	S	100°24'26.3"E	14°51'37.6"N
16. Ang Thong				
16.1 Canal (roadside)	ANT1	M	100°27'12.9"E	14°29'33.7"N
17. Suphan Buri				
17.1 Pond (in front of the Uthong National Museum Suphanburi)	SPB1	M	99°53'30"E	14°22'16"N
18. Uthai Thani				
18.1 Pond (Karung City Police Station)	UTT1	S	99°41'49"E	15°15'31"N
18.2 Bueng Reusi	UTT2	M	100°7'46"E	15°30'2"N
19. Nakhon Pathom				
19.1 Concrete pond (roadside)	NPT1	S	99°58'40"E	13°47'52"N
19.2 Lad Po Marshes	NPT2	S	99°58'40"E	13°47'52"N
20. Loei				
20.1 Pond (in Phurua Hospital)	LOE1	S	99°49'37"E	18°28'27"N
21. Udon Thani				
21.1 Nong Prajak Park	UDT1	M	102°46'47"E	17°25'11"N
22. Nong Khai				
22.1 Pond (roadside)	NKI1	S	103°04'41"E	18°01'07"N
23. Nakhon Phanom				
23.1 Pond (roadside)	NPN1	S	104°43'425"E	17°23'25"N
23.2 Pond (in Nakhon Phanom University)	NPN2	S	104°43'08"E	17°23'50"N

Table 2 (Continued)

Sampling site	Code	Size	Ordinations	
24. Sakon Nakhon				
24.1 Municipal Oxidation Pond	SKN1	S	104°10'09"E	17°09'48"N
25. Mukdahan				
25.1 Pond (in Chaloe Phra Kiat Kanchana Phisek Park)	MDH1	S	104°43'16"E	16°32'32"N
26. Yasothon				
26.1 Canal (in Shithummaram Temple)	YST1	S	104°08'27"E	15°47'06"N
27. Roi Et				
27.1 Pond (roadside)	RET1	M	104°08'27"E	15°47'06"N
27.2 Phalanchai Lake	RET2	M	103°38'55"E	16°03'28"N
28. Kalasin				
28.1 Pond (in Kud Nam Kin Park)	KLS1	M	103°30'00"E	16°25'20"N
29. Khon Kaen				
29.1 Pond (roadside)	KKN1	L	102°50'58"E	16°26'45"N
29.2 Concrete pond (roadside)	KKN2	M	102°50'25"E	16°26'48"N
30. Nakhon Ratchasima				
30.1 Pond (roadside)	NRS1	S	102°15'17"E	14°58'47"N
30.2 Canal (roadside)	NRS2	M	102°06'08"E	14°58'34"N
31. Surin				
31.1 Pond (roadside)	SUR1	S	103°32'11"E	14°52'40"N
31.2 Fish pond (in temple)	SUR2	S	103°32'27"E	14°52'41"N
32. Si Sa Ket				
32.1 Canal (in Somdech Phra Srinagarindra Park)	SSK1	M	104°18'24"E	15°06'10"N
32.2 Pond (Sisaket Marenon Can Park)	SSK2	S	104°19'09"E	15°06'16"N
33. Ubon Ratchathani				
33.1 Pond (Ubon Ratchathani Cultural Center)	UBR1	S	104°50'44"E	15°14'47"N
34. Chachoengsao				
34.1 Pond (in Somdech Phra Srinagarindra Park)	CCS1	S	101°04'05"E	13°41'17"N
35. Chon Buri				
35.1 Pond (in Burapha University)	CBR1	S	105°55'36"E	13°16'36"N
36. Rayong				
36.1 Pond (in Phraphuttha Angkhirot Dhamma Hall)	RYN1	S	101°16'37"E	12°40'30"N
36.2 Klong (roadside)	RYN2	M	101°16'38"E	12°40'29"N
37. Chanthaburi				
37.1 (in King Taksin the Great Park)	CTB1	L	102°06'11"E	12°36'23"N
38. Sa Kaeo				
38.1 Pond (in Sa Kaeo Hospital)	SKO1	S	102°04'20"E	13°48'59"N
39. Tak				
39.1 Pond (in Tak Municipal Wastewater Treatment)	TAK1	M	99°7'51"E	16°51'35"N

Table 2 (Continued)

Sampling site	Code	Size	Ordinations	
40. Kanchanaburi				
40.1 Pond (Chaloem Prakiarti Rama 9 Park)	KCN1	M	99°45'1"E	13°57'13"N
40.2 Pond (in Phanom Thuan Provincial Waterworks Authority)	KCN2	M	99°41'16"E	14°7'21"N
43.3 Fish pond	KCN3	S	99°43'26"E	14°9'57"N
41. Ratchaburi				
41.1 Concrete pond (in Jompratad Health Promoting Hospital)	RBR1	S	99°52'51"E	13°24'34"N
42. Phetchaburi				
42.1 Pond (roadside)	PBR1	S	99°55'4"E	12°52'7"N
43. Chumphon				
43.1 Pond (in Chumphon Khet Udomsakdi Hospital)	CHP1	S	99°11'11"E	10°29'56"N
44. Surat Thani				
44.1 Pond (in temple)	SRT1	S	99°11'05"E	9°23'06"N
44.2 Pond (in Rama 9 Public park)	SRT2	M	99°11'05"E	9°08'19"N
44.3 Pond (in Ban Sadet Subdistrict Administrative Organization)	SRT3	M	99°08'03"E	9°00'19"N
45. Nakhon Si Thammarat		S		
45.1 Pond (in Somdech Phra Srinagarindra Park)	NST1	L	99°57'14"E	8°27'17"N
45.2 Pond (roadside)	NST2	S	99°55'21"E	8°30'46"N
45.3 Pond (in Nakhon Si Thammarat Airport)	NST3	S	99°56'24"E	8°32'31"N
46. Phatthalung				
46.1 Pond (roadside)	PTL1	S	100°08'53"E	7°38'35"N
47. Songkhla				
47.1 Pond Hatyai City Municipality Park	SKA1	M	100°30'17"E	7°02'33"N
48. Satun				
48.1 Pond (roadside)	STN1	S	100°02'59"E	6°37'59"N

3.2 Investigation of *Pediastrum* spp.

3.2.1 Collection of *Pediastrum* spp.

Pediastrum spp. were collected by filtering 10 liters of water from each sampling site with 10 µm pore size plankton net. The samples were preserved by adding 0.7 ml of lugol solution to 100 ml of samples (Wetzel, 2001) and the fresh sample were kept in a cool box for isolation and cultivation.

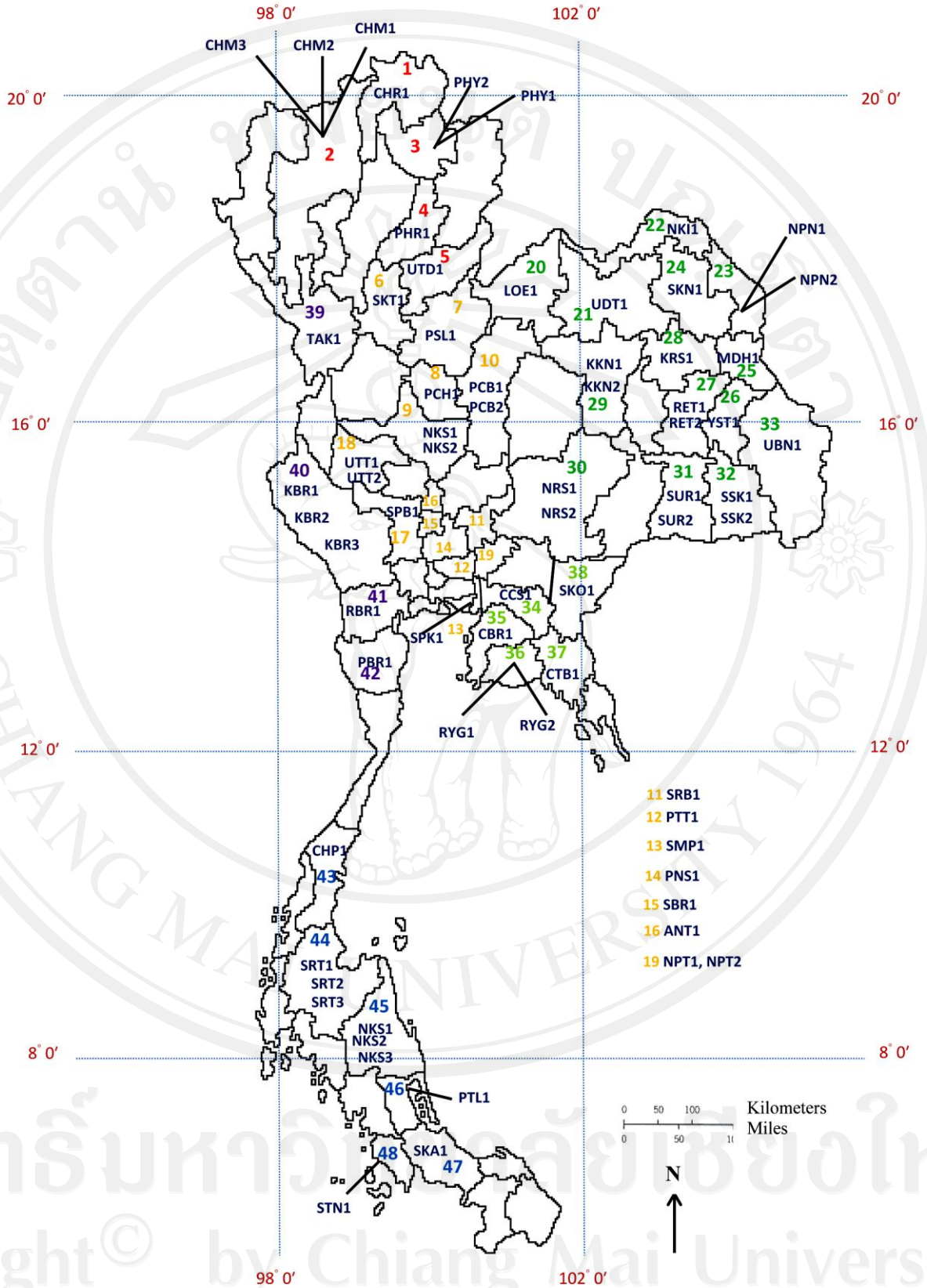


Figure 3 Map of Thailand showed 68 sampling sites in 48 provinces in some freshwater resources

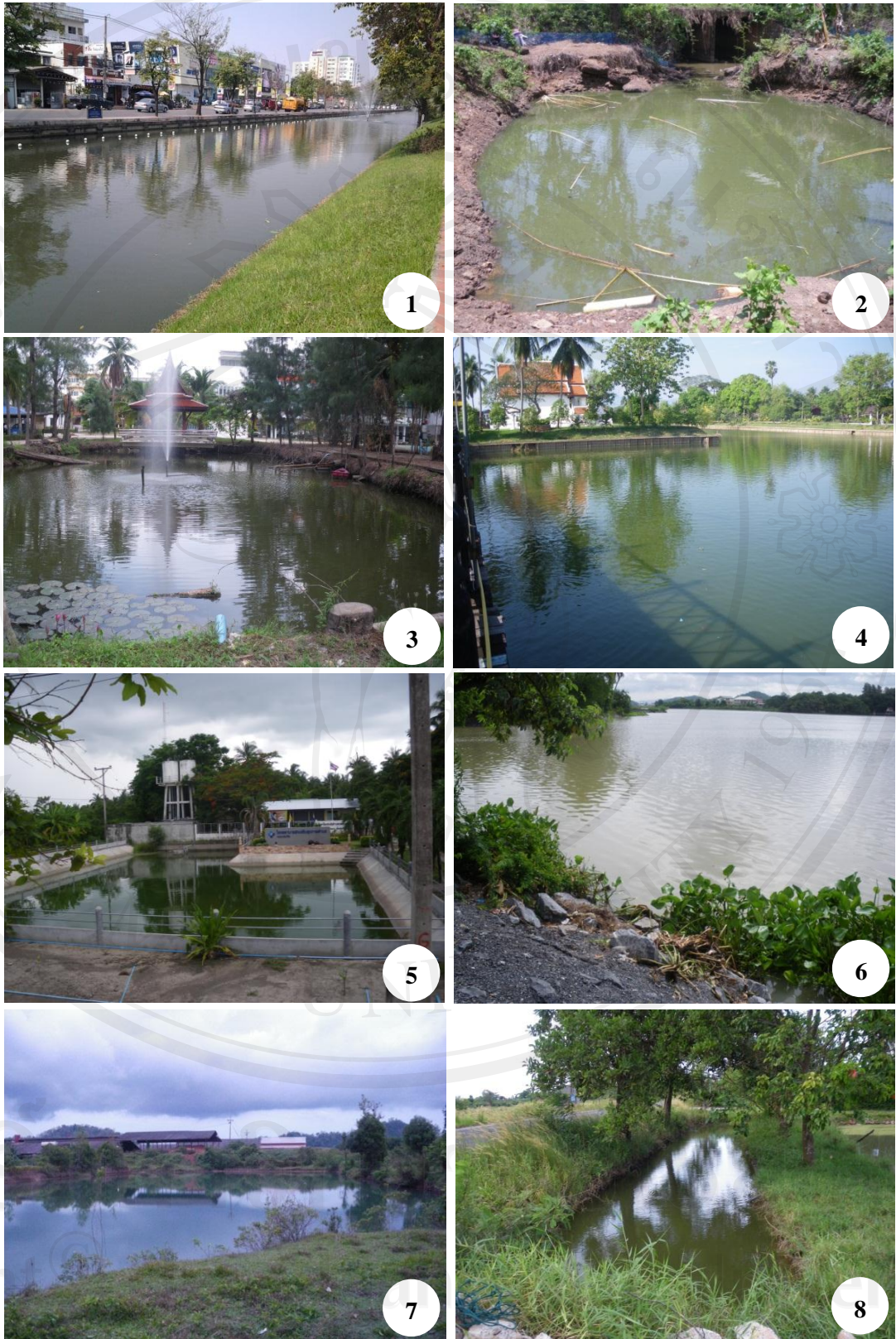


Figure 4 Sampling sites in some freshwater resources of Thailand;

1. Chiang Mai Moat (CHM3)
2. Pond roadside (UTD1)
3. Pond in University of Central Thailand (NKS2)
4. Pond in Wat Traphang Tong (SKT1)
5. Concrete pond in Jompratad Health Promoting Hospital (RBR1)
6. Klong Pure (SRB1)
7. Pond roadside (STN1)
8. Pond roadside (PTL1)

3.2.2 Identification and counting of *Pediastrum* spp.

The *Pediastrum* spp. samples were observed under 40X and 100X light microscope. The specimens were photographed using an Olympus Normaski microscope and reproduced by line drawing. Scanning electron microscope (SEM) study was also carried out. The samples were first rinsed with distilled water, laid on cover glasses and air dried at 30-40 °C and then affixed to aluminium stubs with carbon tape. Finally, the stubs were coated with gold and photographed by SEM according to the method of Wetzel (2001) Kowalska and Wolowski (2010).

Species identification was conducted according to Meneghini (1840), Prescott (1970), Huber-Pestalozzi (1983), Croasdale *et al.* (1994), Chang and Mi (1997), Komarek and Jankovska (2001), John *et al.* (2011) and Kowalska and Wolowski (2010).

The cells were counted by whole count method under light microscope (Rojo *et al.*, 2009) (Appendix A)

3.3 Water sampling procedure

Water samples from each sampling site were collected in polyethylene bottles for nutrient and some physico-chemical analysis. The bottles were kept in a cool box (5-7°C) for analysis in the laboratory. The samples were analyzed as follows:

3.3.1 Physio-chemical parameters at the sampling sites

3.3.1.1 Water and air temperature by a thermometer

3.3.1.2 Light intensity measurement by lux meter (Tecpel 530).

3.3.1.3 Transparency measurement of Secchi depth by Secchi disc

3.3.1.4 Conductivity measurement by a conductivity meter (electrode kit of TRANS Company).

3.3.1.5 pH measurement by a pH meter (electrode kit of TRANS Company)

3.3.1.6 Dissolved oxygen (DO) analysis by azide modification method (Eaton *et al.*, 2005)

3.3.2 Physio-chemical parameters in the laboratory

3.3.2.1 Turbidity analysis by portable data logging spectrophotometer (HACH DR/2010)

3.3.2.2 Alkalinity analysis by the phenolphthalein methyl orange indicator method (Eaton *et al.*, 2005)

3.3.2.3 Biochemical oxygen demand (BOD) by azide modification method (Eaton *et al.*, 2005)

3.3.2.4 Chlorophyll *a* analysis (Saijo, 1975; Winternans and De Mots, 1965)

The nutrient analysis was conducted according to the methods described by Eaton *et al.* (2005)

3.3.2.5 Nitrate nitrogen analysis by cadmium reduction method

3.3.2.6 Ammonium nitrogen analysis by nesslerization method

3.3.2.7 Soluble reactive phosphorus (SRP) analysis by ascorbic acid method

3.4 Cultivation of *Pediastrum* spp.

3.4.1 Isolation

Colonies of *Pediastrum* spp. in the water samples were studied under a microscope and single colony were isolated with a glass micropipette. Each colony was washed at least five times with sterile medium and cultivated in the 12 multi well cell culture plate containing Jaworski's medium (JM). The alga was then purified by streak plate method on JM. Sub-culture was carried out until monoculture was obtained and transferred to broth medium for using as stock culture. The culture was incubated at 25 °C under continuous illumination (Lee *et al.*, 2009).

3.4.2 Cultivation

Pediastrum spp. were cultivated in 3 media: Jaworski's medium, algal broth and bold basal medium for comparison (Appendix D). Then, the algae were cultivated at 4 levels of pH: 6.5, 7.0, 7.5 and 8.0 and finally incubated at 25 °C and room temperature under continuous illumination. The experiment was carried out in 500 ml erlenmeyer flasks containing 300 ml of medium. Cell density was determined spectrophotometrically at a wavelength of 665 nm and cell counts by whole counts method (Rojo *et al.*, 2009). When the growth reached the stationary phase, the cells were harvested by centrifugation and the biomass productivity (P) was calculated as maximum productivity (mg/L/d), according to the equation (Tang *et al.*, 2011).

$$P = 1000 \cdot \frac{(X_1 - X_0)}{(t_1 - t_0)}$$

X₀ is the initial biomass (g/L) at time t₀ (d)

X₁ is the final biomass (g/L) at any time t₁(d)

After the optimal condition for growth was identified, these algae were cultivated and scaled up to 20 L media carrying capacity bottles (Polycarbonate Carboys) for analysis of nutritional value (Lee *et al.*, 2009).

3.5 Analysis of nutritional value

At the end of cultivation, the biomass of the dominant species of *Pediastrum* were selected and collected for nutritional analysis which was conducted according to the methods described by AOAC (1990).

3.5.1 Protein analysis by micro kjeldahl method

3.5.2 Fatty acid analysis by acid hydrolysis method

3.5.3 Fiber analysis by acid detergent method

3.5.4 Ash analysis by burned at 450 °c 1 hour

3.5.5 Carbohydrate analysis by calculating from

$$\text{Carbohydrate (\%)} = 100 - (\text{protein} + \text{fat} + \text{moisture} + \text{ash})$$

3.6 Molecular examination

Genomic DNA was extracted from living cell using the Genomic DNA Extraction Kit (Geneaid) and CTAB method. The 26SrDNA and rbcL rDNA gene were amplified by PCR using specific primer pairs. The PCR conditions for the rbcL and 26S rDNA reactions were 35 cycles at 94 °C, 3 min for predenature, denature at 94 °C for 30 sec, annealing at 55 °C for 30 sec, an extension step at 72 °C for 1 min 30 sec and with a final extension step at 72 °C for 7 min. The QiaQuick Nucleotide Purification Kit (Qiagen Inc.) was used to clean the PCR products, and cycle sequencing reactions of the purified PCR product were done with the ABIPRISM-Big

DyeTM Terminator Cycle Sequencing Reaction Kit (Applied Biosystems, Foster City, CA, USA) and analyzed using DNA sequencing software (McManus and Lewis, 2011).

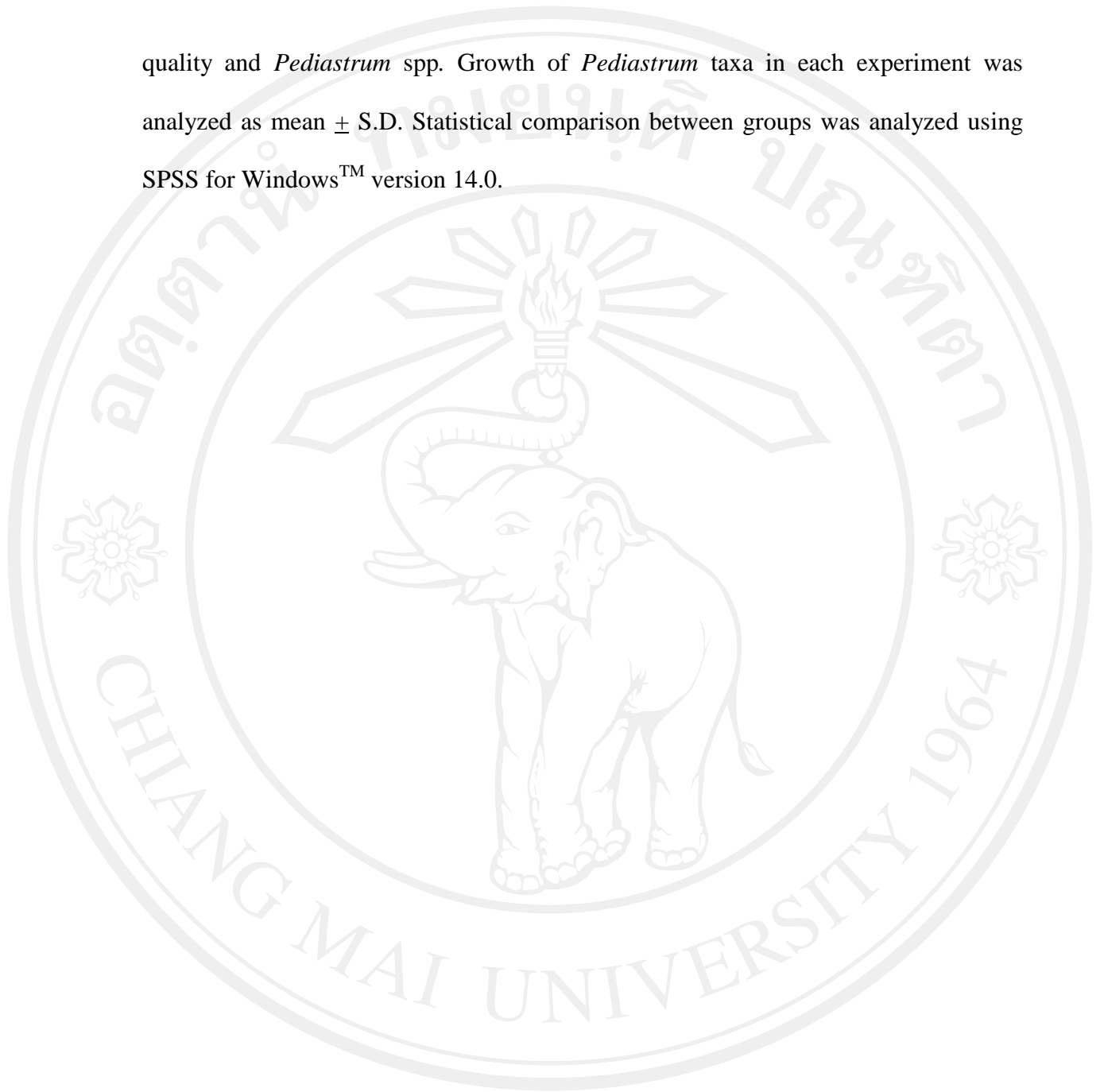
3.7 Phylogenetic tree

The Basic Local Alignment Search Tool (BLAST) at NCBI (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) were used to find sequences similar to those of *Pediastrum* spp. Phylogenetic and molecular evolutionary analyses of the obtained sequences were conducted using the MEGA 5 computer program (Tamura et al. 2011). Alignments were checked manually. A Maximum Likelihood (ML) tree was constructed using software with the best fits model (Tamura three-parameter using a discrete Gamma distribution (_G) with five rate categories and by assuming that a certain fraction of sites is evolutionarily invariable (_I)) based on the lowest Bayesian Information Criterion (BIC) score and the substitution nucleotide matrix parameters calculated by the software. One thousand bootstraps were generated.

3.8 Data evaluation

The trophic status and water quality was classified according to the method of Peerapornpisal *et al.* (2007) based on Wetzel (2001) and Lorraine and Vollenweider (1981) by using selected parameters i.e. dissolved oxygen, biochemical oxygen demand, conductivity, ammonium nitrogen, nitrate nitrogen and soluble reactive phosphorus and chlorophyll *a* (Appendix B and C). The multivariate statistical package (MVSP) for windows was used. Canonical correspondence analysis (CCA) was to find the *Pediastrum* spp. and the relationship between water

quality and *Pediastrum* spp. Growth of *Pediastrum* taxa in each experiment was analyzed as mean \pm S.D. Statistical comparison between groups was analyzed using SPSS for Windows™ version 14.0.



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