



**APPENDICES**

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## APPENDIX A

### Culture Media

#### 1.) Brain Heart Infusion (BHI) Broth

BHI	37.00	g
Distilled water	1000.00	ml

Sterilize by autoclaving at 121°C 15 lbs for 15 min.

#### 2.) BHI agar

BHI agar (dehydrate)	52.00	g
Distilled water	1000.00	ml

Sterilize by autoclaving at 121°C 15 lbs for 15 min. Allow sterilized medium to cool and pour in Petri dishes.

#### 3.) Potato Dextrose Agar (PDA)

PDA agar (dehydrate)	39.00	g
Distilled water	1000.00	ml

Add distilled water, mix thoroughly, heat with frequent agitation and boil for 1 min to completely dissolve the powder. Sterilize by autoclaving at 121°C 15 lbs for 15 min. Allow sterilized medium to cool and pour in Petri dishes.

#### 4.) Malt Extract Agar (MEA)

MEA agar (dehydrate)	50.00	g
Distilled water	1000.00	ml

Add distilled water, mix thoroughly, heat with frequent agitation and boil for 1 min to completely dissolve the powder. Sterilize by autoclaving at 115°C 15 lbs for 10 min. Allow sterilized medium to cool and pour in Petri dishes.

#### 5.) Sabouraud Dextrose Broth (SDB)

SDB (dehydrate)	30.00	g
Distilled water	1000.00	ml

Sterilize by autoclaving at 121°C 15 lbs for 15 min.

### 6.) Luria-Bertani (LB) medium

LB	20.00	g
Deionized water up to	1000.00	ml

Shake until the solutes are dissolved. Sterilize by autoclaving at 121°C 15 lbs for 15 min.

### 7.) Minimal Medium (MM) Agar

NH <sub>4</sub> Cl	2.00	g
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	1.00	g
KCl	0.5	g
NaCl	0.5	g
KH <sub>2</sub> PO <sub>4</sub>	1.00	g
MgSO <sub>4</sub> .7H <sub>2</sub> O	0.5	g
FeSO <sub>4</sub> .7H <sub>2</sub> O	0.02	g
Glucose	10.00	g
Agar	15.00	g

Dissolve in distilled water and adjust pH to 5.0 with 1 N NaOH and adjust the volume to 1000 ml final volume with distilled water. Sterilize by autoclaving at 121°C 15 lbs for 15 min.

### 8.) Aspergillus minimal medium (AMM)

KCl	0.52	g
MgSO <sub>4</sub> .7H <sub>2</sub> O	0.52	g
KH <sub>2</sub> PO <sub>4</sub>	1.52	g
Trace element solution	1	ml

1% D-glucose, 10 mM glutamine

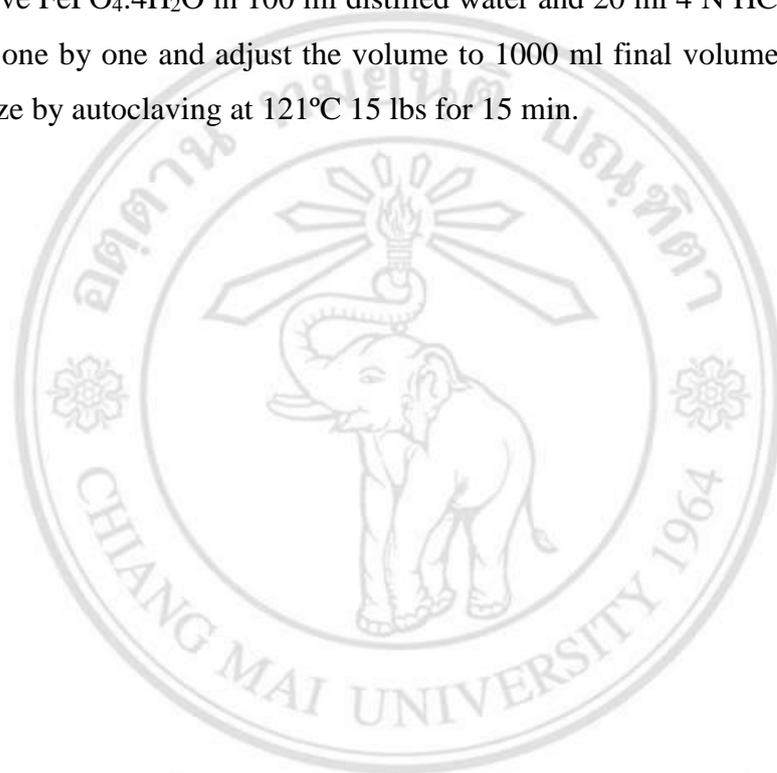
1% Agar

Dissolve in distilled water and adjust pH to 6.5 with 1 N NaOH and adjust the volume to 1000 ml final volume with distilled water. Sterilize by autoclaving at 121°C 15 lbs for 15 min.

### 9.) Trace element solution

$\text{Na}_2\text{B}_4\text{O}_7 \cdot 10\text{H}_2\text{O}$	40.00	mg
$\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$	400.00	mg
$\text{FePO}_4 \cdot 4\text{H}_2\text{O}$	1.00	g
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	600.00	mg
$\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$	800.00	mg
$\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$	8.00	g

Dissolve  $\text{FePO}_4 \cdot 4\text{H}_2\text{O}$  in 100 ml distilled water and 20 ml 4 N HCl, then add the rest elements one by one and adjust the volume to 1000 ml final volume with distilled water. Sterilize by autoclaving at 121°C 15 lbs for 15 min.



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## APPENDIX B

### Alkaline Lysis Buffer for Preparing Plasmid DNA

#### 1.) Solution I

50 mM glucose

25 mM Tris-Cl (pH 8.0)

10 mM EDTA (pH 8.0)

Sterilize by autoclaving for 20 min 15 lb/sq.in. on liquid cycle at store at 4 °C.

#### 2.) Solution II

0.2 N NaOH (freshly dilute from a 10 N stock)

1% SDS

#### 3.) Solution III

5 M Potassium acetate	60.00	ml
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Glacial acetic acid	11.50	ml
---------------------	-------	----

H <sub>2</sub> O	28.50	ml
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The resulting solution is 3 M with the respect to potassium and 5 M with respect to acetate.

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## APPENDIX C

### Solution for Working in Protoplast-mediated Genomic DNA Extraction

#### 1.) Osmotic buffer (pH 8.0)

MgSO <sub>4</sub> .7H <sub>2</sub> O	73.94	g
1 M sodium phosphate buffer pH 7.0	2.50	ml
Distilled water to	250.00	ml

Adjust pH to 5.8. Sterilize by autoclaving at 121°C for 15 min.

#### 2.) ST buffer (pH 5.8)

0.6 M Sorbitol  
100 mM Tri-HCl  
Sterilize by autoclaving for 20 min 15 lb/sq.in. on liquid cycle at store at 4 °C.

#### 3.) Lysis buffer

10 mM Tris  
1 mM EDTA  
1% sodium dodecyl sulphate (SDS)  
Sterilize by autoclaving for 20 min 15 lb/sq.in. on liquid cycle at store at 4 °C.

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## APPENDIX D

### Solution for Protoplast-mediated Transformation

#### 1.) Osmotic buffer (pH 5.8)

MgSO <sub>4</sub> ·7H <sub>2</sub> O	73.94	g
1 M sodium phosphate buffer pH 7.0	2.50	ml
Distilled water to	250.00	ml

Adjust pH to 5.8 with 0.2 M Na<sub>2</sub>HPO<sub>4</sub>. Sterilize by autoclaving at 121°C for 15 min.

#### 2.) Trapping Buffer

D-Sorbitol	54.65	g
1 M Tris-HCl pH 7.0	50.00	ml
Distilled water to	500.00	ml

Sterilize by autoclaving at 121°C for 15 min.

#### 3.) STC (Sorbitol, Tris, CaCl<sub>2</sub>) Buffer

D-Sorbitol	109.30	g
1 M Tris-HCl pH 7.5	5.00	ml
CaCl <sub>2</sub>	0.73	g
Distilled water to	500.00	ml

Sterilize by autoclaving at 121°C for 15 min.

#### 4.) 60% PEG

Polyethylene glycol 4000	150.00	g
1 M CaCl <sub>2</sub>	2.5	ml
1 M Tris-HCl pH 7.5	2.50	ml

Dissolve in distilled water and adjust to 250 ml in volumetric flask. Sterilize by autoclaving at 121°C for 15 min.

## APPENDIX E

### Reagents for General Molecular Biology

#### 1.) 20X SSC

NaCl 175.33 g

Sodium citrate 88.20 g

Dissolve in distilled water and adjust pH to 7.0 with 1 N NaOH and adjust the volume to 1000 ml final volume with distilled water. Sterilize by autoclaving at 121°C for 15 min.

#### 2.) 1 M Tris-HCl pH 7.5

Tris base 121.11 g

Dissolve in distilled water and adjust pH to 7.5 with concentrated HCl and adjust the volume to 1000 ml final volume with distilled water. Sterilize by autoclaving at 121°C for 15 min.

#### 3.) 0.5 M EDTA pH 8.0

EDTA ( $C_{10}H_{14}N_2O_8Na_2 \cdot 2H_2O$ ) 18.6 g

Dissolve in distilled water and adjust pH to 8.0 with NaOH and adjust the volume to 100 ml final volume with distilled water. Sterilize by autoclaving at 121°C for 15 min.

#### 4.) TE Buffer

10 mM Tris-HCl pH 7.5

1 mM EDTA pH 8.0

Sterilize by autoclaving at 121°C for 15 min.

**5.) 50X Tris-acetate (TAE) buffer**

Tris base	242.00	g
Glacial acetic acid	57.10	ml
0.5 M EDTA	100.00	ml

The working solution (1X) contains 0.04 M Tris acetate and 0.0001 M EDTA

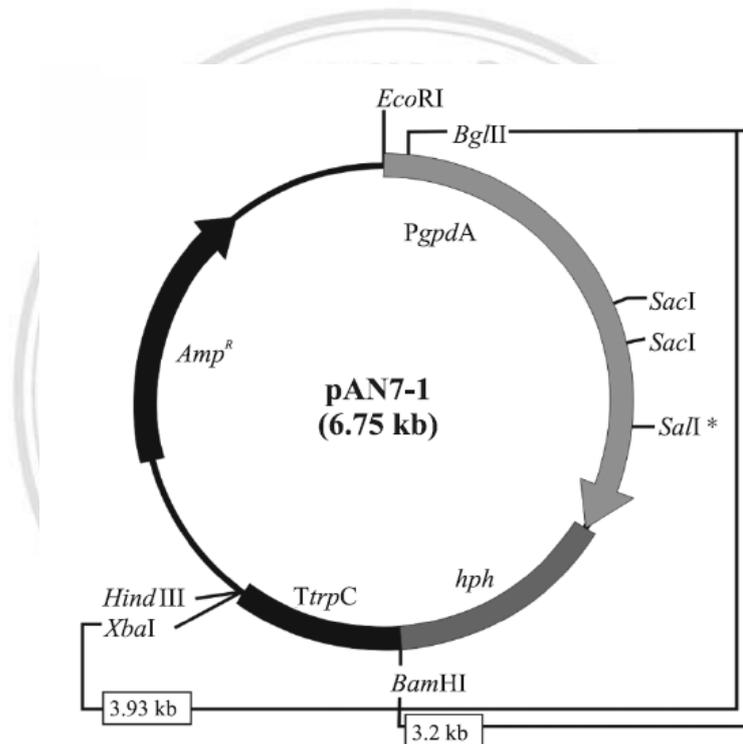


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## Appendix F

### Plasmid Used in This Study

1.) pAN7-1 (Cardoza *et al.*, 2006)



**Expression vector (PAN7-1) DNA, 6756bp**

**GenBank: Z32698.1** (<http://www.ncbi.nlm.nih.gov/nucore/Z32698.1>)

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 ACCESSION Z32698  
 VERSION Z32698.1 GI:475166  
 KEYWORDS hygromycin b phosphotransferase.  
 SOURCE synthetic construct  
 ORGANISM synthetic construct  
 other sequences; artificial sequences.  
 REFERENCE 1  
 AUTHORS Mullaney,E.J., Hamer,J.E., Roberti,K.A., Yelton,M.M. and Timberlake,W.E.  
 TITLE Primary structure of the trpC gene from Aspergillus nidulans

JOURNAL Mol. Gen. Genet. 199 (1), 37-45 (1985)  
 PUBMED [3158796](#)  
 REMARK sites .  
 REFERENCE 2  
 AUTHORS Punt,P.J., Oliver,R.P., Dingemanse,M.A., Pouwels,P.H. and van den Hondel,C.A.  
 TITLE Transformation of Aspergillus based on the hygromycin B resistance marker from Escherichia coli  
 JOURNAL Gene 56 (1), 117-124 (1987)  
 PUBMED [2824287](#)  
 REMARK (sites)  
 REFERENCE 3 (bases 999 to 2572)  
 AUTHORS Punt,P.J., Dingemanse,M.A., Kuyvenhoven,A., Soede,R.D., Pouwels,P.H. and van den Hondel,C.A.  
 TITLE Functional elements in the promoter region of the Aspergillus nidulans gpdA gene encoding glyceraldehyde-3-phosphate dehydrogenase  
 JOURNAL Gene 93 (1), 101-109 (1990)  
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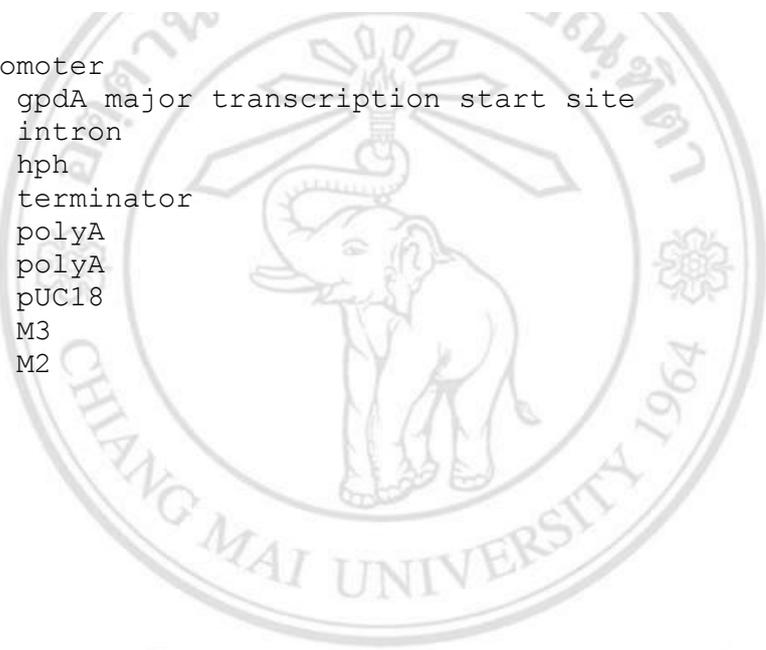




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2302-3321 = hph  
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3590-3600 = polyA  
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4105-6756 = pUC18  
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6711-6735 = M2



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List of single cutter restriction enzymes in pAN7-1

0 cutter

#	Enzyme	Specificity
1	Acc65I	G GTAC△C
2	AfeI	AGC△GCT
3	AflII	C TTAA△G
4	AleI	CACNN△NNGTG
5	ApaI	G△GGCC C
6	AscI	GG△CGCG△CC
7	BsaAI	YAC△GTR
8	BsaBI	GATNN△NNATC
9	BsiWI	C GTAC△G
10	BspDI	AT CG△AT
11	BssHII	G△CGCG△C
12	BstXI	CCAN△NNNN NTGG
13	Bsu36I	CC TNA△GG
14	ClaI	AT△CG△AT
15	CspCI	△NN (N) <sub>11</sub> CAA(N) <sub>5</sub> GTGG(N) <sub>10</sub> △NN
16	EcoNI	CCTNN N△NNAGG
17	EcoRV	GAT△ATC
18	FseI	GG△CCGG CC
19	HpaI	GTT△AAC
20	KpnI	G△GTAC C
21	NotI	GC△GGCC△GC
22	NruI	TCG△CGA
23	PacI	TTA△AT TAA
24	PmeI	GTTT△AAAC
25	PmlI	CAC△GTG
26	PsiI	TTA△TAA
27	PspOMI	G△GGCC△C
28	SexAI	A△CCWGG△T
29	SfiI	GGCCN△NNN NGGCC
30	SgrAI	CR△CCGG△YG
31	SmaI	CCC△GGG
32	SnaBI	TAC△GTA
33	SpeI	A△CTAG△T
34	SwaI	ATTT△AAAT
35	TspMI	C△CCGG△G
36	XmaI	C△CCGG△G

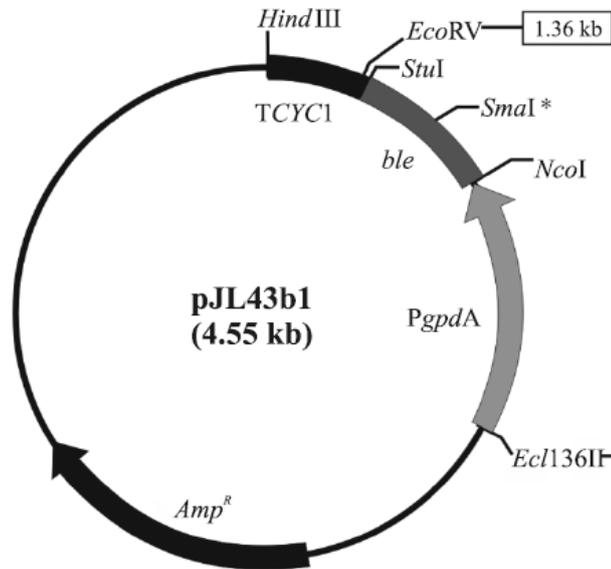
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#	Enzyme	Specificity	Sites & flanks	Cut positions (blunt - 5' ext. - 3' ext.)
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2	AsiSI	GCG↓AT CGC	<a href="#">list</a>	*2660/2658
3	AvrII	C CTAG↓G	<a href="#">list</a>	1718/1722
4	BamHI	G GATC↓C	<a href="#">list</a>	3339/3343
5	BbvCI	CC TCA↓GC	<a href="#">list</a>	1976/1979
6	BclI	T GATC↓A	<a href="#">list</a>	#853/857
7	BglII	A GATC↓T	<a href="#">list</a>	138/142
8	BlpI	GC TNA↓GC	<a href="#">list</a>	3631/3634
9	BmgBI	CAC↓GTC	<a href="#">list</a>	*374
10	BmtI	G↓CTAG C	<a href="#">list</a>	908/904
11	BsrGI	T GTAC↓A	<a href="#">list</a>	1422/1426
12	BstEII	G GTNAC↓C	<a href="#">list</a>	396/401
13	BstZ17I	GTA↓TAC	<a href="#">list</a>	3921
14	HindIII	A AGCT↓T	<a href="#">list</a>	4122/4126
15	KasI	G GCGC↓C	<a href="#">list</a>	*4286/4290
16	MfeI	C AATT↓G	<a href="#">list</a>	1734/1738
17	MluI	A CGCG↓T	<a href="#">list</a>	*3525/3529
18	NaeI	GCC↓GGC	<a href="#">list</a>	*3977
19	NarI	GG CG↓CC	<a href="#">list</a>	*4287/4289
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21	NgoMIV	G CCGG↓C	<a href="#">list</a>	*3975/3979
22	NheI	G CTAG↓C	<a href="#">list</a>	904/908
23	PciI	A CATG↓T	<a href="#">list</a>	6401/6405
24	PluTI	G↓GCGC C	<a href="#">list</a>	*4290/4286
25	PshAI	GACNN↓NNGTC	<a href="#">list</a>	*2323
26	PspXI	VC TCGA↓GB	<a href="#">list</a>	*1076/1080
27	RsrII	CG GWC↓CG	<a href="#">list</a>	*2704/2707
28	SalI	G TCGA↓C	<a href="#">list</a>	*1944/1948
29	SbfI	CC↓TGCA GG	<a href="#">list</a>	4114/4110
30	SfoI	GGC↓GCC	<a href="#">list</a>	*4288
31	StuI	AGG↓CCT	<a href="#">list</a>	465
32	XbaI	T CTAG↓A	<a href="#">list</a>	4058/4062
33	XcmI	CCANNNN↓N NNNNTGG	<a href="#">list</a>	1163/1162

2 cutters

#	Enzyme	Specificity	Sites & flanks	Cut positions (blunt - 5' ext. - 3' ext.)
1	AatII	G $\downarrow$ ACGT C	<u>list</u>	*2323/2319, *4594/4590
2	AccI	GT MK $\downarrow$ AC	<u>list</u>	*1945/1947, 3920/3922
3	AflIII	A $\downarrow$ CRYG $\downarrow$ T	<u>list</u>	3525/3529, 6401/6405
4	AgeI	A $\downarrow$ CCGG $\downarrow$ T	<u>list</u>	*1283/1287, *1431/1435
5	AvaI	C YCGR $\downarrow$ G	<u>list</u>	*694/698, *1076/1080
6	BanII	G $\downarrow$ RGCY C	<u>list</u>	1257/1253, 1420/1416
7	BbsI	GAAGACNN NNN $\downarrow$	<u>list</u>	368/372, 1056/1060
8	BciVI	GTATCC (N) <sub>5</sub> $\downarrow$ N	<u>list</u>	4676/4675, 6203/6202
9	BmrI	ACTGGGNNNN $\downarrow$ N	<u>list</u>	4167/4166, 5473/5472
10	Bpu10I	CC $\downarrow$ TNA $\downarrow$ GC	<u>list</u>	1900/1903, 1976/1979
11	BseYI	C $\downarrow$ CCAG $\downarrow$ C	<u>list</u>	3292/3296, 6097/6101
12	BsgI	GTGCAG (N) <sub>14</sub> $\downarrow$ NN	<u>list</u>	1547/1545, 1947/1945
13	BsmI	GAATG $\downarrow$ CN	<u>list</u>	3958/3956, 4026/4024
14	BsoBI	C YCGR $\downarrow$ G	<u>list</u>	694/698, 1076/1080
15	BspQI	GCTCTTCN NNN $\downarrow$	<u>list</u>	323/326, 6518/6521
16	BstBI	TT CG $\downarrow$ AA	<u>list</u>	*354/356, *786/788
17	BtgZI	GCGATG (N) <sub>10</sub> NNN $\downarrow$	<u>list</u>	*2242/2246, *2968/2972
18	DraIII	CAC $\downarrow$ NNN GTG	<u>list</u>	2584/2581, *2877/2874
19	Eco53kI	GAG $\downarrow$ CTC	<u>list</u>	1255, 1418
20	EcoRI	G $\downarrow$ AATT $\downarrow$ C	<u>list</u>	1/5, 2539/2543
21	FspI	TGC $\downarrow$ GCA	<u>list</u>	*4267, *5290
22	MscI	TGG $\downarrow$ CCA	<u>list</u>	1501, 1911
23	NdeI	CA TA $\downarrow$ TG	<u>list</u>	2746/2748, 4339/4341
24	Paer7I	C $\downarrow$ TCGA $\downarrow$ G	<u>list</u>	*694/698, *1076/1080
25	PpuMI	RG GWC $\downarrow$ CY	<u>list</u>	664/667, 1980/1983
26	SacI	G $\downarrow$ AGCT C	<u>list</u>	1257/1253, 1420/1416
27	SacII	CC $\downarrow$ GC GG	<u>list</u>	*3076/3074, *3337/3335
28	SapI	GCTCTTCN NNN $\downarrow$	<u>list</u>	323/326, 6518/6521
29	SspI	AAT $\downarrow$ ATT	<u>list</u>	681, 4708
30	StyI	C CWWG $\downarrow$ G	<u>list</u>	1718/1722, 2648/2652
31	XhoI	C TCGA $\downarrow$ G	<u>list</u>	*694/698, *1076/1080
32	XmnI	GAANN $\downarrow$ NNTTC	<u>list</u>	707, 4913
33	ZraI	GAC $\downarrow$ GTC	<u>list</u>	*2321, *4592

## 2.) pJL43b1



CTAAATTGTAAGCGTTAATATTTTGTAAAATTCGCGTTAAATTTTGTAAAATCAGCTCATTTTTTAAAC  
 CAATAGGCCGAAATCGGCAAAATCCCTTATAAATCAAAGAATAGACCGAGATAGGGTTGAGTGTGTTC  
 CAGTTTGAACAAGAGTCCACTATTAAGAACGTGGACTCCAACGTCAAAGGGCGAAAAACCGTCTATCA  
 GGGCGATGGCCACTACGTGAACCATCACCTAATCAAGTTTTTGGGGTTCGAGGTGCCGTAAAGCACTA  
 AATCGGAACCCATAAGGGAGCCCCGATTTAGAGCTTGACGGGAAAGCCGGCGAACGTGGCGAGAAAGG  
 AAGGAAGAAAGCGAAAGGAGCGGGCGCTAGGGCGCTGGCAAGTGTAGCGGTCACGCTGCGCGTAACCC  
 CACACCCGCGCGCTTAATGCGCCGCTACAGGGCGCGTCCATTTCGCCATTCAGGCTGCGCAACTGTTGG  
 GAAGGGCGATCGGTGCGGGCCTCTTCGCTATTACGCCAGCTGGCGAAAGGGGGATGTGCTGCAAGGCGAT  
 TAAGTTGGGTAACGCCAGGGTTTTCCAGTCACGACGTTGTAAAACGACGGCCAGTGAGCGCGCGTAATA  
 CGACTCACTATAGGGCGAATTTGGAGCTCTGTACAGTGACCGGTGACTCTTTCTGGCATGCGGAGAGACGG  
 ACGGACGCAGAGAGAAGGGCTGAGTAATAAGCCACTGGCCAGACAGCTCTGGCGGCTCTGAGGTGCAGTG  
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 TCCCTAATTGGCCCATCCGGCATCTGTAGGGCGTCAAATATCGTGCCTCTCCTGCTTTGCCCGGTGTAT  
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 CCGGTGCTCTGCACCTCGACCTGCTGAGGTCCCTCAGTCCCTGGTAGGCAGCTTTGCCCGTCTGTCCGCC  
 CGGTGTGTGGCGGGGTTGACAAGGTGCTTGCGTCACTCAACATTTGTTGCCATATTTTCTGCTCTCC  
 CCACCAGCTGCTCTTTTCTTTTCTTTTCTTTTCCCATCTTCAGTATATTCATCTTCCCATCCAAGAACC  
 TTTATTTCCCTAAGTAAGTACTTTGCTACATCCATACTCCATCCTTCCCATCCCTTATTCCTTTGAACC  
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 GGCCAAGTTGACCAAGTCCGCTTCCGGTGTCCAGCGCGCAGCTCGCCGGAGCGGTCGAGTTCTGGACC  
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 TGTTCATCAGCGCGGTCCAGGACCAGGTGGTCCCGACAACACCCTGGCCTGGGTGTGGGTGCGCGGCCT  
 GGACGAGCTGTACGCCGAGTGGTCCGAGGTGCTGTCCACGAACCTCCGGGACGCTCCGGGCGGCCATG  
 ACCGAGATCGGCGAGCAGCCGTGGGGGCGGGAGTTCGCCCTGCGCGACCCGGCCGCAACTGCGTGCACCT  
 TCGTGGCCGAGGAGCAGGACTGACCGACGCCACCAACACCAGCGGTCCGACGCGGCCCGACGGGTCCGA  
 GGCTCCGGAGATCCGTCCCCCTTTTCTTTTGTGCGATATCATGTAATTAGTTATGTCACGCTTACATTCAC  
 GCCCTCCCCCACATCCGCTCTAACCAGAAAGGAAGGAGTTAGACAACCTGAAGTCTAGGTCCCTATTTA  
 TTTTTTTATAGTTATGTTAGTATTAAGAACGTTATTTATATTTCAAATTTTTCTTTTTTTCTGTACAGA  
 CGCGTGTACGCATGTAACATTATACTGAAAACCTTGCTTGAGAAGGTTTTGGGACGCTCGAAGGCTTTAA  
 TTTGCAAGCTTATCGATACCGTGCACCTCGAGGGGGGGCCCGGTACCCAGCTTTTGTTCCTTTAGTGAG  
 GGTTAATTGCGCGCTTGGCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTATCCGCTCACAAT

TCCACACAACATACGAGCCGGAAGCATAAAGTGTAAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCACA  
TTAATTGCGTTGCGCTCACTGCCCGCTTTCCAGTCGGGAAACCTGTGCTGCCAGCTGCATTAATGAATCG  
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TGCTGGCGTTTTTCCATAGGCTCCGCCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTG  
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CCGACCTGCGCTTACCGGATACCTGTCCGCCTTTCTCCCTTCGGAAGCGTGGCGCTTTTCTCATAGCT  
CACGCTAGGTATCTCAGTTCGGTGTAGGTCTGCTTCCAGCTGGGCTGTGTGCACGAACCCCCCGT  
TCAGCCCCGACCGCTGCGCCTTATCCGGTAACATCGTCTTTCAGTCCAACCCGGTAAGACACGACTTATCG  
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CTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAAACAAACCACCGCTGGTAGCGGTGGTTTTTTTTGTT  
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ACGCTCAGTGAACGAAAACCTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTACCTA  
GATCCTTTTAAATTAATAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAAACTTGGTCTGACAGT  
TACCAATGCTTAATCAGTGAGGCACCTATCTCAGCGATCTGTCTATTTTCGTTTCATCCATAGTTGCCTGAC  
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GGTCTGCAACTTTATCCGCTCCATCCAGTCTATTAATTGTTGCCGGAAGCTAGAGTAAGTAGTTCCG  
CAGTTAATAGTTTGGCAACGTTGTTGCCATTGCTACAGGCATCGTGGTGTACGCTCGTCTGTTTGGTAT  
GGCTTCATTCAGCTCCGGTTCCCAACGATCAAGGCGAGTTACATGATCCCCATGTTGTGCAAAAAAGCG  
GTTAGCTCCTTCGGTCCCTCCGATCGTTGTGCAAGTAAGTTGGCCGAGTGTATCACTCATGGTTATGG  
CAGCACTGCATAATCTCTTACTGTGATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGGTACTCAAC  
CAAGTCATTCGAGAATAGTGTATGCGGCGACCGAGTTGCTCTTGGCCGGCGTCAATACGGGATAATACC  
GCGCCACAGCAGAGAACTTAAAAGTGCTCATCTGGAAAACGTTCTTTCGGGGCGAAAACCTCAAGGA  
TCTTACCAGTGTGAGATCCAGTTCGATGTAACCCACTCGTGCACCCAACTGATCTCAGCATCTTTTAC  
TTTTACCAGCGTTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAAGGGAATAAGGGGACA  
CGGAAATGTTGAATACTCATACTCTTCTTTTTCAATATTATTGAAGCATTATCAGGGTTATTGTCTCA  
TGAGCGGATACATATTTGAATGTATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACATTTCCCCGAAA  
AGTGCCAC

653-1538 (in red) gpdA promoter (886 bp)  
1539-1965 (in blue) phleomycin gene (427 bp)  
1966-2251 (in green) terminator region of CYC1 gene

List of single cutter restriction enzymes in pJL43b1

#	Enzyme	Specificity	Sites & flanks	Cut positions (blunt - 5' ext. - 3' ext.)
1	AatII	G $\downarrow$ ACGT C	<u>list</u>	*1585/1581
2	Acc65I	G GTAC $\downarrow$ C	<u>list</u>	*2282/2286
3	AgeI	A CCGG $\downarrow$ T	<u>list</u>	*668/672
4	AhdI	GACNN $\downarrow$ N NNGTC	<u>list</u>	3573/3572
5	ApaI	G $\downarrow$ GGCC C	<u>list</u>	*2280/2276
6	AvrII	C CTAG $\downarrow$ G	<u>list</u>	955/959
7	BaeI	$\uparrow$ (N) $\downarrow$ <sub>5</sub> (N) <sub>10</sub> ACN $\downarrow$ NNNGTAYC(N) $\uparrow$ <sub>7</sub> (N) $\downarrow$ <sub>5</sub>	<u>list</u>	951/946+984/979
8	BbvCI	CC TCA $\downarrow$ GC	<u>list</u>	1213/1216

9	BfuAI	ACCTGCNNNN NNNN▲	<a href="#">list</a>	1217/1221
10	BmgBI	CAC▲GTC	<a href="#">list</a>	*1673
11	BpmI	CTGGAG (N) <sub>14</sub> ▲NN	<a href="#">list</a>	3643/3641
12	BsaAI	YAC▲GTR	<a href="#">list</a>	*227
13	BsaI	GGTCTCN NNNN▲	<a href="#">list</a>	3634/3638
14	BseRI	GAGGAG (N) <sub>8</sub> ▲NN	<a href="#">list</a>	1914/1912
15	BsmBI	CGTCTCN NNNN▲	<a href="#">list</a>	*688/692
16	BspDI	AT CG▲AT	<a href="#">list</a>	*2253/2255
17	BspMI	ACCTGCNNNN NNNN▲	<a href="#">list</a>	1217/1221
18	BspQI	GCTCTCN NNN▲	<a href="#">list</a>	2564/2567
19	ClaI	AT CG▲AT	<a href="#">list</a>	*2253/2255
20	Eco53kI	GAG▲CTC	<a href="#">list</a>	655
21	EcoRV	GAT▲ATC	<a href="#">list</a>	1996
22	FseI	GG▲CCGG CC	<a href="#">list</a>	*1815/1811
23	HindIII	A AGCT▲T	<a href="#">list</a>	2246/2250
24	KpnI	G▲GTAC C	<a href="#">list</a>	2286/2282
25	MfeI	C AATT▲G	<a href="#">list</a>	971/975
26	MluI	A CGCG▲T	<a href="#">list</a>	*2170/2174
27	NcoI	C CATG▲G	<a href="#">list</a>	1537/1541
28	PaeR7I	C TCGA▲G	<a href="#">list</a>	*2267/2271
29	PciI	A CATG▲T	<a href="#">list</a>	2680/2684
30	PflFI	GACN N▲NGTC	<a href="#">list</a>	1282/1283
31	PsiI	TTA▲TAA	<a href="#">list</a>	99
32	PspOMI	G GGCC▲C	<a href="#">list</a>	*2276/2280
33	PspXI	VC TCGA▲GB	<a href="#">list</a>	*2267/2271
34	RsrII	CG GWC▲CG	<a href="#">list</a>	*1935/1938
35	SacI	G▲AGCT C	<a href="#">list</a>	657/653
36	SapI	GCTCTCN NNN▲	<a href="#">list</a>	2564/2567
37	SexAI	A CCWGG▲T	<a href="#">list</a>	#1702/1707
38	SgrAI	CR CCGG▲YG	<a href="#">list</a>	*1653/1657
39	SmaI	CCC▲GGG	<a href="#">list</a>	*1629
40	StuI	AGG▲CCT	<a href="#">list</a>	1962
41	TliI	C TCGA▲G	<a href="#">list</a>	*2267/2271
42	TspMI	C CCGG▲G	<a href="#">list</a>	*1627/1631
43	Tth111I	GACN N▲NGTC	<a href="#">list</a>	1282/1283
44	XhoI	C TCGA▲G	<a href="#">list</a>	*2267/2271
45	XmaI	C CCGG▲G	<a href="#">list</a>	*1627/1631
46	XmnI	GAANN▲NNTTC	<a href="#">list</a>	4172
47	ZraI	GAC▲GTC	<a href="#">list</a>	*1583

## CURRICULUM VITAE

- Name** Ms. Panjaphorn Nimmanee
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- 2001 Bachelor Degree of Science (Medical Technology), Faculty of Allied Health Science, Thammasart University, Thailand
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